

**Manuscript version: Author's Accepted Manuscript**

The version presented in WRAP is the author's accepted manuscript and may differ from the published version or Version of Record.

**Persistent WRAP URL:**

<http://wrap.warwick.ac.uk/108652>

**How to cite:**

Please refer to published version for the most recent bibliographic citation information. If a published version is known of, the repository item page linked to above, will contain details on accessing it.

**Copyright and reuse:**

The Warwick Research Archive Portal (WRAP) makes this work by researchers of the University of Warwick available open access under the following conditions.

Copyright © and all moral rights to the version of the paper presented here belong to the individual author(s) and/or other copyright owners. To the extent reasonable and practicable the material made available in WRAP has been checked for eligibility before being made available.

Copies of full items can be used for personal research or study, educational, or not-for-profit purposes without prior permission or charge. Provided that the authors, title and full bibliographic details are credited, a hyperlink and/or URL is given for the original metadata page and the content is not changed in any way.

**Publisher's statement:**

Please refer to the repository item page, publisher's statement section, for further information.

For more information, please contact the WRAP Team at: [wrap@warwick.ac.uk](mailto:wrap@warwick.ac.uk).

**Metallome of cerebrovascular endothelial cells infected with *Toxoplasma gondii* using  $\mu$ -XRF imaging and inductively coupled plasma mass spectrometry**

Alaa T. Al-sandaqchi <sup>a,†</sup>, Chris Brignell <sup>b</sup>, Joanna F. Collingwood <sup>c</sup>, Kalotina Geraki <sup>d</sup>, Evgeny M. Mirkes <sup>e</sup>, Kenny Kong <sup>f</sup>, Marcos Castellanos <sup>g</sup>, Sean T. May <sup>g</sup>, Carl W. Stevenson <sup>a</sup>, Hany M. Elsheikha <sup>h\*</sup>

<sup>a</sup> *School of Biosciences, University of Nottingham, Sutton Bonington Campus, Loughborough, LE12 5RD, UK*

<sup>b</sup> *School of Mathematical Sciences, University of Nottingham, Nottingham, NG7 2RD, United Kingdom*

<sup>c</sup> *School of Engineering, University of Warwick, Coventry, CV4 7AL, UK*

<sup>d</sup> *Diamond Light Source, Harwell Science and Innovation Campus, Didcot, OX11 0DE, UK*

<sup>e</sup> *Mathematics Department, University of Leicester, University Road, Leicester, LE1 7RH, UK*

<sup>f</sup> *School of Physics and Astronomy, University of Nottingham, University Park, Nottingham NG7 2RD, UK*

<sup>g</sup> Nottingham Arabidopsis Stock Centre, School of Biosciences, University of Nottingham,  
Loughborough LE12 5RD, United Kingdom

<sup>h</sup> Faculty of Medicine and Health Sciences, School of Veterinary Medicine and Science,  
University of Nottingham, Sutton Bonington Campus, Leicestershire, LE12 5RD, UK

\* Corresponding author

Hany Elsheikha

Email address: [hany.elsheikha@nottingham.ac.uk](mailto:hany.elsheikha@nottingham.ac.uk)

Current address:

† Department of Veterinary Public Health, College of Veterinary Medicine and Science,  
Basra University, Iraq

**Table of contents entry (up to 30 words)**

Elemental profiling using synchrotron  $\mu$ -XRF imaging coupled with ICP-Mass Spectrometry and microarray expression of metallothionein genes identified elements significantly associated with *Toxoplasma gondii* infection of cerebrovascular endothelial cells.

## Abstract

In this study, we measured the levels of elements in human brain microvascular endothelial cells (ECs) infected with *T. gondii*. ECs were infected with tachyzoites of the RH strain, and at 6, 24, and 48 hours post infection (hpi), the intracellular concentrations of elements were determined using a synchrotron-microfocus X-ray fluorescence microscopy ( $\mu$ -XRF) system. This method enabled the quantification of the concentrations of Zn and Ca in infected and uninfected (control) ECs at the sub-micron spatial resolution. *T. gondii*-hosting ECs contained less Zn than uninfected cells only at 48 hpi ( $p < 0.01$ ). The level of Ca was not significantly different between infected and control cells ( $p > 0.05$ ). Inductively Coupled Plasma Mass Spectrometry (ICP-MS) analysis revealed infection-specific metallome profiles characterized by significant increases in the intracellular levels of Zn, Fe, Mn and Cu at 48 hpi ( $p < 0.01$ ), and significant reductions in the extracellular concentrations of Co, Cu, Mo, V, and Ag at 24 hpi ( $p < 0.05$ ) compared with control cells. Zn constituted the largest part (74%) of the total metal composition (metallome) of the parasite. Gene expression analysis showed infection-specific upregulation in the expression of five genes, MT1JP, MT1M, MT1E, MT1F, and MT1X, belonging to the metallothionein gene family. These results point to a possible correlation between *T. gondii* infection and increased expression of MT1 isoforms and altered intracellular levels of elements, especially Zn and Fe. Taken together, a combined  $\mu$ -XRF and ICP-MS approach is promising for studies of the role of elements in mediating host-parasite interaction.

**Keywords:** *Toxoplasma gondii*, blood-brain barrier, host-pathogen interaction, label-free imaging, X-ray fluorescence spectromicroscopy; Mass Spectrometry

## 1. Introduction

*Toxoplasma gondii* is an obligate intracellular apicomplexan pathogen causing diseases by reiterating its lytic cycle, comprising host cell invasion, parasite replication and parasite egress. The ability of this parasite to invade host cells, disseminate through tissues and cause disease depends critically on its ability to locate and engage with the target host central nervous system (CNS). The blood-brain barrier (BBB) plays an important role as the biological interface that separates the host neural tissues from circulating blood. Human brain microvascular endothelial cells (ECs) constitute the fundamental component of the BBB, and together with basal lamina, astrocytic foot processes, pericytes, and tight junctions strictly regulate substance entry to neuronal tissue.<sup>1-3</sup> Overcoming this protective barrier is a fundamental step in the establishment of brain infection.<sup>4</sup>

*T. gondii* invades and survives in host cells by modulating host cell processes and evading innate defenses, but the mechanisms are not fully defined. Metal dyshomeostasis during *T. gondii* infection implicates a significant role for metals in mediating parasite-host interaction. Fe, Cu and Zn are essential elements required for a multitude of cellular functions, such as enzymatic reactions, DNA synthesis, metabolic processes, and gene expression.<sup>5</sup> These metals play key roles in the host response to infection, including the development of the immune response and influencing the virulence of microorganisms.<sup>6-8</sup> The intracellular homeostasis of these elements is tightly regulated, and alteration of their levels can have adverse impacts on the host cell and its ability to respond to microbial infection. *T. gondii* infection has been shown to alter the levels of Zn, Fe, Mg and Cu in the blood of seropositive sheep<sup>9</sup> and humans,<sup>10</sup> compared to their seronegative counterparts. Therefore, it is reasonable to hypothesize that host cell elemental content will change in response to *T. gondii* infection, especially in tissues with high metabolic demand such as the CNS.

This study establishes a novel use of synchrotron-microfocus X-ray fluorescence microscopy ( $\mu$ -XRF) to determine changes in the cellular levels of Fe, Cu, Zn, and other elements in ECs challenged with *T. gondii*.  $\mu$ -XRF is a chemical, non-invasive, element imaging technique, which can be used to generate x-ray fluorescent 2D elemental maps of biological samples,<sup>11-12</sup> with detection sensitivity and spatial resolution well-suited to characterize host-parasite interaction.  $\mu$ -XRF imaging can enable *in situ* interrogation of the spatial distribution of one or more elements to submicron spot sizes and offers enough sensitivity and precision to detect metal distribution even at the single cell and subcellular levels.<sup>13</sup> However, absolute quantification of elements is not so straightforward using  $\mu$ -XRF imaging, and in this respect the method is usually considered semi-quantitative. We therefore compared these results with those obtained from inductively coupled plasma mass spectrometry (ICP-MS), which is currently a gold-standard technique for the determination of absolute trace element concentrations and has been widely used for the elemental analysis of various tissues.<sup>14</sup>

## **2. Experimental**

### **2.1. Parasite strain**

*Toxoplasma gondii* genotype I (RH strain) tachyzoites were maintained by passage in Madin-Darby Canine Kidney (MDCK) cell cultures grown in complete Dulbecco's modified Eagle's medium (DMEM), supplemented with 10% heat-inactivated fetal bovine serum (FBS), 2 mM glutamine, and 1% antibiotic-antimycotic solution at 37°C under a humidified atmosphere of 5% CO<sub>2</sub>. The tachyzoites were purified from their feeder MDCK cell cultures by passage through PD-10 desalting columns filled with Sephadex, as described previously.<sup>15</sup> The purified parasites were centrifuged at 800×g, re-suspended in fresh medium, and quantified

using a hemocytometer. The final volume of pure tachyzoite suspension was adjusted with fresh Roswell Park Memorial Institute (RPMI) 1640 medium.

## **2.2. Cell culture**

Human brain microvascular endothelial cells (ECs) were used at passage 9 and were maintained *in vitro*, as described previously.<sup>16, 17</sup> Briefly, ECs were propagated in tissue-culture medium composed of RPMI 1640 medium supplemented with 20% (v/v) heat inactivated FBS, 2mM L-glutamine, 1mM sodium pyruvate, 1% non-essential amino acids, 1% MEM vitamins, and 2% penicillin/streptomycin (Invitrogen, GIBCO, UK). Cells were maintained in an incubator in a humidified atmosphere at 37°C and 5% CO<sub>2</sub>. Once confluent (~3 days), cells were trypsinized using trypsin-EDTA (Invitrogen, GIBCO, UK). Cells were examined daily under light microscopy and were considered confluent when their expansion had reached a point where cells touched each other on all sides, leaving no intercellular gaps. To exclude the possibility that cell viability has influenced elemental concentration of the ECs or parasite interactions with the ECs and any subsequent measurement, the viability of the cells was assessed for a minimum of 100 cells using 0.15% trypan blue exclusion assay prior to use in any experiment.

## **2.3. Quantification of elements using ICP-MS**

ECs were seeded in T-175 cm<sup>2</sup> tissue culture flasks at 10<sup>10</sup> cells/flask and grown in RPMI medium, as described above. Once a confluent cell monolayer was formed (~24 h), tachyzoites were added at a multiplicity of infection (MOI) of 2 (i.e., a host:parasite ratio of 1:2). The culture medium (500 µl) was collected 6, 24 and 48 hours post-infection (hpi) from the infected and non-infected (control) cultures and diluted to 10 ml (1:20 dilution) with 1% nitric acid, before running on the inductively coupled plasma mass spectrometry (ICP-MS;

Varian Ultramass, Melbourne, VIC, Australia) system fitted with a direct injection nebulizer (CETAC, Omaha, NE, USA) using Rh (10 ng/l) as an internal standard in order to quantify the concentrations of the extracellular trace elements. At identical time points after infection (**Supplementary file; Figure S1**), infected and uninfected cells were harvested using a sterile cell scraper, followed by washing three times with deionized water. The cells (infected or uninfected) and purified tachyzoites were centrifuged for 5 min at 800xg. The cell or tachyzoite pellets were lyophilized by a Modulyo freeze-dryer (Thermo Savant, USA). These lyophilized pellets were digested for 1 h at ambient temperature using a solution containing 3 ml HNO<sub>3</sub>, 3 ml H<sub>2</sub>O, and 2 ml H<sub>2</sub>O<sub>2</sub>. The samples were heated in a microwave for 90 min. The intracellular concentration of the elements was then determined using ICP-MS. The accuracy of the method for element analysis was evaluated using bovine liver standard reference material (SRM) obtained from the National Institute of Standards and Technology (NIST SRM 1577b, USA). A total of 24 elements was measured, including iron (Fe), manganese (Mn), calcium (Ca), potassium (K), sulfur (S), barium (Ba), zinc (Zn), magnesium (Mg), phosphorous (P), boron (B), sodium (Na), vanadium (V), chromium (Cr), cobalt (Co), nickel (Ni), copper (Cu), arsenic (As), selenium (Se), rubidium (Rb), strontium (Sr), molybdenum (Mo), cesium (Cs), cadmium (Cd), and silver (Ag).

#### **2.4. Cell infection and sample preparation for $\mu$ -XRF imaging**

Endothelial cells were seeded on quartz slides (UQG Ltd., Cambridge, England) until they formed a monolayer (~ 24 h). *T. gondii* tachyzoites were then added to the cell monolayer at a MOI of 2. Control samples included quartz slides that were also seeded with the same number of cells, but without addition of tachyzoites. At 6, 24 and 48 hpi, three slides from each of the infected and non-infected ECs were washed 3X in sterile phosphate buffered saline (PBS; 137 mM sodium chloride, 2.7 mM potassium chloride, 10 mM disodium

phosphate, and 1.8 mM monopotassium phosphate; pH 7.4) and fixed in 4% paraformaldehyde in PBS for 30 minutes at ambient temperature. The slides were washed again 3X in PBS and left at 4°C in deionized water (17.8 MΩ-cm) until μ-XRF imaging.

## **2.5. Microfocus X-ray fluorescence imaging of metals**

The synchrotron microprobe of beamline I18<sup>18</sup> at Diamond Light Source, Oxfordshire, UK was utilized to map, and thereby quantify and compare, the individual distributions of Fe, Ca, and Zn within the ECs. Control samples were also analyzed to establish a baseline for the levels of these metals. Incubation times of 6, 24 and 48 hpi were measured for control and infected cells in order to evaluate the relative metal levels within the host cells at different stages after infection. The elemental distribution map of Fe could not be determined because the signals did not exceed the background level. It is likely that the ECs might have taken up Fe from the cell culture serum because ECs were cultured in media supplemented with animal serum. In contrast, the elemental distribution maps of Ca and Zn were successfully produced at a resolution of approximately 3 microns, over sample areas of 400 μm<sup>2</sup>. As the levels measured were known to be in the low parts per million (ppm) region, 4 sec/point dwell time was used for improved counting statistics. The elemental concentration was determined by measuring a reference material for μ-XRF measurements (AXO, Dresden GmbH) in the same conditions as the samples in order to deduce the photon flux on the sample. The reference material was composed of nm-thick layers of metals with known numbers of atoms per unit area. The resulting concentration maps were produced using Pymca, where the reference material and our sample were modelled in terms of main composition, density and thickness.<sup>19</sup> This analysis enabled fitting of the acquired spectra and translation of the fluorescence intensity maps for individual elements into estimated concentration distributions with an approximation to the composition of the cellular matrix.

The net peak areas from the sample spectra were translated into concentrations resulting in elemental maps in units of ppm (mg/g dry weight), which were then imported as tiff images in ImageJ for further statistical analysis. From the concentration maps regions of interest (ROIs) within cells were identified based on contrast to surrounding using the auto-threshold tool in ImageJ. From each ROI an average elemental value was extracted and plotted in scatter plots (Fig. 6).

The dehydration of the samples did not change the quantity of each material element sampled by the beam, as hydrogen and oxygen were not considered here, and the X-ray beam penetrated the full thickness of the sample. However, the absolute concentrations were necessarily an estimate due to the limitations of matrix matching with the reference standards, and to the inevitable variation in the thickness profile of individual cells. Also, it was important to note that the examined cell matrix was not a homogeneous bulk sample. ECs were not uniform in shape, or volume, their diameter ranged from 35-50  $\mu\text{m}$ , and the cell monolayer thickness might also vary. Therefore, considerable heterogeneity effects might have occurred, which can reduce the accuracy of estimated concentrations. The obtained values were tested for statistically significant differences between control and infected cells using Student's *t* test (two tailed). A *p*-value of  $< 0.05$  was considered the cut-off for a significant difference.

## **2.6. Elemental analysis at a single cell level**

We have developed a data-processing pipeline to estimate the elemental concentration in single cells (Fig. 1). To avoid issues of pile-up cells, we excluded spots where there was microscopic evidence of crowding and overlapping of cells. First, we aligned the bright field image (Fig. 1a) to the  $\mu$ -XRF fluorescent image (Fig. 1b) to ensure that the offset, scale, orientation and resolution of both images were completely matched. From the bright field

image, ROIs (i.e. individual cells) were selected, and the contour of each individual cell was manually drawn (Fig. 1c). These outlined cells were used as masks to crop and extract the information only from the ROIs in the  $\mu$ -XRF fluorescent images. The image annotated with the ROIs (Fig. 1c) was then transformed into a Boolean image, where the ROIs were coded as 1 and anything else as 0, as shown in Fig. 1d. The Boolean image was resized to the same resolution as the  $\mu$ -XRF fluorescent image (Fig. 1b) to ensure the correctness of the measured fluorescence intensity (Fig. 1e). The fluorescence intensity of the ROIs within the  $\mu$ -XRF fluorescent binary image was extracted (Fig. 1f). We used the connected-component labelling algorithm in MATLAB 2016b (MathWorks, Inc.) to quantify the fluorescence intensity signal from individual cells, at a single pixel scale, by labelling adjacent pixels (Fig. 1f). Each area of a connected component represented the number of the adjacent pixels in a single cell. The data represent the means of the element concentrations from at least 20 individual cells per sample  $\pm$  the SD. Although our image processing approach may not have fully quantified the complete fluorescence that represents the XRF spectra from single cells, its sensitivity was sufficient to quantify metal content inside individual cells in an objective and quantitative manner.

## **2.7. Gene expression analysis**

We explored whether *T. gondii* infection induced metal dyshomeostasis through modulation of metal transporters or metallothionein. EC monolayers were harvested using trypsin and lysed with QIAshredder columns (Qiagen). Total RNA was isolated using an RNeasy Mini Kit (Qiagen), and eluted with nuclease-free water. RNA was stored at  $-80$  °C for 1 week prior to microarray analysis. All subsequent sample handling, labelling and microarray (GeneChip<sup>TM</sup> Human Gene 2.1 ST, Affymetrix) processing was performed at the Nottingham Arabidopsis Stock Centre (NASCC). The RNA concentration and quality was assessed using

the Agilent 2100 Bioanalyzer (Agilent Technologies Inc., Palo Alto, CA) and the RNA 600 Nano Kit (Caliper Life Sciences, Mountain View, CA). Samples with a minimum RNA concentration of 100 ng/μl and RNA Integrity Number (RIN)  $\geq 8$  were used for gene expression analysis. Single stranded complimentary DNA was prepared from 200 ng of total RNA as per the GeneChip<sup>TM</sup> WT PLUS Reagent Kit (Applied Biosystems and Affymetrix). Total RNA was first converted to cDNA, followed by *in vivo* transcription to make cRNA. Single stranded cDNA was synthesized, end labeled and hybridized for 16 h at 45°C to GeneChip<sup>TM</sup> Human Gene 2.1 ST arrays. All steps were performed by a GeneAtlas<sup>TM</sup> Personal Microarray system (Affymetrix). Gene expression data were analyzed using Partek Genomics Suite 7.0 software (Partek Incorporated). The raw CEL files were normalized using the RMA background correction with quantile normalization, log base 2 transformation and mean probe-set summarization with adjustment for GC content. Differentially expressed genes were considered significant if *p*-value with false discovery rate (FDR) was  $\leq 0.05$  and fold change of  $>2$  or  $<-2$ .

## **2.8. Statistical analysis of the ICP-MS data**

Differences in the Mass Spectrometry-based elemental concentrations between control and infected cells at different time points were assessed by two-way ANOVA and Tukey's multiple comparison test (\*,  $p < 0.05$ ; \*\*,  $p < 0.01$ ; \*\*\*,  $p < 0.001$  ) using Graphpad Prism version 5.0 (Graphpad Software Inc., San Diego, CA). The data were presented as the mean and standard deviation (SD) from at least three independent experiments. A heatmap was produced to demonstrate the average elemental concentrations over the three time points post-infection, with the elements ordered according to the cluster analysis. The dendrograms were calculated using Ward's method on Euclidean distances. Because the dataset was not normally distributed, statistical analysis was performed on the transformed data ( $\log_{10}$ ). Next,

we, performed a volcano plot analysis where for each element and intra/extra-cellular condition we considered the most significant time point (the time point with the smallest  $p$ -value). Briefly, a linear model was fitted for each element, with the measurement as the response variable. The covariates were the time (treated as a factor with three levels) and the interaction between time and the control/infected condition, giving six parameters. The parameter estimate for the interaction parameters provided an estimate of the difference between the infected and control groups. The standard errors,  $t$ -values and  $p$ -values for the estimates were also evaluated. After correcting for multiple comparisons, a threshold of  $p < 0.00088$  was deemed to be significant. To classify sample by measurement of one element we applied a z-score classifier: mean and standard deviations of infected and uninfected samples were estimated. This method corresponds to one-dimensional Fisher's linear discriminant with a significance level of 99.9%.

### **3. Results**

#### **3.1. Elemental levels in *T. gondii* -infected ECs using ICP-MS**

The levels of elements within ECs and in culture medium infected with *T. gondii* compared with uninfected cells were determined using ICP-MS. Notably, the majority of elements did not show significant differences between control and infected cells. However, our analysis identified patterns associated with *T. gondii* infection and detected significant differences ( $p < 0.001$ ) between control and infected cell cultures (Fig. 2). We also noted that the intra- and extra-cellular measurements often differed by orders of magnitude. The intracellular concentrations of Zn, Fe, Mn, and Cu increased over time ( $p < 0.01$ ), whereas the levels of their counterparts in the extracellular culture medium, except Cu, remained nearly

unchanged. Intracellular levels of Co, Cu, Mo, V, and Ag remained unaltered, whereas their concentrations in the extracellular culture medium decreased over time ( $p < 0.01$ ).

To further validate the results, we looked at the differences in individual elements between infected and control cells using t-tests. The intracellular levels of Zn, Fe and Mn were significantly increased in infected cells compared with uninfected cells ( $p < 0.01$ ). The intracellular level of Cu also showed significant differences at 6 and 48 hpi ( $p < 0.001$ , as determined by ANOVA). The extracellular concentrations of Co, V, Mo, and Ag showed no difference as indicated by the overlap of intervals of infected and uninfected samples at 6 and 48 hpi and significant decrease in samples obtained at 24 hpi ( $p < 0.05$  for V and Mo;  $p < 0.01$  for Co and Ag, as determined by ANOVA). The accuracy of the z-score classifier for the intracellular level of Cu was 98.3%, 50% and 99.9%, whereas for the extracellular level it was 65.5%, 99.9% and 86.3%, at 6, 24 and 48 hpi, respectively.

The difference in elemental levels between control and infected cells was plotted against the corresponding  $-\log$  (base 10)  $p$ -value for testing significant differences. As shown in the supplementary Fig.S2, the Cu inside and outside the cells was the only element that was located above the horizontal line that represents the threshold of significance. This finding, together with the result showed in Fig. 2, suggests that the level of Cu was significantly different between infected and control samples.

We also performed unsupervised hierarchical clustering of the element concentrations. In the constructed dendrograms, the samples were separated based on the mean concentration of elements in the control and infected samples at 6, 24 and 48 hpi. The heatmap provided a graphical display of the temporal changes in the concentration of each element, with one row representing the mean control (Con) and one row representing the mean infected (Inf) cells of each sample (Fig. 3). Results shown in the heatmap further point out the significant increase in the concentration of Zn ( $p < 0.01$ ) and Fe ( $p < 0.05$ ) at 48 hpi. Next, the dendrogram (tree)

in the heatmap was divided to produce six different groups. The mean ( $\pm$  1 standard deviation) of the individual replicates of the elements that constituted each group were plotted at 6, 24, and 48 hpi. Elements within each of the six cluster groups shared an overall unique pattern (Fig. 4).

### **3.2. Elemental content of purified parasite preparation**

The elemental composition of *T. gondii* tachyzoites was also determined. Zn and Cu were the most abundant elements, representing 74% and 13% of the total parasite elemental content, respectively. Fe, Mn and P constituted 5%, 3% and 2%, respectively, whereas each of Na, S, and Rb accounted for  $\sim$ 1 % (Fig. 5).

### **3.3. Trace element levels in *T. gondii*-infected ECs using synchrotron $\mu$ -XRF**

The micrometer resolution of synchrotron-based X-ray fluorescence microscopy ( $\mu$ -XRF) enabled mapping and quantification of the level of elements in ECs infected with *T. gondii*. Maps of Ca and Zn at 6, 24, and 48 hpi of infected and control cells are shown in supplementary Figs. S3-4). Elemental imaging showed that Zn was significantly reduced only at 48 hpi compared with uninfected cells ( $p < 0.05$ ), whereas Ca was increased at 48 hpi in infected cells, but the difference was not statistically significant (Fig. 6). Average concentration of the metal content inside single cells was calculated using a connected-components labelling algorithm. The levels of Ca and Zn obtained using this image processing approach showed a similar trend to the results obtained based on the analysis of hot spots in the cell monolayer (Fig. 7).

### **3.4. Gene expression microarray data**

Affymetrix GeneChip™ Human Gene 2.1 ST microarray analysis of mRNA isolated from ECs, 6, 24, and 48 hpi revealed significant changes in overall gene expression. Principal component analysis (PCA) of gene expression profiles showed distinct transcriptional signatures among the six groups of samples (Fig. 8). Further analysis showed significant gene expression values ( $p$ -value with FDR was  $\leq 0.05$  and fold change  $>2$  or  $<-2$ ) in the metallothioneins (MT) - cysteine-rich, metal-binding proteins. Intensity values (LS mean) of the MT1JP, MT1M, MT1E, MT1F, and MT1X genes increased as the time of the infection reached 48 hpi (Fig. 9). These results were also confirmed by the heatmap and hierarchical clustering dendrogram (Fig. 10). Differential expression of MT1M and MT1JP genes was detected at 6, 24 and 48 hpi. Interestingly, MT1E, MT1F, MT1X genes showed lower gene expression at 6 hpi compared to MT1M and MT1JP. Gene expression increased gradually at 24 hpi for these three genes. The five genes showed similar gene expression after 48 hpi. These results show that the expression of MT1 isoforms JP, E, F, and X increased significantly in cells infected with *T. gondii* compared to uninfected cells.

#### **4. Discussion**

The availability and restriction of elements are important aspects of the host-pathogen interaction.<sup>7</sup> In the present study, we determined the changes in the concentrations of 24 trace elements in *T. gondii*-infected and uninfected ECs, the main constituent of the BBB, at 6, 24 and 48 hpi using ICP-MS. Significant increases were noted in the intracellular levels of Zn, Fe, Mn, and Cu ( $p < 0.01$ ) in infected cells compared with control cells. Significant decreases were noted in the extracellular levels of Co, Cu, Mo, V, and Ag in infected cells compared with the controls. Zn was also the most abundant element in the parasite elemental composition.

We also used synchrotron radiation-based  $\mu$ -XRF with  $< 5$  micron spatial resolution to assess the alterations in levels of Zn, Ca, Fe, and Cu distribution, as well as other elements in ECs following *T. gondii* infection. This elemental imaging showed Zn as the element most likely to be influenced during *T. gondii* infection of ECs. We also developed an image analysis pipeline to uniquely identify and analyse individual cells within cell monolayers and to quantify the metal content inside the selected individual cells. Results obtained by  $\mu$ -XRF imaging, whether based on complete cell monolayers or single cell levels, were consistent and showed a reduction in the level of Zn in infected cells at 48 hpi, indicating that single cell analysis was sufficiently sensitive to be representative of the intracellular metal concentration in the infected cell population. Reduction of Zn in infected cells at 48 hpi may be attributed to Zn sequestration by host cells, as a defense strategy to limit the intracellular growth of this parasite.

While the bulk ICP-MS analysis suggested elevated Zn in the infected cells,  $\mu$ -XRF imaging showed reduced concentration of Zn at 48 hpi. This discrepancy between the results obtained by the two methods might be accounted for by factors such as the heterogeneity of the cell population, as the ECs can have variable sizes, metal content, and in some cases cell overlapping occurred, which makes absolute quantification of metal content from the measured  $\mu$ -XRF difficult. By contrast, the bulk ICP-MS sampling could not be restricted to individual cells or well-characterized single monolayers. It is also likely that the concentration of Zn examined by  $\mu$ -XRF imaging may have been altered by fixation. The spatial distribution and concentration of the elements can be significantly altered in formalin-fixed samples<sup>20</sup> and in samples subjected to immunochemical treatment prior to imaging.<sup>21</sup> For example, 40% and 77% reduction in endogenous Fe and Zn, respectively has been observed in brain tissues fixed in formalin.<sup>22</sup> Substantial leaching of elements from the brain tissue into the formalin has been also detected, and the leaching varied considerably between

different elements and was time-dependent.<sup>23</sup> Therefore, fixation of the infected ECs, for health and safety reasons, with 4% paraformaldehyde prior to  $\mu$ -XRF imaging may have inevitably introduced an artificial effect and caused leaching of some elements from the cells into the fixative. It is therefore appropriate to cautiously interpret  $\mu$ -XRF imaging-based metal data derived from fixed cells in the present study. Future studies could compare the elemental differences between fixed samples and fresh or frozen samples, and assess new ways of preparing the samples with less potentially-disruptive protocols for  $\mu$ -XRF imaging analysis.

Microarray analysis of gene expression performed on infected and control cells using Affymetrix GeneChip revealed infection-specific upregulation of five genes; MT1JP, MT1M, MT1E, MT1F, and MT1X. These genes belong to the metallothionein (MT) gene family, encoding small, cysteine-rich, heavy metal-binding proteins, which play an important role in the homeostasis of transition metals (e.g. Fe, Zn and Cu) and detoxification of non-essential trace elements, such as cadmium (Cd) and mercury (Hg)<sup>24</sup>, and cell proliferation.<sup>25</sup> MTs are also stress response proteins that are induced in response to triggers, such as oxidative stress, infection, inflammation, and heavy metals,<sup>26</sup> to protect against reactive oxygen species (ROS),<sup>27</sup> via their free radical scavenging ability.<sup>28, 29</sup> Because agents that produce reactive oxygen intermediates are known to induce MT, one should anticipate a marked induction of MTs during *T. gondii* infection. Increased expression of these MT genes in infected cells compared to uninfected cells support the results obtained by ICP-MS, emphasizing the key role of Zn and other differentially abundant elements in mediating the host-parasite interaction.

Although the main focus of our work was a search for differential elemental levels in infected *versus* uninfected cells, it is noteworthy to highlight the clinical relevance of our findings. *T. gondii* infection stimulates immune cells to produce ROS and Th1-derived

cytokines, such as interferon-gamma (IFN- $\gamma$ ) to limit parasite growth.<sup>4</sup> The increased level of Zn in infected cells might be related to its anti-inflammatory and antioxidant activities<sup>30-32</sup> to protect host cells from oxidative stress and DNA damage,<sup>33-34</sup> known to be associated with *T. gondii* infection.<sup>35</sup> Zn can reduce the production of tumor necrosis factor-alpha (TNF- $\alpha$ ) and prevents the formation of free radicals.<sup>32</sup> This anti-oxidative stress (OS) effect of Zn has been attributed to down-regulation of the expression of ROS-producing inflammatory cytokines, such as TNF- $\alpha$  and interleukin-1 $\beta$ .<sup>36</sup> Zn can also limit nitric oxide (NO) production in endothelial cells by inhibiting NF- $\kappa$ B-dependent expression of inducible NO synthase (iNOS).<sup>37-38</sup> Further, Zn plays a role in maintaining the integrity of vascular endothelial cells, possibly by regulating signaling events to inhibit host cell death.<sup>39-41</sup> Zn deficiency can induce apoptosis<sup>42</sup> and disrupts cell membrane barrier integrity and increases the secretion of IL-8 and neutrophil transmigration.<sup>43</sup>

Our results also showed elevated levels of Fe in infected cells, indicating that Fe is essential for parasite growth. Fe is a cofactor of many enzymes involved in diverse cellular processes including respiration and DNA replication.<sup>44</sup> Obligatory intracellular parasites, such as *T. gondii*, possess mechanisms to obtain Fe from the host cell to sustain its own replication. This parasite hijacks Fe from the host cells by binding to transferrin receptor (TfR) during cell entry. The host expression of TfR was up-regulated in *T. gondii*-infected cells, suggesting that the parasite uses the host endocytic pathway and H.O.S.T. structure for Fe acquisition.<sup>45</sup> Our data also showed increases in the levels of Mn and Cu in *T. gondii*-infected cells compared with uninfected controls. Finally, a significant decrease in the levels of Co, Cu, Mo, V, and Ag was detected between infected and uninfected cells. The biological relevance of the differential levels of these elements in the pathogenesis of *T. gondii* infection remains to be elucidated.

## **5. Conclusion**

Elemental profiling of ECs infected with *T. gondii* was accomplished using a combination of sensitive ICP-MS analysis and non-invasive  $\mu$ -XRF imaging. Overall, our *in vitro* time course study showed a correlation between *T. gondii* infection and increased intracellular levels of Zn, Fe, Mn and Cu in the ECs, and increased expression of MT1 isoforms. These ICP-MS-based data suggest that *T. gondii* parasite promotes a rise in the intracellular level of Zn, potentially to sustain its own growth. Five metallothionein genes; MT1JP, MT1M, MT1E, MT1F, and MT1X, were upregulated in infected cells, providing support for increased intracellular zinc levels. These genes should be included in further studies of the effects of *T. gondii* infection on the regulation of metal transporters in host cells.  $\mu$ -XRF imaging of a heterogeneous cell population both at monolayer and individual cell levels revealed that Zn decreases in infected cells at 48 hpi. Nevertheless, these findings demonstrate the importance of further understanding of the regulatory processes involved in metal dyshomeostasis during *T. gondii* infection.

## **Conflicts of interest**

There are no conflicts of interest to declare.

## **Acknowledgements**

The authors gratefully acknowledge the support provided by The UK's National Synchrotron Science facility, Diamond Light Source at Harwell Science and Innovation Campus, Oxfordshire, for access to I18-the microfocus spectroscopy beamline (proposal number SP8504) that contributed to the X-ray fluorescence results presented in the manuscript.

## Notes and references

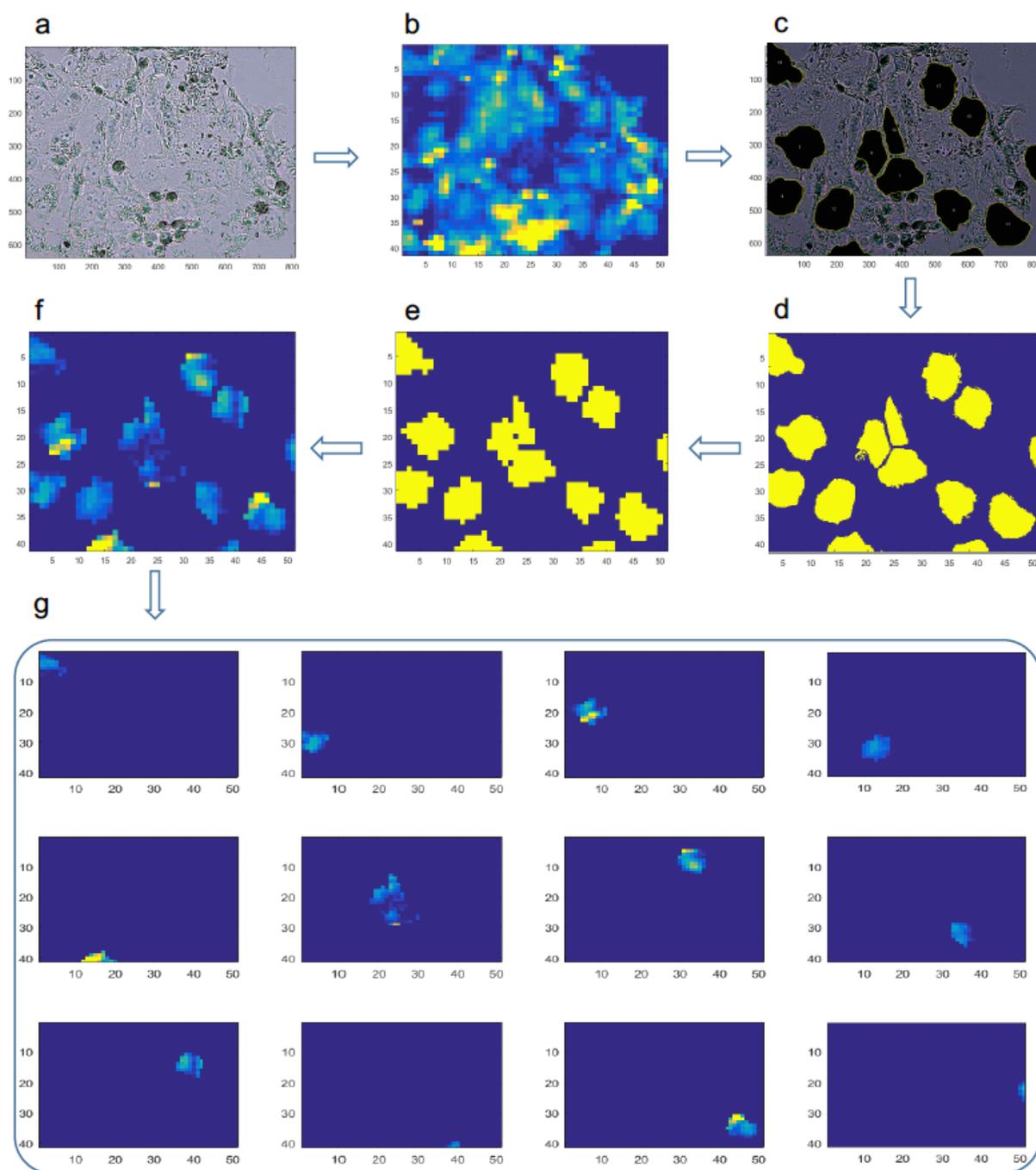
- 1 N. J. Abbott, Astrocyte–endothelial interactions and blood–brain barrier permeability, *J. Anat.*, 2002, **200**, 629–638.
- 2 U. Kniesel and H. Wolburg, Tight junctions of the blood–brain barrier, *Cell. Mol. Neurobiol.*, 2000, **20**, 57–76.
- 3 S. Eyal, P. Hsiao and J. D. Unadkat, Drug interactions at the blood–brain barrier: fact or fantasy?, *Pharmacol. Ther.*, 2009, **123**, 80–104.
- 4 H. M. Elsheikha and N. A. Khan, Protozoa traversal of the blood–brain barrier to invade the central nervous system, *FEMS Microbiol. Rev.*, 2010, **34**, 532–553.
- 5 Berg JM, Shi Y, The galvanization of biology: a growing appreciation for the roles of zinc, *Science* 1996, **271**,1081–1085.
- 6 A. H. Shankar and A. S. Prasad, Zinc and immune function: the biological basis of altered resistance to infection, *Am. J. Clin. Nutr.*, 1998, **68**, 447S–463S.
- 7 U. E. Schaible and S. H. Kaufmann, Iron and microbial infection, *Nat. Rev. Microbiol.*, 2004, **2**, 946–953
- 8 E. D. Weinberg, Iron loading and disease surveillance. *Emerg. Infect. Dis.*, 1999, **5**, 346–352.
- 9 K. Seyerk, S. Pasa, F. Kiral, A. Bildik, C. Babur and S. Kilic, Levels of zinc, copper and magnesium in sheep with toxoplasmosis, *Uludag. Univ. J. Fac. Vet. Med.*, 2004, **23**, 39–42.
- 10 E. M. Al-Khshab and H. S. Al-Bakry, Zinc and copper levels in seropositive toxoplasmosis women in Mosul – Iraq, *Rafidain J. Sci.*, 2009, **20**, 37– 41.
- 11 L. Yang, R. McRae, M. M. Henary, R. Patel, B. Lai, S. Vogt and C. J. Fahrni, Imaging of the intracellular topography of copper with a fluorescent sensor and by synchrotron x-ray fluorescence microscopy, *Proc. Natl. Acad. Sci. USA.*, 2005, **102**, 11179 –11184.
- 12 C. J. Fahrni, Biological applications of X-ray fluorescence microscopy: exploring the subcellular topography and speciation of transition metals, *Curr. Opin. Chem. Biol.*, 2007, **11**,121–127.

- 13 E. Malucelli, M. Fratini, A. Notargiacomo, A. Gianoncelli, L. Merolle, A. Sargenti, C. Cappadone, G. Farruggia, S. Lagomarsino and S. Iotti, Where is it and how much? Mapping and quantifying elements in single cells, *Analyst*, 2016, **141**, 5221–5235.
- 14 D. Hare, C. Austin and P. Doble, Quantification strategies for elemental imaging of biological samples using laser ablation-inductively coupled plasma-mass spectrometry. *Analyst*, 2012, **137**, 1527–1537.
- 15 H. M. Elsheikha, B. M. Rosenthal, A. J. Murphy, D. B. Dunams, D. A. Neelis and L. S. Mansfield, Generally applicable methods to purify intracellular coccidia from cell cultures and to quantify purification efficacy using quantitative PCR, *Vet. Parasitol.*, 2006, **135**, 223–234.
- 16 H. M. Elsheikha, C. L. McKinlay, N. A. Elsaied and P. A. Smith, Effects of *Neospora caninum* infection on brain microvascular endothelial cells bioenergetics. *Parasit. Vectors*, 2013, **6**, 24.
- 17 H. M. Elsheikha, M. Alkurashi, K. Kong and X. Q. Zhu, Metabolic footprinting of extracellular metabolites of brain endothelium infected with *Neospora caninum* in vitro, *BMC Res. Notes* 2014, **7**, 406.
- 18 J. F. Mosselmans, P. D. Quinn, A. J. Dent, S. A. Cavill, S. D. Moreno, A. Peach, P. J. Leicester, S. J. Keylock, S. R. Gregory, K. D. Atkinson and J. R. Rosell, I18-the microfocus spectroscopy beamline at the Diamond Light Source, *J. Synchrotron Radiat.*, 2009, **16**, 818–824.
- 19 V. A. Solé, E. Papillon, M. Cotte, P. H. Walter and J. Susini, A multiplatform code for the analysis of energy-dispersive X-ray fluorescence spectra, *Spectrochim Acta Part B: Atomic Spectroscopy*, 2007, **62**, 63–68.
- 20 M. Morello, A. Canini, P. Mattioli, R. P. Sorge, A. Alimonti, B. Bocca, G. Forte, A. Martorana, G. Bernardi and G. Sancesario, Sub-cellular localization of manganese in the basal ganglia of normal and manganese-treated rats - An electron spectroscopy imaging and electron energy-loss spectroscopy study. *Neurotoxicology*, 2008, **29**, 60–72.
- 21 Robison G, Zakharova T, Fu S, Jiang W, Fulper R, Barrea R, et al. (2012) X-Ray Fluorescence Imaging: A New Tool for Studying Manganese Neurotoxicity. *PLoS ONE* 7(11): e48899.

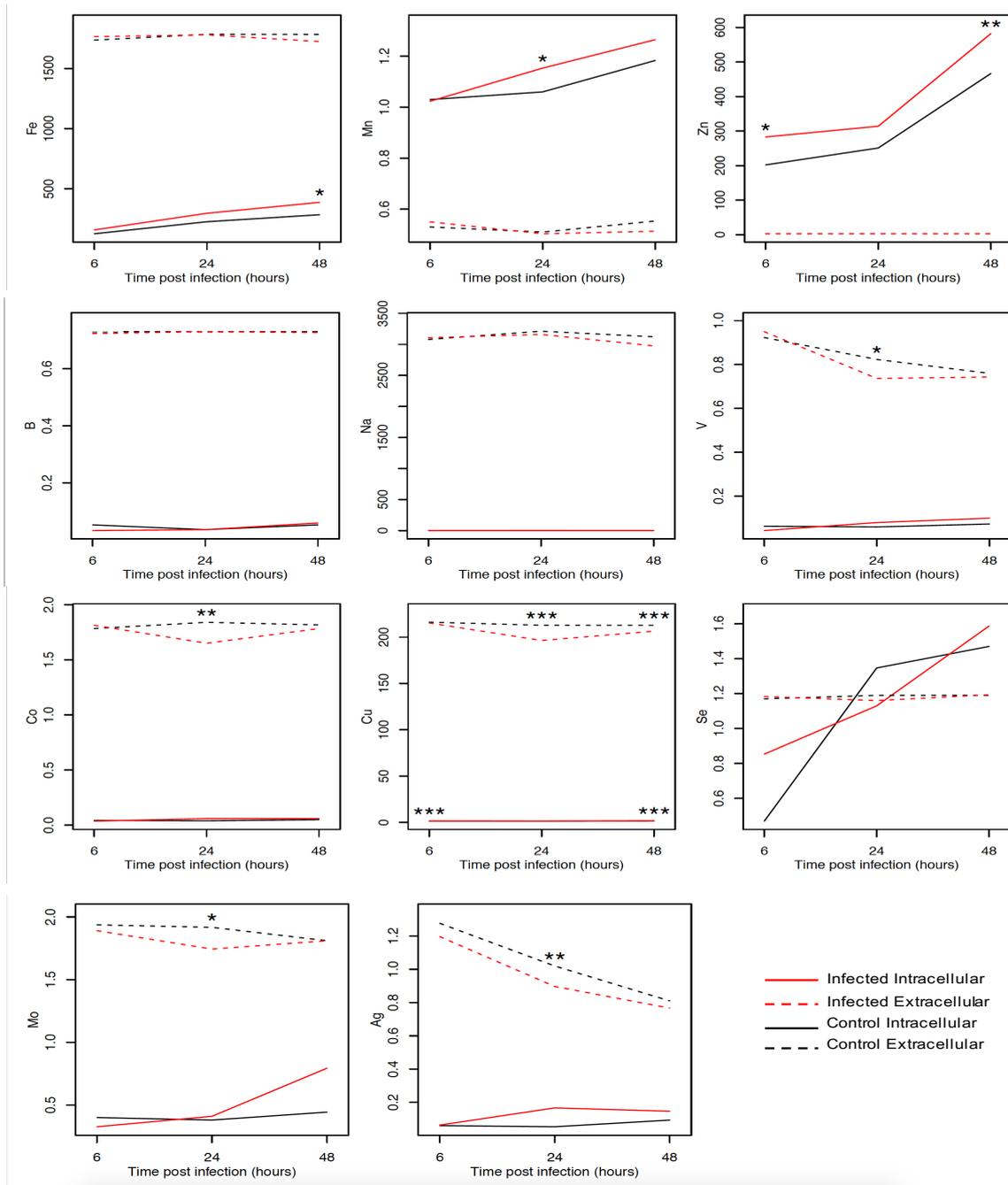
- 22 Schrag M, Dickson A, Jiffry A, Kirsch D, Vinters HV, Kirsch W. Biometals. The effect of formalin fixation on the levels of brain transition metals in archived samples. 2010 Dec;23(6):1123-7.
- 23 Gellein K, Flaten TP, Erikson KM, Aschner M, Syversen T. Leaching of trace elements from biological tissue by formalin fixation. *Biol Trace Elem Res.* 2008 Mar;121(3):221-5.
- 24 C. D. Klaassen, J. Liu and B. A. Diwan, Metallothionein protection of cadmium toxicity, *Toxicol. Appl. Pharmacol.*, 2009, **238**, 215–220.
- 25 R. Jin, J. Huang, P. H. Tan and B. H. Bay, Clinicopathological significance of metallothioneins in breast cancer. *Pathol. Oncol. Res.*, 2004, **10**, 74–79.
- 26 M. Yoshida, Y. Saegusa, A. Fukuda, Y. Akama and S. Owada, Measurement of radical scavenging ability in hepatic metallothionein of rat using in vivo electron spin resonance spectroscopy, *Toxicology*, 2005, **213**, 74–80.
- 27 B. Ruttkay-Nedecky, L. Nejdil, J. Gumulec, O. Zitka, M. Masarik, T. Eckschlager, M. Stiborova, V. Adam and R. Kizek, The role of metallothionein in oxidative stress, *Int. J. Mol. Sci.*, 2013, **14**, 6044–6066.
- 28 G. K. Andrews, Regulation of metallothionein gene expression by oxidative stress and metal ions, *Biochem. Pharmacol.*, 2000, **59**, 95–104.
- 29 M. Sato and I. Bremner, Oxygen free radicals and metallothionein, *Free Radic. Biol. Med.*, 1993, **14**, 325–337.
- 30 B. Bao, A. S. Prasad, F. W. Beck, J. T. Fitzgerald, D. Snell, G. W. Bao, T. Singh and L. J. Cardozo, Zinc decreases C-reactive protein, lipid peroxidation, and inflammatory cytokines in elderly subjects: a potential implication of zinc as an atheroprotective agent, *Am. J. Clin. Nutr.*, 2010, **91**,1634–1641.
- 31 A. S. Prasad, Effects of zinc deficiency on Th1 and Th2 cytokine shifts, *J. Infect. Dis.*, 2000, **182**, S62–68.
- 32 A. S. Prasad, F. W. Beck, B. Bao, J. T. Fitzgerald, D. C. Snell, J. D. Steinberg and L. J. Cardozo, Zinc supplementation decreases incidence of infections in the elderly: effect of zinc on generation of cytokines and oxidative stress, *Am J Clin Nutr.*, 2007, **85**, 837–844.

- 33 P. I. Oteiza, M. S. Clegg, M. P. Zago and C. L. Keen, Zinc deficiency induces oxidative stress and AP-1 activation in 3T3 cells, *Free Radic. Biol. Med.*, 2000, **28**, 1091–1099.
- 34 E. Ho, C. Courtemanche and B. N. Ames, Zinc deficiency induces oxidative DNA damage and increases p53 expression in human lung fibroblasts, *J. Nutr.*, 2003, **133**, 2543–2548.
- 35 E. Y. Denkers, L. Kim and B. A. Butcher, In the belly of the beast: subversion of macrophage proinflammatory signalling cascades during *Toxoplasma gondii* infection, *Cell. Microbiol.*, 2003, **5**, 75–83.
- 36 A. S. Prasad, B. Bao, F. W. Beck, O. Kucuk and F. H. Sarkar, Anti-oxidant effect of zinc in humans, *Free Radic. Biol. Med.*, 2004, **37**, 1182–1190.
- 37 J. D. Kruse-Jarres, The significance of zinc for humoral and cellular immunity, *J. Trace Elem. Electrolytes Health Dis.*, 1989, **3**, 1–8.
- 38 M. M. Cortese-Krott, L. Kulakov, C. Opländer, V. Kolb-Bachofen, K. D. Kröncke and C. V. Suschek, Zinc regulates iNOS-derived nitric oxide formation in endothelial cells. *Redox Biol.*, 2014, **2**, 945–954.
- 39 P. Meerarani, P. Ramadass, M. Toborek, H. C. Bauer, H. Bauer and B. Hennig, Zinc protects against apoptosis of endothelial cells induced by linoleic acid and tumor necrosis factor alpha, *Am. J. Clin. Nutr.*, 2000, **71**, 81–87.
- 40 F. W. Jr. Sunderman, The influence of zinc on apoptosis, *Ann. Clin. Lab. Sci.*, 1995, **25**, 134–142.
- 41 D. Beyersmann and H. Haase, Functions of zinc in signaling, proliferation and differentiation of mammalian cells, *Biometals*, 2001, **14**, 331–341.
- 42 M. Seve, F. Chimienti and A. Favier, Role of intracellular zinc in programmed cell death, *Pathol. Biol. (Paris)*, 2002, **50**, 212–221.
- 43 A. Finamore, M. Massimi, L. Conti Devirgiliis and E. Mengheri, Zinc deficiency induces membrane barrier damage and increases neutrophil transmigration in Caco-2 cells, *J. Nutr.*, 2008, **138**, 1664–1670.
- 44 J. E. Cassat and E. P. Skaar, Iron in infection and immunity, *Cell Host Microbe.*, 2013, **13**, 509–519.

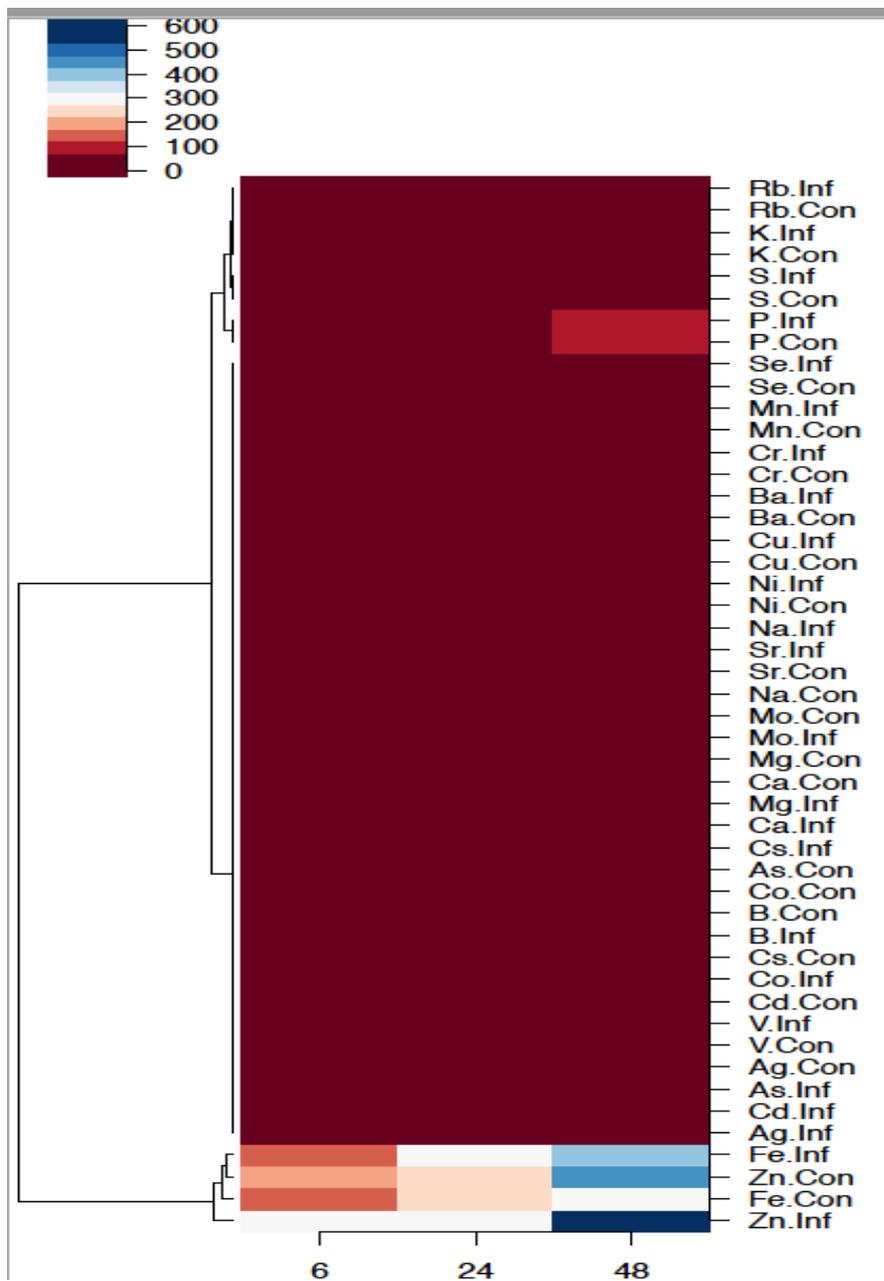
45 M. Gail, U. Gross and W. Bohne, Transferrin receptor induction in *Toxoplasma gondii*-infected HFF is associated with increased iron-responsive protein 1 activity and is mediated by secreted factors, *Parasitol. Res.*, 2004, **94**, 233–239.



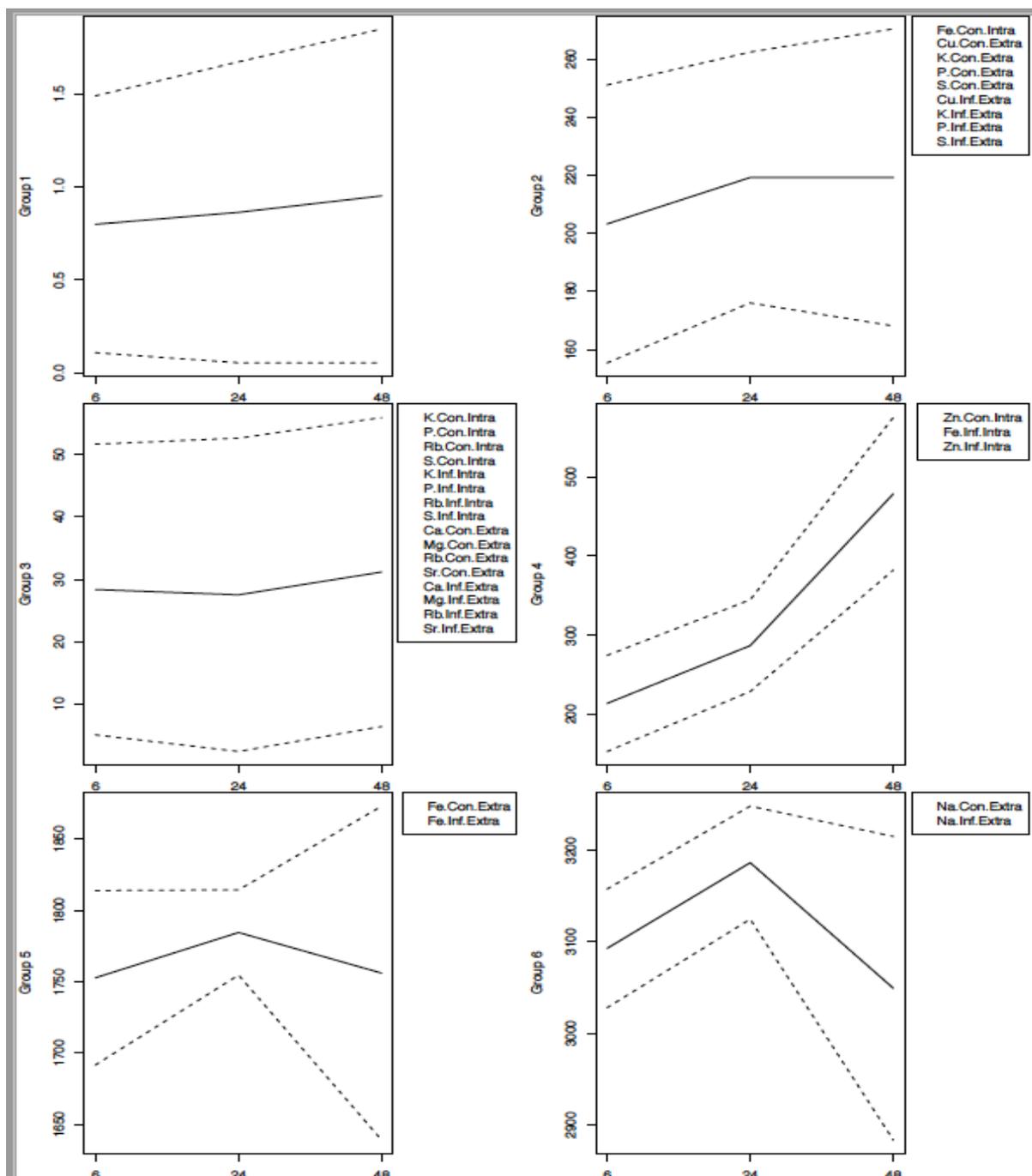
**Fig. 1** Single cell elemental analysis workflow. (a) The bright field image was aligned against (b) the  $\mu$ -XRF elemental map, and (c) regions of interest (i.e. single cells) in the bright field image were outlined. We then masked the regions of interest on the  $\mu$ -XRF elemental map (d-f). Individual cells were separated, and the mean and standard deviation of each element were derived per cell, followed by calculating the mean and standard error for each time point (g).



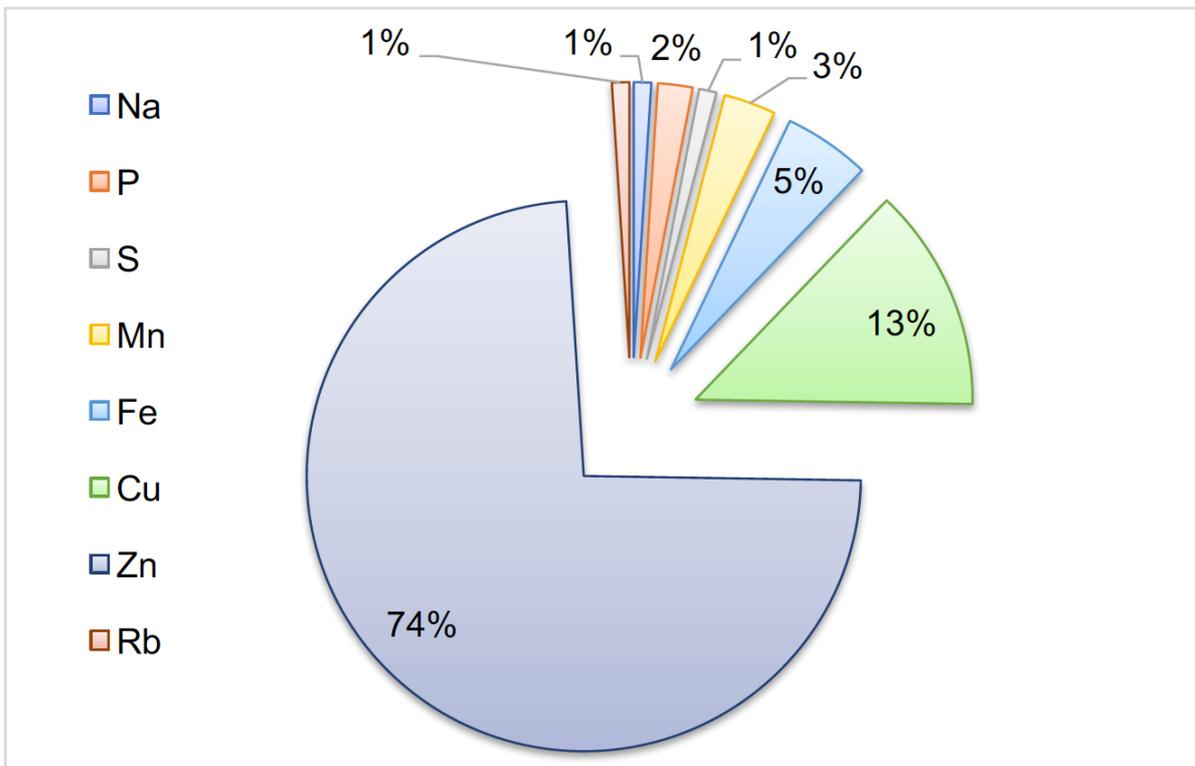
**Fig. 2** Plots showing dynamic changes in the levels of elements within cells and in culture medium following infection using ICP-MS. Results shown represent the mean value with one standard deviation width interval for each group after taking the arithmetic average across the three replicates available for each element at each time point. Each panel shows the kinetic signature of one element detected inside the EC culture or in the supernatant at 6, 24 and 48 hours post infection (hpi) with *T. gondii* (X-axis). The Y-axis denotes concentrations of the metals ( $\mu\text{g/L}$ ). Black and red lines represent control and infected samples, respectively. Solid and dashed lines represent the intracellular and extracellular elemental concentrations, respectively. Following infection the levels of Zn, Fe, Mn, and Se displayed increasing levels inside the cells, whereas Co, Cu, Mo, and Ag displayed decreasing levels outside the cells (\* $p < 0.05$ , \*\* $p < 0.01$ . \*\*\* $p < 0.001$ ).



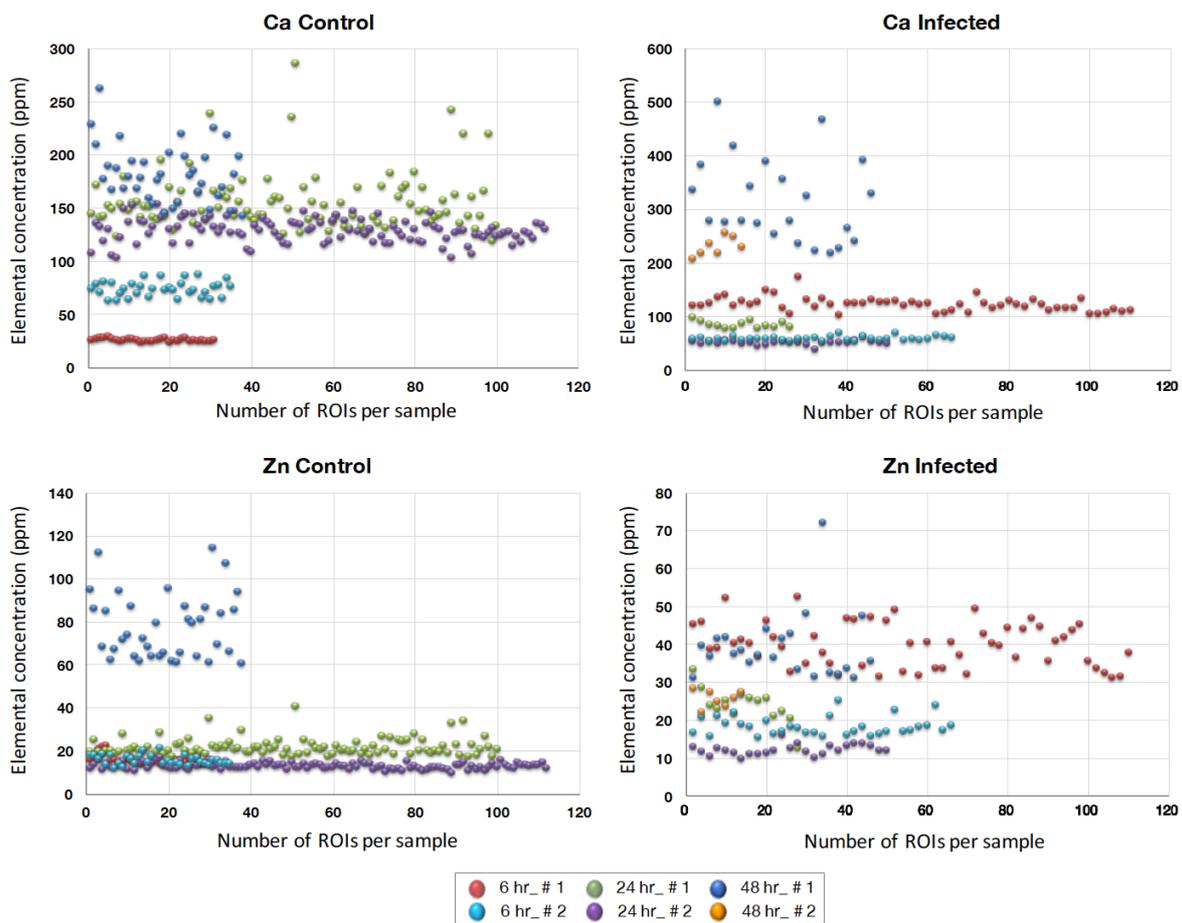
**Fig. 3** A heatmap and cluster tree representation showing the fluctuations in the concentration of elements between infected and uninfected ECs over the course of infection using a colour-scale, with the elements ordered according to the cluster analysis. Average concentrations of the control and infected samples over the course of infection were calculated for each element by taking the mean over control replicates and infected replicates, respectively. Hierarchical cluster analysis combined with a heat map revealed trends across time points after infection (X-axis) and between the measured elements (Y-axis). Each element is represented by two rows, average control (Con) and average infected (Inf), and each time point after infection by a column. The colour scale indicates the concentration value (dark red indicates lower elemental concentration values and dark blue indicates higher concentration values). The dendrogram was calculated using Ward's method on Euclidean distances between the average response vectors.



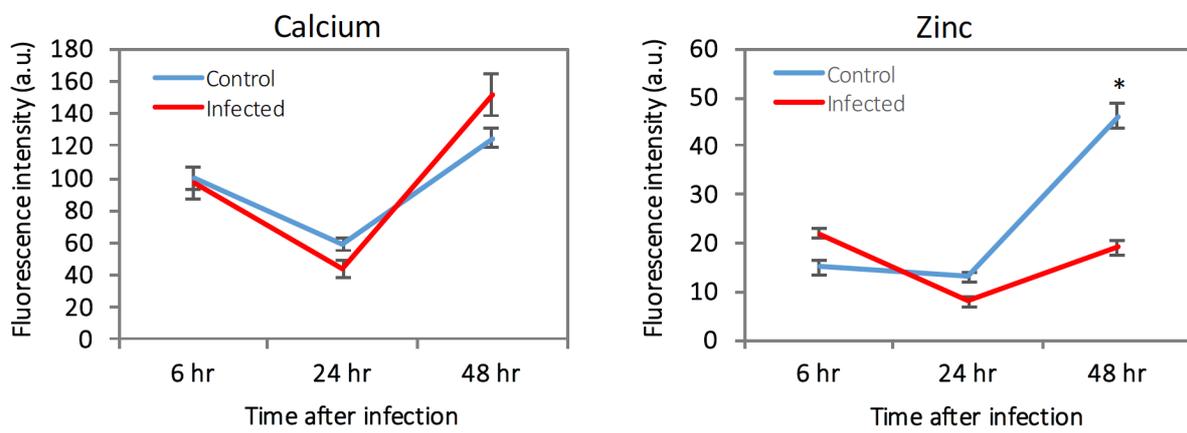
**Fig. 4** Overall trends of the elements within the sub-clusters. The hierarchical clustering was divided into six groups of elements. The number of elements, including infected (Inf) and control (Con), and intracellular (Intra) and extracellular (Extra), in each cluster are listed in the box next to the corresponding group, with the exception of group 1, which contained the remaining elements. The mean (solid line) was calculated across the individual replicates, which constituted the elements in each group. The kinetic shape templates (mean  $\pm$  1 SD; dashed line) represent the span of all the elemental profiles of similar dynamic patterns in a single cluster. The x-axis denotes time in hours post infection and the y-axis represents the elemental concentration.



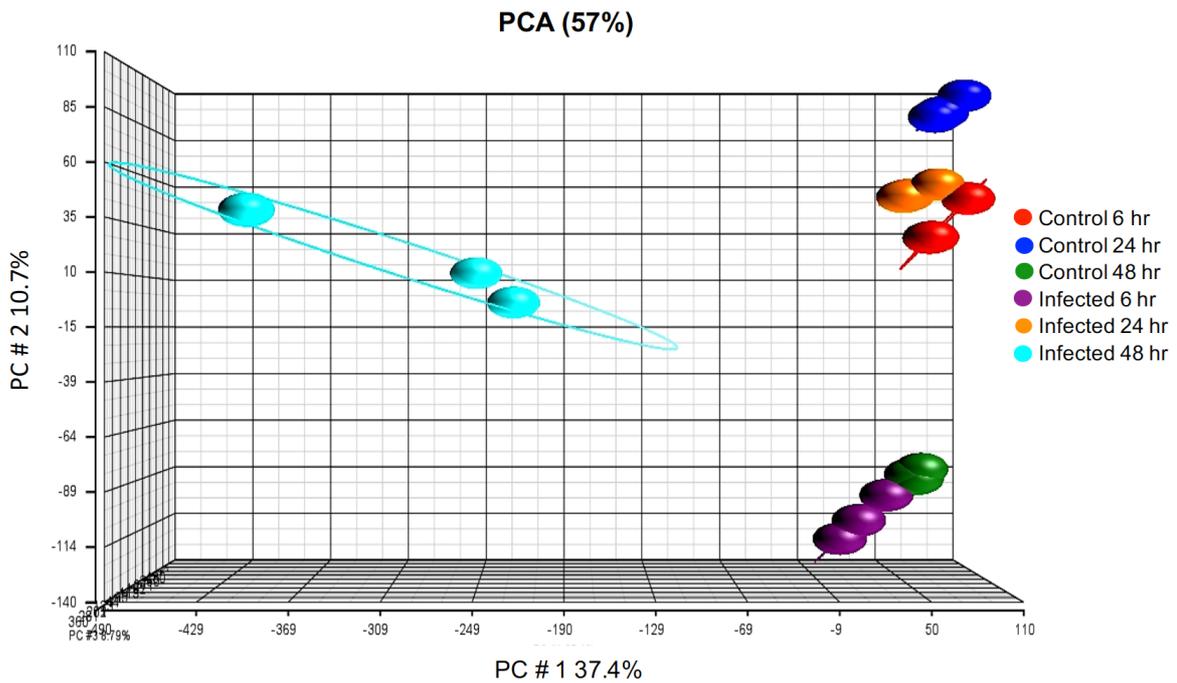
**Fig. 5** Elemental composition of the parasite. Proportions of trace elements in *T. gondii* tachyzoites. Sodium (Na), phosphorous (P), sulfur (S), manganese (Mn), iron (Fe), copper (Cu), zinc (Zn), and rubidium (Rb).



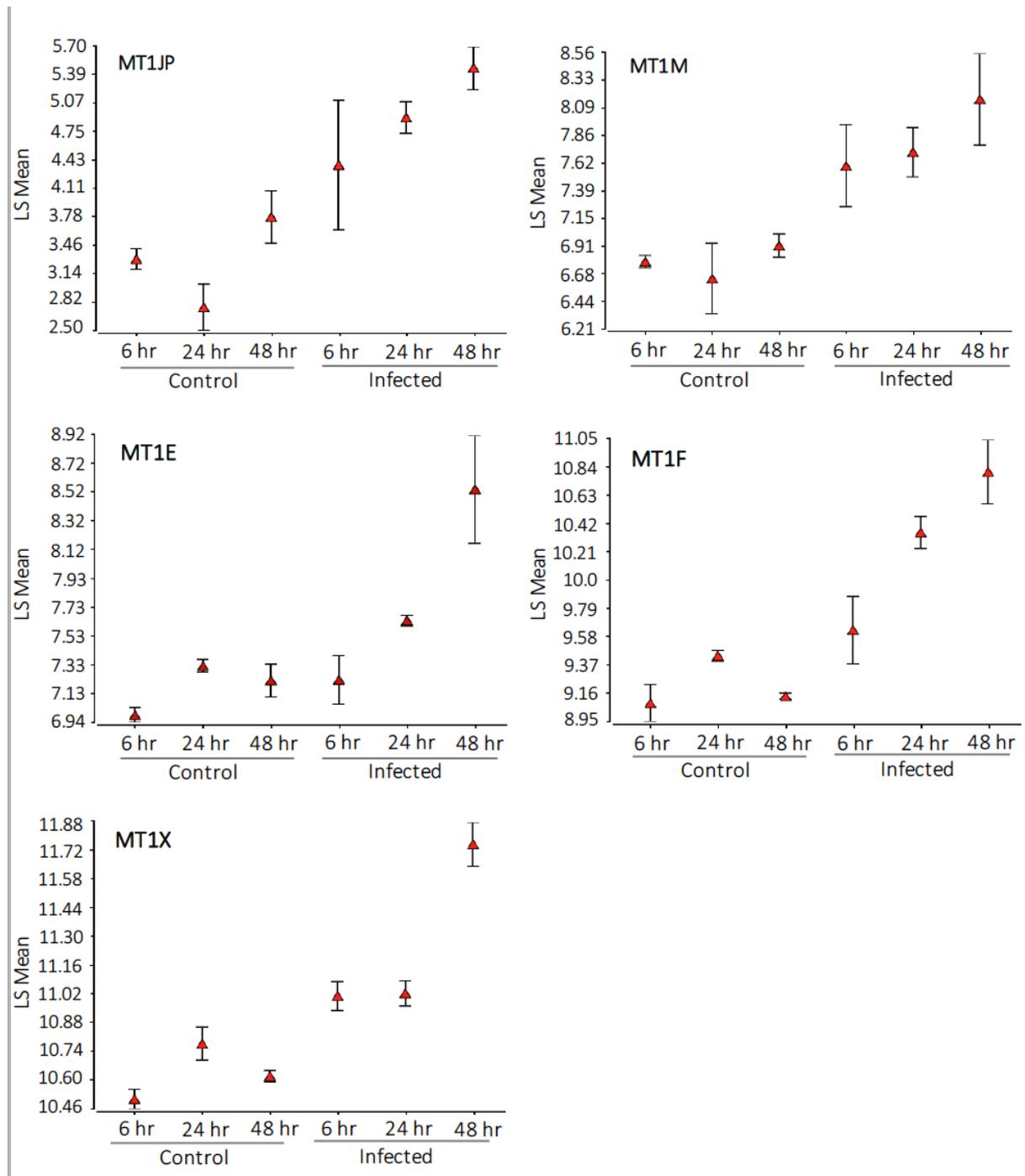
**Fig. 6** Scatterplots showing the differences in the levels of Ca and Zn measured by  $\mu$ -XRF at different time points after infection, in the infected cells and their corresponding controls. The X-axis indicates the number of the regions of interest (ROIs) analyzed per sample and the Y-axis represents the concentrations of the elements (ppm). The values represent two biological replicates (indicated as # 1 and # 2) from infected and uninfected (control) samples at the three indicated time points following infection.



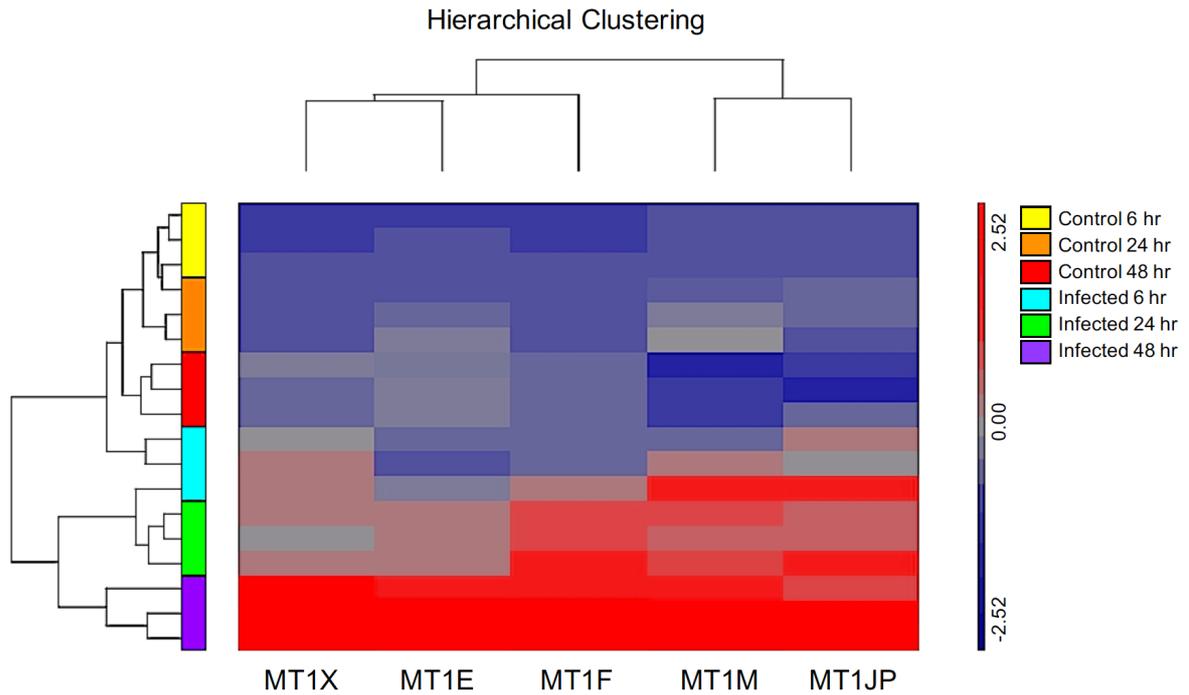
**Fig. 7** The intracellular levels of Ca and Zn determined by single cell analysis of the  $\mu$ -XRF elemental maps.  $p < 0.05$  is indicated by (\*).



**Fig. 8** Principal component analysis (PCA) showing transcriptomic differences between expression microarray data of both uninfected and infected ECs. Samples were analyzed at 6, 24 and 48 hr after exposure to *T. gondii*. Each sample group is represented by a sphere and is color-coded to indicate the corresponding control/infection category. There was a clear difference in the transcriptomic clustering at 48 hr between uninfected and infected samples, suggesting that infection had an influence on gene expression changes in the host cells over time.

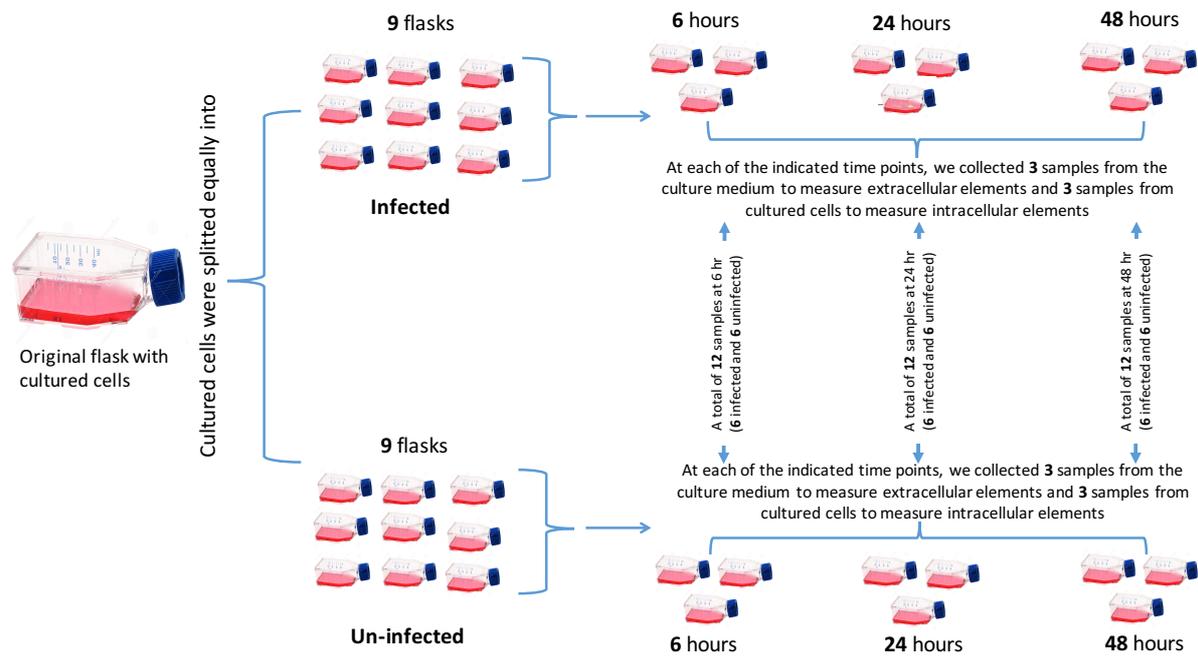


**Fig. 9** XY plots showing the intensity of each metallothionein gene (MT1X, MT1E, MT1F, MT1M and MT1JP) across sample groups. Each point (triangle) is the LS mean value of all samples from the same group. The intensity value for the five metallothionein genes increased as the time of the infection progressed compared to controls.

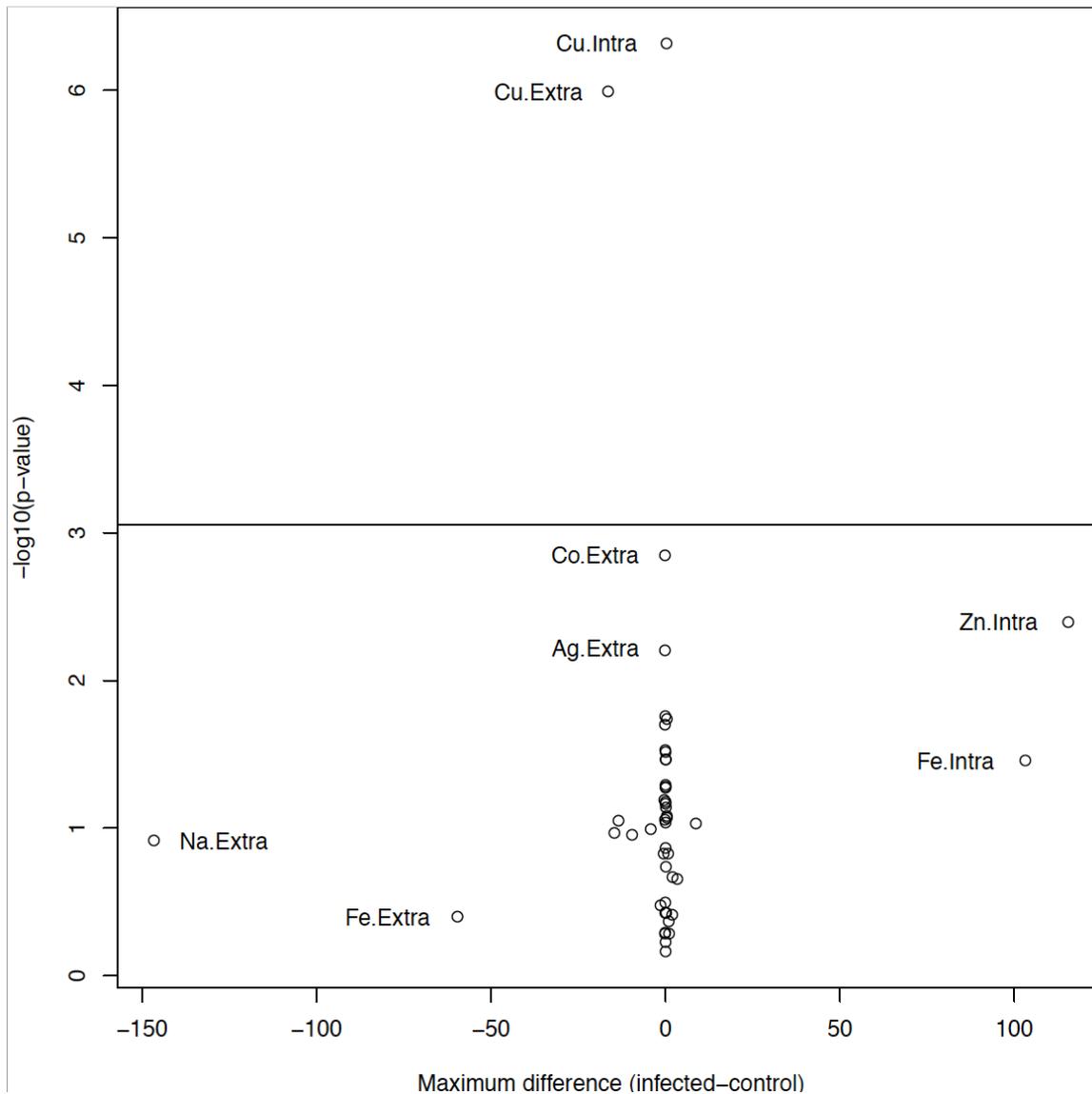


**Fig. 10** Heatmap and hierarchical clustering dendrogram for five metallothionein genes (MT1X, MT1E, MT1F, MT1M and MT1JP). Results are expressed as fold change of gene expression of infected ECs compared to uninfected ECs at 6 hr, 24 hr and 48 hr after exposure to *T. gondii*. The red color represents upregulation of expression, grey color indicates an unchanged expression, and blue color represents downregulation of expression. The pattern and length of the branches in the dendrogram reflect the relatedness of the samples. Expression of the metallothionein genes was mostly downregulated in all uninfected samples, whereas in infected samples gene expression increases as infection progresses.

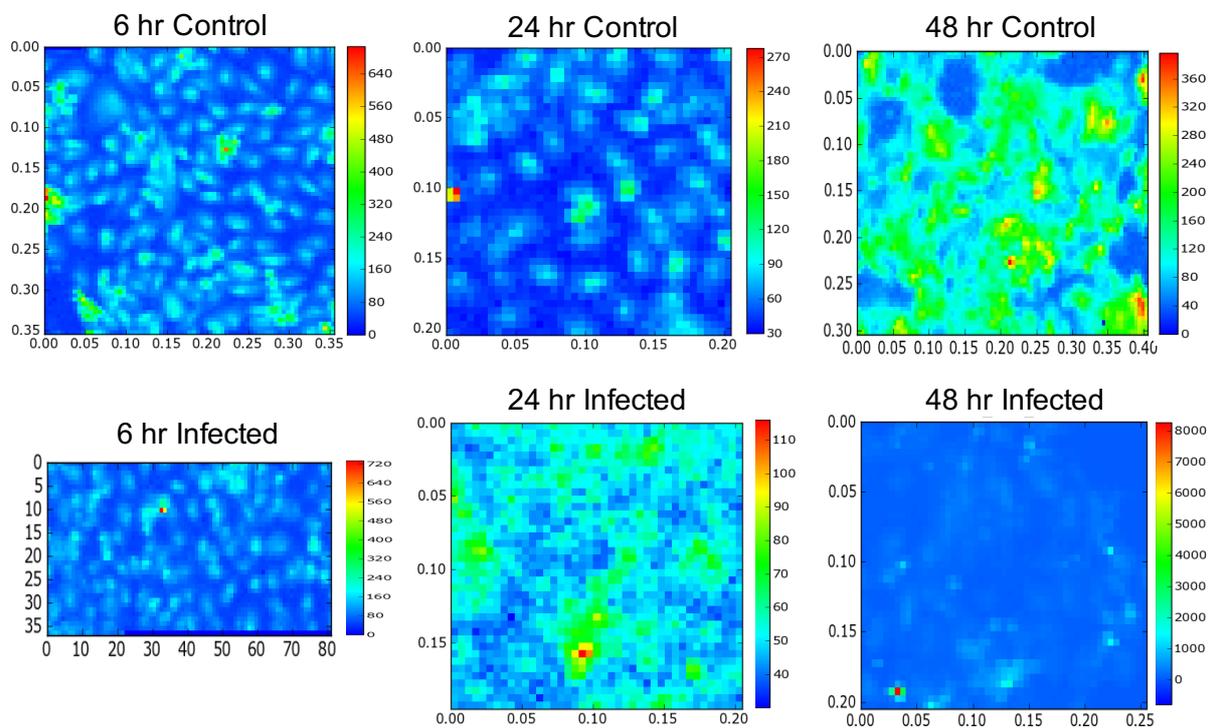
## Supplementary Figures



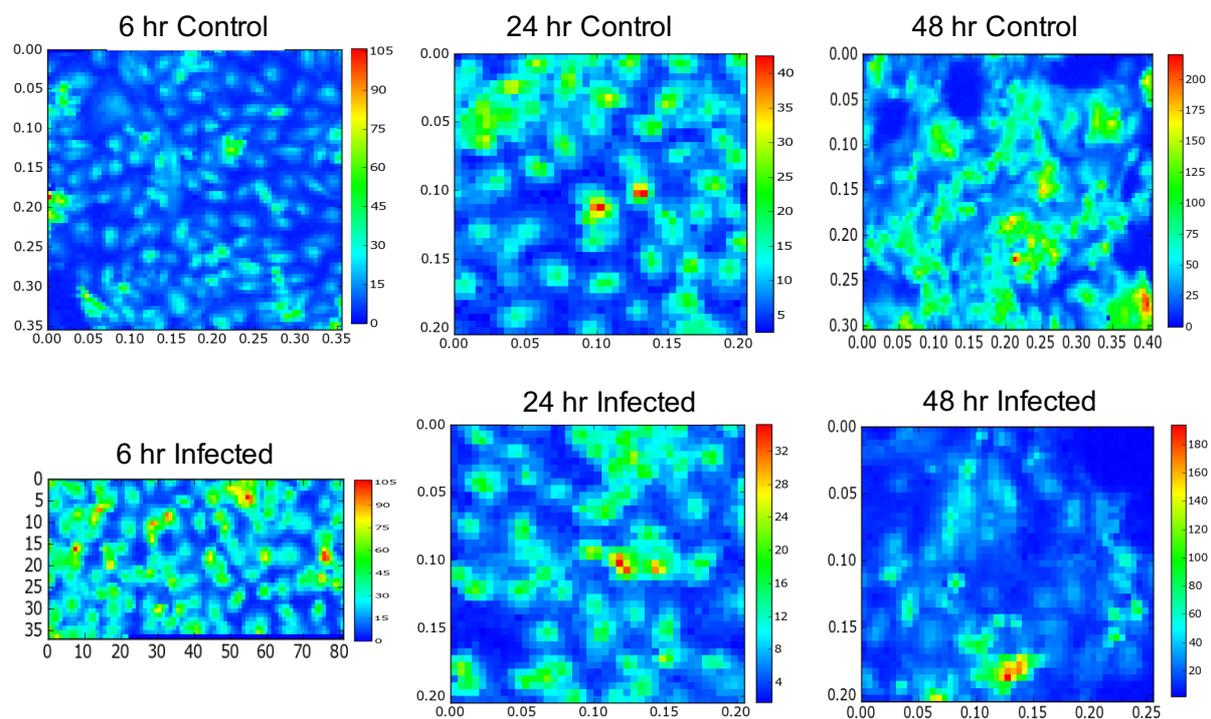
**Suppl. Figure 1: Schematic illustration outlining the groups of samples used to examine the levels of intracellular elements.** Following collection of the culture medium the cultured cells were harvested, extracted and the concentrations of the intracellular elements were determined.



**Suppl. Figure 2: Volcano plot showing the differentially abundant elements.** For each element, a linear model was fitted with the measurement as the response variable. The parameter estimate for the interaction parameters between time and control/infected condition provided an estimate of the difference between the infected and control samples at each time point. The most significant differences (the time point with the smallest  $p$ -value) for intra and extra cellular elements are shown in the plot. Elements are represented as individual circles and plotted along the x-axis by the effect size (the maximum elemental concentration difference between control and infected samples across the three time points after infection), and on the y-axis by the significance level,  $-\log_{10}(p\text{ value})$ . The horizontal line indicates the significance threshold ( $p < 0.00088$ ) after correcting for multiple tests, with elements above the line being significant and those below the line on-significant. Non-labelled circles represent those elements that had non-significant differences between infected and control samples.



**Suppl. Figure 3: High-resolution elemental map of Calcium (Ca) in endothelial cells infected with *Toxoplasma gondii* acquired by  $\mu$ -XRF at  $\sim 3$  microns resolution. (A-C) Ca  $\mu$ -XRF maps showing changes in the distribution and quantity of Ca at 6 hr, 24 hr, and 48 h post infection compared to the corresponding controls (D-F). Note the specific focal accumulation of the element (arrows). The rainbow-colored scale bars reflect the signal intensity and correspond to elemental concentrations in the range presented for each map, with darker pixels representing areas of lower concentration and brighter pixels representing areas of higher concentration.**



**Suppl. Figure 4: High-resolution elemental map of Zinc (Zn) in endothelial cells infected with *Toxoplasma gondii* acquired at the  $\mu$ -XRF at  $\sim 3$  microns resolution. (A-C) Zn  $\mu$ -XRF maps showing changes in the distribution and quantity of Zn at 6, 24 and 48 hr after infection compared to the corresponding controls (D-F). Note the specific focal accumulation of the element (arrows). The rainbow-colored scale bars reflect the signal intensity and correspond to elemental concentrations in the range presented for each map, with darker pixels representing areas of lower concentration and brighter pixels representing areas of higher concentration.**