



# General Commentary on: Alternatives to Antibiotic Growth Promoters in Animals

Blessing O. Anonye\*

Microbiology and Infection Unit, Division of Biomedical Sciences, Warwick Medical School, University of Warwick, Coventry, UK

**Keywords:** antibiotics growth promoters, phytogetic feed additives, microbiota, antibacterial activity, probiotics

## A commentary on

### Development and Evaluation of a Herbal Formulation with Antipathogenic Activities and Probiotics Stimulatory Effects

by Qian Z, Si-Si W, Guang Y, Wen Z, Hui-Ling L. *J Integr Agric* (2016) 15:1103–11. doi:10.1016/S2095-3119(15)61146-7

### Phytogetic Feed Additives as an Alternative to Antibiotic Growth Promoters in Broiler Chickens

by Murugesan GR, Syed B, Haldar S, Pender C. *Front Vet Sci* (2015) 2:21. doi:10.3389/fvets.2015.00021

## OPEN ACCESS

### Edited by:

Guillermo Tellez,  
University of Arkansas, USA

### Reviewed by:

Kenneth James Genovese,  
United States Department  
of Agriculture, USA  
Lisa Bielke,  
Ohio State University, USA

### \*Correspondence:

Blessing O. Anonye  
b.anonye@warwick.ac.uk

### Specialty section:

This article was submitted to  
Veterinary Infectious Diseases,  
a section of the journal  
Frontiers in Veterinary Science

**Received:** 04 June 2016

**Accepted:** 23 August 2016

**Published:** 31 August 2016

### Citation:

Anonye BO (2016)  
General Commentary on:  
Alternatives to Antibiotic  
Growth Promoters in Animals.  
*Front. Vet. Sci.* 3:74.  
doi: 10.3389/fvets.2016.00074

Recent studies have evaluated the impact of alternatives to antibiotic growth promoters (AGPs) such as phytogetic feed additives (PFAs) *in vitro* and *in vivo*. Zhou and colleagues studied the antibacterial properties of 30 herbs on pathogenic Gram-negative and positive bacteria (1). Thirteen of the 30 herbs exerted a significant effect against *Escherichia coli* ATCC 25922 ( $p < 0.01$ ,  $n = 11$  and  $p < 0.05$ ,  $n = 2$ ). More than 30% of the herbs exhibited activity against *Salmonella enteritidis* ATCC 13076 and *Salmonella typhimurium* ATCC 14028. These pathogenic organisms commonly infect humans and animals especially poultry such as chickens leading to loss and decrease in their market value. Additionally, Zhou reported that more than 50% of the herbal extracts ( $n = 16$ ) possessed antibacterial activity against *Staphylococcus aureus* ATCC 25923.

Furthermore, based on the results of the individual herbs on the pathogenic bacteria, two formulations were performed with five of the herbs. For formulation 1, the herb, *Fructus mume* was the main ingredient (35%) and *Galla chinensis* (30%) for formulation 2 with varying proportions of four other herbs. Both formulations 1 and 2 had significant antibacterial activity against the pathogenic bacteria ( $p < 0.05$ ) with no significant difference in activity when compared to the AGPs, aureomycin, and flavomycin (1). Formulation 1 also led to increased counts of *Lactobacillus acidophilus* ATCC 4356 and *Bifidobacterium longum* ATCC 15707 relative to the control, indicating the possibility that these herbs could have a synergistic effect on beneficial bacteria in the intestinal microbiota. This raises the potential of using herbs as an alternative to antibiotics to increase growth in animals and modulate the microbiota. However, these herbs must be carefully chosen as formulation 2 did not produce the same probiotic effects as formulation 1 but led to reduced amounts of the *L. acidophilus* and *B. longum* compared to the control.

Similarly, Murugesan et al. compared the effects of Digestarom® Poultry, a commercial PFA produced by BIOMIN, to the AGP, bacitracin methylene disalicylate in broiler chickens (2). Chicks were randomly assigned to receive either a corn-soybean meal only or supplemented with the PFA or AGP, respectively, over a 39-day period. This period was divided into pre-starter (days 1–7), starter (days 8–21), and grower (days 22–39) phases. The authors noted differences based on the period of growth. For example, in the starter phase, AGP-fed birds gained more body weight relative to control, while PFA-fed birds had increased body weight in the grower phase. Also, increase in the villus height

across the small intestine was observed in birds fed with AGP or PFA relative to control (2). As the villi help to increase the surface area of the intestinal walls, an increase in digestion and absorption of nutrients is likely to be observed (3). Coliforms were significantly decreased ( $p < 0.01$ ), and *Lactobacillus* spp. was significantly increased ( $p < 0.01$ ) through plating of the cecal microbiota when compared to the control or AGP-fed birds. Similar results have been obtained with respect to increased *Lactobacillus* spp. in PFA-fed birds using similar PFA as Murugesan et al. (4) or a phytoncide (5). However, next-generation sequencing could provide a better picture of the changes taking place in the cecal microbiota with respect to the bacterial groups.

The results from these and other studies suggest the ability of the PFAs to modulate the intestinal microbiota. These could occur through various mechanisms by influencing the digestibility of nutrients and thereby enhancing the growth performance of the animals (1, 2, 5). PFAs can potentially stimulate the secretion of digestive enzymes, thereby promoting gut functions. Moreover, the bioactive compounds produced by the PFAs have been shown to possess antibacterial properties *in vivo* against chickens challenged with *S. enteritidis*, *E. coli*, and *Clostridium perfringens* (4, 6). PFAs, such as *F. mume*, may exert their antibacterial effect through the production of organic acids, leading to increased acidity as revealed by HPLC (7). Another possible mechanism by

which PFAs exert their beneficial effects is by acting as antioxidant against oxidative stress in animals.

However, the search for PFAs with these desirable properties is not trivial. Single and different combinations of PFAs need to be tested against different strains of pathogenic bacteria *in vitro* and *in vivo* to determine their antimicrobial activity. A desirable PFA ideally should be able to stimulate the gut microbiota in a number of ways. This could be through increasing colonization resistance without having any adverse effect on beneficial bacteria and creating a favorable environment for increased nutrient intake leading to weight gain. Both studies described above have shown to some extent these desirable characteristics, but more studies will be needed to determine the exact mode of action of these PFAs.

It would also be worth looking at other alternatives to antibiotics such as prebiotics, probiotics, and bacteriocins to determine their effects on animals (8, 9). The development of PFAs and other substances that can give similar or more beneficial outcomes, as the AGPs will go a long way in reducing the increase of antibiotic-resistant bacteria.

## AUTHOR CONTRIBUTIONS

The author confirms being the sole contributor of this work and approved it for publication.

## REFERENCES

- Zhou Q, Wang S, Yang G, Zhao W, Li H. Development and evaluation of a herbal formulation with anti-pathogenic activities and probiotics stimulatory effects. *J Integr Agric* (2016) 15:1103–11. doi:10.1016/S2095-3119(15)61146-7
- Murugesan GR, Syed B, Haldar S, Pender C. Phytogetic feed additives as alternative to antibiotic growth promoters in broiler chickens. *Front Vet Sci* (2015) 2:21. doi:10.3389/fvets.2015.00021
- Koh-en Y. Review on chicken intestinal villus histological alterations related with intestinal function. *J Poult Sci* (2002) 39:229–42. doi:10.2141/jpsa.39.229
- Wati T, Ghosh TK, Syed B, Haldar S. Comparative efficacy of a phytogetic feed additive and an antibiotic growth promoter on production performance, caecal microbial population and humoral immune response of broiler chickens inoculated with enteric pathogens. *Anim Nutr* (2015) 1:213–9. doi:10.1016/j.aninu.2015.08.003
- Li HL, Zhao PY, Lei Y, Hossain MM, Kim IH. Phytoncide, phytogetic feed additive as an alternative to conventional antibiotics, improved growth performance and decreased excreta gas emission without adverse effect on meat quality in broiler chickens. *Livest Sci* (2015) 181:1–6. doi:10.1016/j.livsci.2015.10.001
- Cho JH, Kim HJ, Kim IH. Effects of phytogetic feed additive on growth performance, digestibility, blood metabolites, intestinal microbiota, meat color and relative organ weight after oral challenge with *Clostridium perfringens* in broilers. *Livest Sci* (2014) 160:82–8. doi:10.1016/j.livsci.2013.11.006
- Chen Y, Wong RWK, Seneviratne CJ, Hägg U, McGrath C, Samaranyake LP, et al. The antimicrobial efficacy of *Fructus mume* extract on orthodontic bracket: a monospecies-biofilm model study *in vitro*. *Arch Oral Biol* (2015) 56:16–21. doi:10.1016/j.archoralbio.2010.08.006
- Huyghebaert G, Ducatelle R, Immerseel FV. An update on alternatives to antimicrobial growth promoters for broilers. *Vet J* (2011) 187:182–8. doi:10.1016/j.tvjl.2010.03.003
- Caly DL, D’Inca R, Auclair E, Drider D. Alternatives to antibiotics to prevent necrotic enteritis in broiler chickens: a microbiologist’s perspective. *Front Microbiol* (2015) 6:1336. doi:10.3389/fmicb.2015.01336

**Conflict of Interest Statement:** The author declares that the research was conducted in the absence of any commercial or financial relationships that could be construed as a potential conflict of interest.

Copyright © 2016 Anonye. This is an open-access article distributed under the terms of the Creative Commons Attribution License (CC BY). The use, distribution or reproduction in other forums is permitted, provided the original author(s) or licensor are credited and that the original publication in this journal is cited, in accordance with accepted academic practice. No use, distribution or reproduction is permitted which does not comply with these terms.