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## iLIR database: A web resource for LIR motif-containing proteins in eukaryotes

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### ABSTRACT

Atg8-family proteins are the best-studied proteins of the core autophagic machinery. They are essential for the elongation and closure of the phagophore into a proper autophagosome. Moreover, Atg8-family proteins are associated with the phagophore from the initiation of the autophagic process to, or just prior to, the fusion between autophagosomes with lysosomes. In addition to their implication in autophagosome biogenesis, they are crucial for selective autophagy through their ability to interact with selective autophagy receptor proteins necessary for the specific targeting of substrates for autophagic degradation. In the past few years it has been revealed that Atg8-interacting proteins include not only receptors but also components of the core autophagic machinery, proteins associated with vesicles and their transport, and specific proteins that are selectively degraded by autophagy. Atg8-interacting proteins contain a short linear LC3-interacting region/LC3 recognition sequence/Atg8-interacting motif (LIR/LRS/AIM) motif which is responsible for their interaction with Atg8-family proteins. These proteins are referred to as LIR-containing proteins (LIRCPs). So far, many experimental efforts have been carried out to identify new LIRCPs, leading to the characterization of some of them in the past 10 years. Given the need for the identification of LIRCPs in various organisms, we developed the iLIR database (<https://ilir.warwick.ac.uk>) as a freely available web resource, listing all the putative canonical LIRCPs identified in silico in the proteomes of 8 model organisms using the iLIR server, combined with a Gene Ontology (GO) term analysis. Additionally, a curated text-mining analysis of the literature permitted us to identify novel putative LIRCPs in mammals that have not previously been associated with autophagy.

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### Introduction

Autophagy is a cellular catabolic process allowing for the degradation of numerous cytoplasmic components in a controlled and specific manner through the action of protein receptors that interact with Atg8/LC3/GABARAP-family proteins (hereafter refers as ‘Atg8-family proteins’).<sup>1</sup>

The term selective autophagy has been coined to refer to the targeted degradation of organelles (mitophagy, reticulophagy or pexophagy),<sup>2–4</sup> bacteria and viruses (xenophagy),<sup>5</sup> ribosomes (ribophagy),<sup>3</sup> lipid droplets (lipophagy)<sup>6</sup> and protein aggregates (aggrephagy).<sup>7</sup> Due to the large variety of substrates, selective autophagy employs various receptors able to recognize and tether specific substrates to phagophores.

Various studies pointed out that the interaction between receptors and Atg8-family proteins is mediated by an LC3-interacting region (LIR), also known as LC3 recognition sequence (LRS) or Atg8-interacting motif (AIM).<sup>8–15</sup> Thus, the presence of a LIR appears as a hallmark of the Atg8-interacting proteins.

The LIR corresponds to the shortest sequence required for the interaction with an Atg8-family protein. Previously described as the WxxL motif (where x can be any amino acid),

we and others recently extended this sequence to 6 amino acids based on the multiple alignment of LIR sequences from proteins described to interact in a LIR-dependent manner with Atg8-proteins.<sup>10,16,17</sup> Based on the in silico analysis of experimentally verified functional LIR motifs, we redefined the sequence of the LIR motif. The resulting consensus sequence—referred to as the xLIR motif—is (ADEFGLPRSK)(DEGMSTV)(WFY)(DEILQTV)(ADEFHIKLMPTV)(ILV), where the residues marked in bold (positions 3 and 6) correspond to the most crucial residues for the interaction with Atg8-family proteins. An xLIR motif overlapping a region with the potential to transit from a disordered to an ordered state provides a reliable candidate for a functional binding motif.<sup>10,17,18</sup>

In addition to selective autophagy receptors, Atg8-family proteins can bind a variety of proteins in an LIR-dependent manner. Indeed, many LIR motif-containing proteins (LIRCPs) are required for the formation of the autophagosome,<sup>16,19–22</sup> or vesicular transport,<sup>23,24</sup> or they are proteins that are directly targeted to the phagophore for autophagic clearance.<sup>25–27</sup>

It is worth mentioning that LIR motif-independent modes of interaction with Atg8-family proteins have also been reported

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both in selective autophagy receptors and in other autophagy-related proteins.<sup>28</sup>

In this report, we describe the use of the iLIR server<sup>17</sup> combined with a Gene Ontology (GO) term analysis to sort the genes from 8 model organisms (*Arabidopsis thaliana*, *Caenorhabditis elegans*, *Danio rerio*, *Gallus gallus*, *Homo sapiens*, *Mus musculus*, *Rattus norvegicus* and *Saccharomyces cerevisiae*) encoding proteins containing at least one xLIR motif inside an intrinsically disordered region. The data have been collected in the iLIR database (<https://ilir.warwick.ac.uk>), with the aim to provide a useful resource to researchers interested in studying the Atg8-family proteins interactome. Additionally, a curated text-mining analysis of the literature permitted us to sort human and mouse proteins known to be a part of the Atg8-family proteins interactome or to be involved in pathways linked to autophagy, and also to identify novel putative LICRPs that have not been associated with autophagy previously.

## Results and discussion

### Content of the iLIR database

The iLIR database is a web resource freely available at <https://ilir.warwick.ac.uk>. The website has been designed to give the user an easy way to browse available data and perform BLAST-based searches using a protein sequence of interest against part or all the sequences available in the database for proteins containing a similar xLIR motif. The website also provides hyperlinks to the UniProt database for each entry and the possibility to download the data.

Within the iLIR database different functionalities are organized under specific menus. The ‘LIRCPs’ menu gives

access to the full list of putative LIRCPs listed in the database for the different model organisms analyzed. For a specific organism, data are presented in a table containing the following information for each entry: (i) the UniProtKB accession of the protein, (ii) the position, sequence and position-specific scoring matrix score of the xLIR,<sup>17</sup> (iii) similar LIR motif in experimentally characterized LIRCPs (if any), (iv) the name of the protein, and (v) the UniProt derived GO terms associated with the molecular function, biological process and cellular component classes. The full table of data can be downloaded as an Excel file (Fig. 1).

The ‘Search’ menu offers the user to screen their sequence of interest for the presence of LIR (xLIR and WxxL) motifs using the iLIR server as described elsewhere.<sup>17</sup> In addition, the user has the possibility to search in the database using specific keywords: gene name, protein description or UniProt identifier. The user may also look directly for the presence of similar proteins with the ‘BLAST’ page using PSI-BLAST.<sup>29</sup> The search can be run against Swiss-Prot and TrEMBL entries from the UniProt database (a total of 276,499 FASTA sequences). The results page shows pattern positions in the query sequence and the corresponding matching positions in the subject sequences from the database along with the alignments between them. Red asterisks match the position of the conserved xLIR motif in the subject sequences. Subject sequences matched are named by their UniProtKB accession number and a link permits the redirection to the UniProtKB page for each entry (Fig. 2).

Finally, the ‘GO Annotation’ menu provides pre-computed information relative to the GO terms distribution for the LIRCPs identified for each organism. Three types of analyses are available: (i) The ‘GO Slim’ submenu directs users to a list of reduced GO terms and their abundance for each category in

**iLIR Autophagy Database**

Home LIRCPs BLAST GO Annotation Search Bibliography Help

LIR-containing proteins in *Homo sapiens* >> EXPORT DATA

<< prev 1 2 3 4 5 6 7 ... 121 122 next >>

S.No.	Uniprot ID	Start	End	LIR Sequence	PSSM Score	Similar LIR Motifs	Protein Description	Molecular Function	Biological Process	Cellular Component
1	<a href="#">A0JP02</a>	622	627	ASYVTL	13		PLEKHA5 protein			
2	<a href="#">A0M8Q6</a>	69	74	SSYLSL	12		Ig lambda-7 chain C region	antigen binding (GO:0003823)		extracellular region (GO:0005576)
3	<a href="#">A1A4T8</a>	53	58	FVFLPL	7		Uncharacterized protein C14orf182			
4	<a href="#">A1A4T8-2</a>	53	58	FVFLPL	7		Isoform 2 of Uncharacterized protein C14orf182			
5	<a href="#">A1L443</a>	23	28	SVFTAL	7		NUT family member 2F			
6	<a href="#">A1X283</a>	506	511	AGYEEI	13		SH3 and PX domain-containing protein 2B	protein binding (GO:0005515)		cell projection (GO:0042995)
7	<a href="#">A1XBS5</a>	185	190	SEFITI	13		Protein FAM92A1			
8	<a href="#">A1Z1Q3</a>	32	37	RDYIPL	16		O-acetyl-ADP-ribose deacetylase MACROD2	deacetylase activity (GO:0019213)		nucleus (GO:0005634)
9	<a href="#">A1Z1Q3-2</a>	32	37	RDYIPL	16		Isoform 2 of O-acetyl-ADP-ribose deacetylase MACROD2			
10	<a href="#">A2A2V2</a>	67	72	PVYVPV	7		RNA-binding protein 34 (Fragment)	nucleic acid binding (GO:0003676); nucleotide binding (GO:0000166)		
11	<a href="#">A2A312</a>	20	25	SSYQLL	12		Forkhead box protein P4 (Fragment)			
12	<a href="#">A2A312-2</a>	20	25	SSYQLL	12		Brain-specific angiogenesis			

**Figure 1.** Screenshot of an iLIR database data page. In the ‘LIRCPs’ menu, the user can access the full data available in the database for each model organism. The data are arranged in a table giving various information for each entry, such as the Uniprot Accession ID and protein name, the position and sequence of the xLIR as well as the position-specific scoring matrix (PSSM) score and the similarity of other validated LIR motifs. The data can be downloaded directly.

```

Query= sp|Q13501|SQSTM_HUMAN Sequestosome-1 OS=Homo sapiens GN=SQSTM1 PE=1
SV=1

Length=440
1 occurrence(s) of pattern:
[AEFGHILPRSK][DEGMSTV][FWY][DEILQTV][AEFHIKLMPTV][ILV] at position(s) 336 of query sequence
pattern probability=0.00135645

Sequences producing significant alignments:

Score      E
(Bits)     Value

sp|Q13501|SQSTM_HUMAN Sequestosome-1 OS=Homo sapiens GN=SQSTM1 P... 900 0.0
sp|Q64337|SQSTM_MOUSE Sequestosome-1 OS=Mus musculus GN=Sqstm1 P... 819 0.0
sp|O08623|SQSTM_RAT Sequestosome-1 OS=Rattus norvegicus GN=Sqstm... 807 0.0
tr|F1NA86|F1NA86_CHICK Uncharacterized protein OS=Gallus gallus ... 493 3e-144
tr|F1Q5Z8|F1Q5Z8_DANRE Sequestosome 1 OS=Danio rerio GN=sqstm1 P... 362 1e-104
tr|F1R4V2|F1R4V2_DANRE Uncharacterized protein OS=Danio rerio GN... 244 4e-69
tr|D3ZRI4|D3ZRI4_RAT Iroquois related homeobox 4 (Drosophila) (P... 24.3 0.008

Significant alignments for pattern occurrence 1 at position 336

>sp|Q13501|SQSTM_HUMAN Sequestosome-1 OS=Homo sapiens GN=SQSTM1 PE=1 SV=1
Length=440

Score = 900 bits (2334), Expect = 0.0
Identities = 440/440 (100%), Positives = 440/440 (100%), Gaps = 0/440 (0%)

Query 1 MASLTVKAYLLGKEDAAREIRRFSFCCSPEPEAEAEAAAAGPGPCERLLSRVAALFPALRP 60
Sbjct 1 MASLTVKAYLLGKEDAAREIRRFSFCCSPEPEAEAEAAAAGPGPCERLLSRVAALFPALRP 60

Query 61 GGFQAHYRDEDGDLVAFSSDEELTMAMSYVKDDIFRIYIKEKKECRRDHRPPCAQEAPRN 120
Sbjct 61 GGFQAHYRDEDGDLVAFSSDEELTMAMSYVKDDIFRIYIKEKKECRRDHRPPCAQEAPRN 120

Query 121 MVHPNVICDGCNGPVVVGTRYKCSVCPDYDLCSVCEGKGLHRGHTKLAFFSPFGHLSGFS 180
Sbjct 121 MVHPNVICDGCNGPVVVGTRYKCSVCPDYDLCSVCEGKGLHRGHTKLAFFSPFGHLSGFS 180

Query 181 HSRWLRKVKHGHFGWPGWEMGPPGNWSRPPRAGEARPGPTAESASGPSSEDPVSNFLKNV 240
Sbjct 181 HSRWLRKVKHGHFGWPGWEMGPPGNWSRPPRAGEARPGPTAESASGPSSEDPVSNFLKNV 240

Query 241 GESVAAALSPLGIEVDIDVEHGGKRSRLTPVSPESSTEEKSSSQPSSCCSDPSKPGGNV 300
Sbjct 241 GESVAAALSPLGIEVDIDVEHGGKRSRLTPVSPESSTEEKSSSQPSSCCSDPSKPGGNV 300

pattern
Query 301 EGATQSLAEQMRKIALESEGRPEEQMESDNCSSGGDDWTHLSSKEVDPSTGELQSLQMPE 360
Sbjct 301 EGATQSLAEQMRKIALESEGRPEEQMESDNCSSGGDDWTHLSSKEVDPSTGELQSLQMPE 360

Query 361 SEGPSLSDPSQEGPTGLKEAALYPHLPPEADPRLIESLSQMLSMGFSDEGGWLRLLQTK 420
Sbjct 361 SEGPSLSDPSQEGPTGLKEAALYPHLPPEADPRLIESLSQMLSMGFSDEGGWLRLLQTK 420

Query 421 NYDIGAALDTIQYSKHPPPL 440
Sbjct 421 NYDIGAALDTIQYSKHPPPL 440

>sp|Q64337|SQSTM_MOUSE Sequestosome-1 OS=Mus musculus GN=Sqstm1 PE=1 SV=1

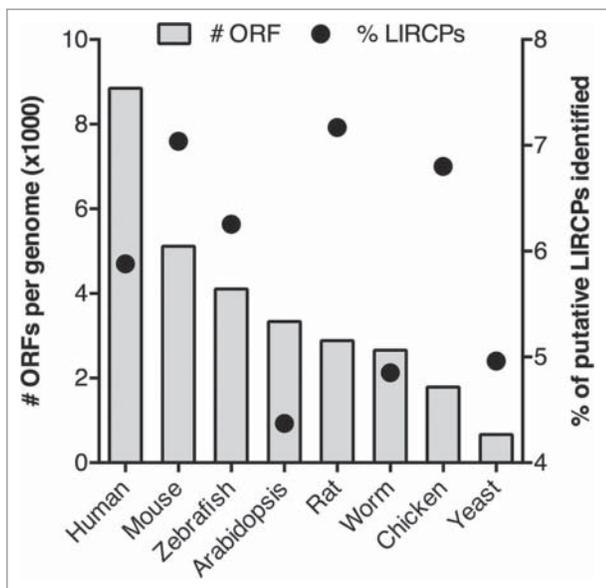
```

**Figure 2.** Screenshot of the iLIR database BLAST results page. Using the 'BLAST' menu, the user has the possibility to blast the sequence of interest against the sequences for one or all organisms available in the database in order to identify similar putative LIRCPs. The results page gives the list of similar sequences and the position of the putative LIR motif is indicated with red asterisks. The 'sp' and 'tr' preceding the FASTA header of the sequences producing a significant alignment refer to UniProtKG/Swiss-Prot (reviewed and manually annotated sequences) and UniProtKG/TrEMBL (unreviewed, automatically annotated sequences from large-scale screens), respectively.

a specific organism. The user can sort the entries based on their counts or adjusted p-value. (ii) The 'Distribution' submenu directs users to a bar chart view of the GO terms distribution for each organism. (ii) The 'Enrichment' submenu permits the visualization of the proportion of entries for each GO term for the LIRCPs for any pair of species available in the iLIR database (Fig. S1).

### **Prediction of the LIR-containing proteins (LIRCP) in the proteome of model organisms**

Using iLIR, a computational approach for predicting LC3-interaction regions in proteins,<sup>17</sup> we identified putative LIRCPs from 8 model organisms (see Methods for details). We found that the proportion of putative LIRCPs varies between 4% to



**Figure 3.** Representation of the number of ORFs (bar chart, plotted on the left axis) and percentage of putative LIRCPs identified (dot chart, plotted on the right axis) for each of the model organisms analyzed.

7% of the total ORFs for each organism but we observed no correlation between the proportion of LIRCPs and the size of the proteome (number of ORFs) (Fig. 3 and Table 1).

### Text-mining analysis for the identification of novel LIRCPs in mammals

In order to further investigate novel putative LIRCPs in mammals, we first concentrated on the human and mouse proteomes. Our batch analysis led to the identification of 6087 and 4218 entries, respectively. Consecutively to the application of the statistical significance for each GO slim category for these organisms, we decided to eliminate the entries sorted as ‘non significant’ (adjusted p-value > 0.1) from the rest of the analysis. This procedure permitted us to sort a total of 1766 and 1976 entries for the human and mouse proteome, respectively, with a low to high significance level ( $p\text{-adj} \leq 0.1$ ). We made use of these significant hits for further analysis.

Previous studies have identified and described 31 proteins encoded by the human, yeast and *Arabidopsis thaliana* genomes involved in autophagy through their interaction with at least one protein belonging to the Atg8-family and containing a functional, verified LIR motif.<sup>16,28,30</sup> However, the LIR motifs of a few of these proteins are not contained within an

intrinsically disordered region such as human ATG4B or yeast Atg3 and Atg19.<sup>17</sup> From the 31 verified LIRCPs, all 21 proteins with a LIR motif within an anchor region have been successfully identified in our computational analysis, thus validating the sorting procedure (these proteins constitute the group ‘A’ in the rest of the text) (Table S1).

From these proteins, we extracted their associated GO slim categories for the 3 GO classes (Molecular Function, Biological Process and Cellular Component). Totally, 26 different GO terms were obtained (6 for the Molecular Function class, 8 for the Biological Process class and 12 for the Cellular Component class) (Fig. 4 and Table S1). We noticed that only 4 of these proteins have been assigned to the GO term ‘GO:0006914|autophagy’ as a Biological Process; other proteins have been assigned to GO terms that can be related to autophagy such as GO:0005739|mitochondrion, GO:0030904|retromer complex (Cellular Component), GO:0006810|transport (Biological Process), GO:0005515|protein binding and GO:0042277|peptide binding (Molecular Function). Additionally, various GO terms not directly related to autophagy have been pinpointed such as GO:0005634|nucleus, GO:0005576|extracellular region, GO:0009986|cell surface, GO:0006457|protein folding, GO:0007049|cell cycle, GO:0004871|signal transducer activity or GO:0042562|hormone binding. This suggests that many proteins whose original function is not related to autophagy might interact with Atg8-family proteins in a way that remains unknown. In order to test this assumption, we decided to screen all the putative LIRCPs with a significant adjusted p-value (sorted as previously described) for the human and mouse proteomes, which are associated with at least one of the 26 GO terms correlated with the 21 experimentally validated human and yeast LIRCPs. Over 1,000 entries have thus been filtered. A manually curated search of these entries using PubMed, permitted us to sort 18 proteins already described to interact with an Atg8-family protein, irrespective of further evidence of a direct interaction (referred to hereafter as group ‘B’, Table S2). Three of these proteins—GPM1/AGS3,<sup>31,32</sup> NCOA4<sup>33</sup> and MAPK8IP1/JIP1<sup>34</sup>—had been shown to interact directly (i.e., through in vitro studies) with some members of the Atg8-family. The 15 remaining proteins—PICALM,<sup>35</sup> PCMI,<sup>36</sup> STAT1,<sup>37,38</sup> UBQLN1 and UBQLN2,<sup>39,40</sup> PEG3,<sup>41,42</sup> HTT,<sup>43</sup> SYNPO2,<sup>44</sup> UBR4,<sup>45,46</sup> MAP1S,<sup>47</sup> BCL10,<sup>48</sup> OFD1,<sup>36</sup> FNIP2,<sup>49</sup> APC<sup>50</sup> and CSPG4<sup>50</sup>—have been identified to function in complexes containing Atg8-family proteins *in cellulo* by co-immunoprecipitation and/or colocalization experiments (Table S2). In line with the functions of the LIRCPs containing experimentally verified LIR motifs (Table S1), it appears that

**Table 1.** Summary of the numbers of putative LIRCPs and LIR identified for the 8 model organisms studied.

		# ORF	# LIRCPs	LIR motifs	% LIRCPs	Ratio #LIRs:LIRCP
Human	<i>Homo sapiens</i>	88479	5204	6087	5.88	1.17
Mouse	<i>Mus musculus</i>	51130	3599	4218	7.04	1.17
Zebrafish	<i>Danio rerio</i>	41102	2571	3007	6.26	1.17
Arabidopsis	<i>Arabidopsis thaliana</i>	33350	1458	1592	4.37	1.09
Rat	<i>Rattus norvegicus</i>	28849	2068	2425	7.17	1.17
Worm	<i>Caenorhabditis elegans</i>	26595	1290	1526	4.85	1.18
Chicken	<i>Gallus gallus</i>	17864	1215	1400	6.80	1.15
Yeast	<i>Saccharomyces cerevisiae</i>	6693	332	363	4.96	1.09

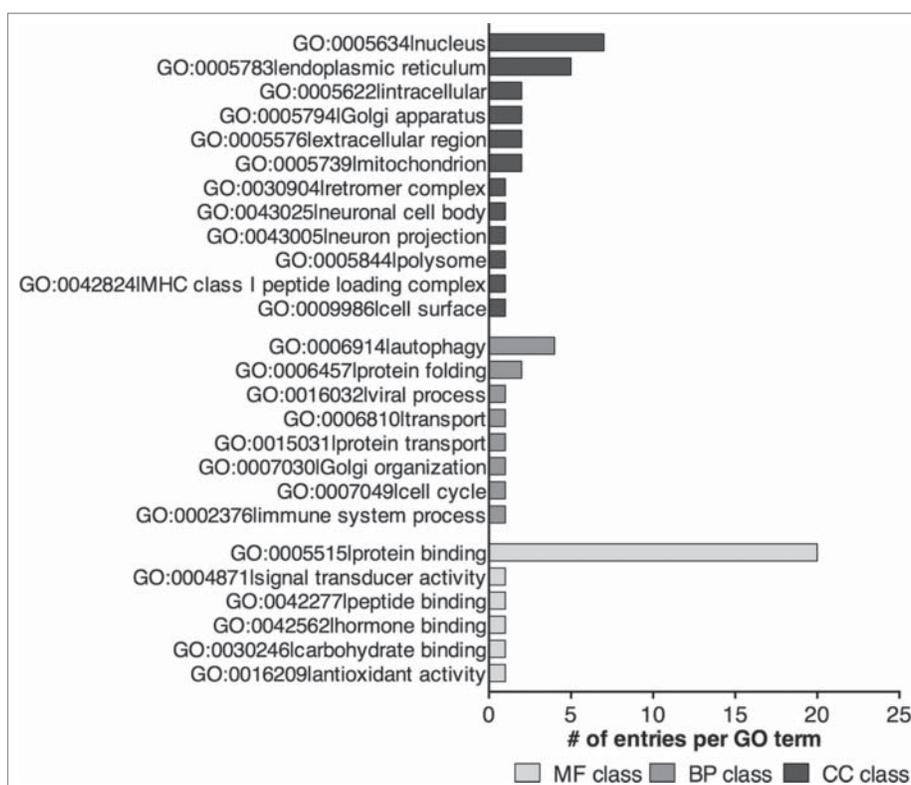


Figure 4. Distribution of the GO terms of the 21 human proteins listed in Kalvari et al. which have a verified xLIR in an intrinsically disordered region (see also Table S1). MF, Molecular Function; BP, Biological Process; CC, Cellular Component.

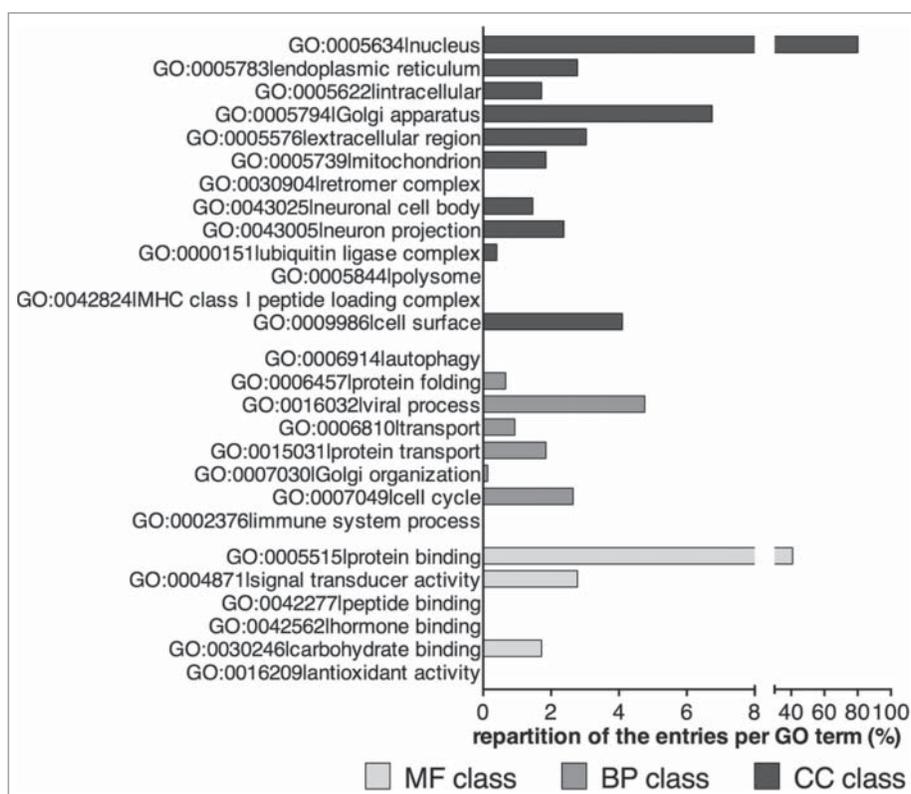


Figure 5. Distribution of the GO terms of the 756 human entries that have not been linked to autophagy-associated processes (see also Table S4). MF, Molecular Function; BP, Biological Process; CC, Cellular Component.

the proteins interacting with Atg8-family members we sorted can be related to the autophagy process in various ways. Some of these Atg8-interacting proteins are selective autophagy receptors for the targeting of specific cargos (e.g., NCOA4, PIC-ALM, PCMI, STAT1),<sup>33,35,37,38</sup> whereas others are degraded themselves by autophagy (e.g., BCL10, OFD1).<sup>36,48</sup> Yet some others are implicated in the regulation of the autophagic process (e.g., GPSM1/AGS3, MAPK8IP1/JIP1, UBQLN, PEG3, HTT, SYNPO2, UBR4, MAP1S, FNIP2) (Table S2).<sup>32,34,39-47,49</sup>

In addition, our text-mining analysis permitted us to sort 256 supplementary entries corresponding to proteins that have been demonstrated to be involved in the regulation of autophagy, the degradation of specific substrates, or to be themselves degraded by autophagy without any evidence of interaction with Atg8-family proteins (referred to hereafter as group 'C', Table S3). These proteins have been described to take part in a broad range of processes related to autophagy, such as immunity (NFKBIA/I $\kappa$ B $\alpha$ ,<sup>51</sup> IRF1 [interferon regulatory factor 1],<sup>52,53</sup> PPP1R13L/iASPP,<sup>54</sup> EIF2AK2/PKR,<sup>55</sup> RELA/NF- $\kappa$ B-p65<sup>56,57</sup>) or oncogenesis (BRCA1,<sup>58,59</sup> MYC,<sup>60,61</sup> RB1 [retinoblastoma 1],<sup>62,63</sup> TSC2/Tuberin,<sup>64</sup> FOXO1,<sup>65</sup> XIAP<sup>66,67</sup>). A few posttranslational modification enzymes have also been identified, such as 2 ubiquitin ligases (HERC1<sup>68,69</sup> and XIAP<sup>66,67</sup>), 4 kinases (PKD2/polycystin 2,<sup>70,71</sup> MARK4,<sup>72</sup> SIK2,<sup>73</sup> CAMKK2/CaMKK $\beta$ <sup>74</sup>), one deacetylase (HDAC4<sup>75</sup>), one methyltransferase (EHMT2/G9a<sup>76,77</sup>) and one phosphatase (PTPN13/PTPL1<sup>78</sup>) (Table S3).

Finally, we sorted proteins that have not been shown to be linked to autophagy or associated pathways, totaling for the human proteome 756 entries sharing their GO terms with the 21 human and yeast proteins that contain experimentally verified LIR motifs in an intrinsically disordered region (refers hereafter as group 'D', Table S4). The most represented GO terms are GO:0005634|nucleus (80.16%) for the Cellular Component class, GO:0005515|protein binding (40.87%) for the Molecular Function class and GO:0016032|viral process (4.76%) for the Biological Process class (Fig. 5). This observation suggest that these proteins are promising candidates for further investigation.

## Conclusion

Autophagy is a vital catabolic process for the maintenance of cell and tissue homeostasis by the selective degradation and recycling of macromolecules and organelles. In recent years, great efforts have been made for the identification and characterization of new receptors for selective autophagy, leading to the discovery of the LC3-interacting region.<sup>8-10</sup> Additional studies showed that LIR-containing proteins (LIRCPs) participate in a broad range of autophagic functions such as the selective targeting of cargo for degradation, the initiation and maturation of the autophagosome or vesicular transport.<sup>79</sup> Given the need for the identification of novel LIRCPs, we used the iLIR server to generate the iLIR database, a comprehensive bioinformatics resource for all the putative LIRCPs identified from the proteome of 8 model organisms. Our comprehensive manual literature analysis of human and mouse proteomes shows that our database includes already experimentally validated LIRCPs and novel putative functional LIRCPs.

Of course, there are some limitations to the iLIR database. At the moment, the iLIR server is not able to predict the non-canonical LIR motifs such as the one allowing for the interaction between CALCOCO2/NDP52 and LC3C.<sup>80</sup> Therefore the iLIR database cannot currently offer the list of unconventional LIRCPs.

In summary, we anticipate that the iLIR database will help autophagy researchers to test their candidates of interest, and elucidate the full set of LIRCPs in eukaryotes.

## Methods

### *Proteomes of model organisms and prediction of the LIR-containing proteins (LIRCPs)*

We selected 8 model organisms: *Arabidopsis thaliana*, *Caenorhabditis elegans*, *Danio rerio*, *Gallus gallus*, *Homo sapiens*, *Mus musculus*, *Rattus norvegicus* and *Saccharomyces cerevisiae*. The protein sequences encoding the complete genomes of these model organisms were obtained from the UniProt database (UniProt.org, (2014). *UniProt*. [online] Available at: <http://www.uniprot.org/> [Accessed 06 February 2014]). A stand-alone version of iLIR was employed to process the data in batch mode and predict LIRCPs based on the presence of at least one xLIR within an intrinsically disordered region.

### *Gene Ontology (GO) enrichment analysis*

The GO enrichment analysis was performed by downloading the ID (identifiers) mapping data for each organism from UniProt. These data contains cross-references for a given UniProt identifier with mappings to multiple databases such as Entrez-Gene, RefSeq, GI, PDB, GO, PIR, NCBI-taxon, UniGene etc. each recorded as an identifier of the respective database. We also downloaded the Gene Ontology Protein Information Resource slim generic categories from the online GO database (Geneontology.org, (2014). *GO Database*. [online] Available at: [http://www.geneontology.org/ontology/subsets/goslim\\_generic.obo](http://www.geneontology.org/ontology/subsets/goslim_generic.obo) [Accessed 19 June 2014].).

Using the mapping, GO slim and UniProt files together with the list of LIRCPs for a model organism, we generated GO class distribution files with counts of proteins having a particular GO slim category. One distribution file for each GO top level hierarchy (i.e., Biological Process, Cellular Component and Molecular Function) has been generated.

We assessed the statistical significance of each GO slim category of the model organisms using a hypergeometric test, employed through the Perl module Math::Pari (Search.cpan.org, (2014). Math-Pari-2.010808 Retrieved from: <http://search.cpan.org/CPAN/authors/id/I/IL/ILYAZ/modules/Math-Pari-2.010808.zip>.) based on the following criteria:

- Number of proteins assigned to a particular GO slim category in the model organism (n)
- Number of putative LIRCPs assigned to the same GO slim category in the model organism (x)
- Total number of proteins in the model organism (N)
- Total number of putative LIRCPs in the model organism (k)

The formula used for predicting the probability using hypergeometric test (h) is given below:<sup>81</sup>

$$h(x; N, n, k) = \frac{[{}_k C_x][{}_{N-k} C_{n-x}]}{[{}_N C_n]}.$$

To control the false discovery rate, we have also generated p-adjusted values employing the Benjamini-Hochberg method from Perl's Statistics::Multtest module (Search.cpan.org, (2014). Statistics-Multtest-0.13. Retrieved from: <http://search.cpan.org/CPAN/authors/id/J/JO/JOKERGOO/Statistics-Multtest-0.13.tar.gz>). Following the hypergeometric test and false discovery rate correction, the GO distribution files were updated with p-value and p-adjusted values. Then, the GO slim categories data of model organisms was further classified based on different cut-offs for p-adjusted (p-adj) values as:

- (i) Highly significant, (p-adj <= 0.01)
- (ii) Significant (p-adj > 0.01 and P-adj <= 0.05)
- (iii) Low significance (p-adj > 0.05 and p-adj <= 0.1)
- (iv) Not significant (p-adj > 0.1)

### Web application

The iLIR database has been developed for making the list of putative LIRCPs from the complete proteome of selected model organisms available for researchers worldwide. This web resource is based on well-established web technologies, including HTML, CSS, JavaScript, PHP (v5.3.28), JpGraph (v3.5.0b1) (Jpgraph.net, (2014). v3.5.0b1 Retrieved from: <http://jpgraph.net/download/download.php?p=5>) and the Apache web server technologies to develop and serve the web application.

### Abbreviations

AIM	Atg8-interacting motif
GO	gene ontology
LIR	LC3-interacting region
LIRCP	LIR-containing protein
LRS	LC3 recognition sequence
MAP1LC3/LC3	microtubule associated protein 1 light chain 3
xLIR	extended LIR motif

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No potential conflicts of interest were disclosed.

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### References

- [1] Shpilka T, Weidberg H, Pietrokovski S, Elazar Z. Atg8: an autophagy-related ubiquitin-like protein family. *Genome Biol* 2011; 12:226; PMID:21867568; <http://dx.doi.org/10.1186/gb-2011-12-7-226>
- [2] Youle RJ, Narendra DP. Mechanisms of mitophagy. *Nat Rev Mol Cell Biol* 2011; 12:9-14; PMID:21179058; <http://dx.doi.org/10.1038/nrm3028>
- [3] Ceblollo E, Reggiori F, Kraft C. Reticulophagy and ribophagy: regulated degradation of protein production factories. *Int J Cell Biol* 2012; 2012:182834; PMID:22481944; <http://dx.doi.org/10.1155/2012/182834>
- [4] Dunn WA, Jr., Cregg JM, Kiel JA, van der Klei IJ, Oku M, Sakai Y, Sibirny AA, Stasyk OV, Veenhuis M. Pexophagy: the selective autophagy of peroxisomes. *Autophagy* 2005; 1:75-83; PMID:16874024; <http://dx.doi.org/10.4161/auto.1.2.1737>
- [5] Boyle KB, Randow F. The role of 'eat-me' signals and autophagy cargo receptors in innate immunity. *Curr Opin Microbiol* 2013; 16:339-48; PMID:23623150; <http://dx.doi.org/10.1016/j.mib.2013.03.010>
- [6] Weidberg H, Shvets E, Elazar Z. Lipophagy: selective catabolism designed for lipids. *Dev Cell* 2009; 16:628-30; PMID:19460339; <http://dx.doi.org/10.1016/j.devcel.2009.05.001>
- [7] Lamark T, Johansen T. Aggrephagy: selective disposal of protein aggregates by macroautophagy. *Int J Cell Biol* 2012; 2012:736905; PMID:22518139; <http://dx.doi.org/10.1155/2012/736905>
- [8] Pankiv S, Clausen TH, Lamark T, Brech A, Bruun JA, Overvatn A, Bjorkoy G, Johansen T. p62/SQSTM1 binds directly to Atg8/LC3 to facilitate degradation of ubiquitinated protein aggregates by autophagy. *J Biol Chem* 2007; 282:24131-45; PMID:17580304; <http://dx.doi.org/10.1074/jbc.M702824200>
- [9] Ichimura Y, Kumanomidou T, Sou YS, Mizushima T, Ezaki J, Ueno T, Kominami E, Yamane T, Tanaka K, Komatsu M. Structural basis for sorting mechanism of p62 in selective autophagy. *J Biol Chem* 2008; 283:22847-57; PMID:18524774; <http://dx.doi.org/10.1074/jbc.M802182200>
- [10] Noda NN, Ohsumi Y, Inagaki F. Atg8-family interacting motif crucial for selective autophagy. *FEBS letters* 2010; 584:1379-85; PMID:20083108; <http://dx.doi.org/10.1016/j.febslet.2010.01.018>
- [11] Kirkin V, Lamark T, Sou YS, Bjorkoy G, Nunn JL, Bruun JA, Shvets E, McEwan DG, Clausen TH, Wild P, et al. A role for NBR1 in autophagosomal degradation of ubiquitinated substrates. *Mol Cell* 2009; 33:505-16; PMID:19250911; <http://dx.doi.org/10.1016/j.molcel.2009.01.020>
- [12] Thurston TL, Ryzhakov G, Bloor S, von Muhlinen N, Randow F. The TBK1 adaptor and autophagy receptor NDP52 restricts the proliferation of ubiquitin-coated bacteria. *Nat Immunol* 2009; 10:1215-21; PMID:19820708; <http://dx.doi.org/10.1038/ni.1800>
- [13] Schwarten M, Mohrluder J, Ma P, Stoldt M, Thielmann Y, Stangler T, Hersch N, Hoffmann B, Merkel R, Willbold D. Nix directly binds to GABARAP: a possible crosstalk between apoptosis and autophagy. *Autophagy* 2009; 5:690-8; PMID:19363302; <http://dx.doi.org/10.4161/auto.5.5.8494>
- [14] Novak I, Kirkin V, McEwan DG, Zhang J, Wild P, Rozenknop A, Rogov V, Lohr F, Popovic D, Occhipinti A, et al. Nix is a selective autophagy receptor for mitochondrial clearance. *EMBO Rep* 2010; 11:45-51; PMID:20010802; <http://dx.doi.org/10.1038/embo.2009.256>
- [15] Wild P, Farhan H, McEwan DG, Wagner S, Rogov VV, Brady NR, Richter B, Korac J, Waidmann O, Choudhary C, et al. Phosphorylation of the autophagy receptor optineurin restricts Salmonella growth. *Science* 2011; 333:228-33; PMID:21617041; <http://dx.doi.org/10.1126/science.1205405>
- [16] Alemu EA, Lamark T, Torgersen KM, Birgisdottir AB, Larsen KB, Jain A, Olsvik H, Overvatn A, Kirkin V, Johansen T. ATG8 family proteins act as scaffolds for assembly of the ULK complex: sequence requirements for LC3-interacting region (LIR) motifs. *J Biol Chem* 2012; 287:39275-90; PMID:23043107; <http://dx.doi.org/10.1074/jbc.M112.378109>
- [17] Kalvari I, Tsompanis S, Mulakkal NC, Osgood R, Johansen T, Nezis IP, Promponas VJ. iLIR: A web resource for prediction of Atg8-family interacting proteins. *Autophagy* 2014; 10:913-25; PMID:24589857; <http://dx.doi.org/10.4161/auto.28260>
- [18] Popelka H, Klionsky DJ. Analysis of the native conformation of the LIR/AIM motif in the Atg8/LC3/GABARAP-binding proteins. *Autophagy* 2015; 11:2153-9; PMID:26565669; <http://dx.doi.org/10.1080/15548627.2015.1111503>
- [19] Kraft C, Kijanska M, Kalie E, Siergiejuk E, Lee SS, Semplicio G, Stofel I, Brezovich A, Verma M, Hansmann I, et al. Binding of the Atg1/ULK1 kinase to the ubiquitin-like protein Atg8 regulates autophagy. *EMBO J* 2012; 31:3691-703; PMID:22885598; <http://dx.doi.org/10.1038/emboj.2012.225>

- [20] Nakatogawa H, Ohbayashi S, Sakoh-Nakatogawa M, Kakuta S, Suzuki SW, Kirisako H, Kondo-Kakuta C, Noda NN, Yamamoto H, Ohsumi Y. The autophagy-related protein kinase Atg1 interacts with the ubiquitin-like protein Atg8 via the Atg8 family interacting motif to facilitate autophagosomal formation. *J Biol Chem* 2012; 287:28503-7; PMID:22778255; <http://dx.doi.org/10.1074/jbc.C112.387514>
- [21] Satoo K, Noda NN, Kumeta H, Fujioka Y, Mizushima N, Ohsumi Y, Inagaki F. The structure of Atg4B-LC3 complex reveals the mechanism of LC3 processing and delipidation during autophagy. *EMBO J* 2009; 28:1341-50; PMID:19322194; <http://dx.doi.org/10.1038/emboj.2009.80>
- [22] Yamaguchi M, Noda NN, Nakatogawa H, Kumeta H, Ohsumi Y, Inagaki F. Autophagy-related protein 8 (Atg8) family interacting motif in Atg3 mediates the Atg3-Atg8 interaction and is crucial for the cytoplasm-to-vacuole targeting pathway. *J Biol Chem* 2010; 285:29599-607; PMID:20615880; <http://dx.doi.org/10.1074/jbc.M110.113670>
- [23] Popovic D, Akutsu M, Novak I, Harper JW, Behrends C, Dikic I. Rab GTPase-activating proteins in autophagy: regulation of endocytic and autophagy pathways by direct binding to human ATG8 modifiers. *Mol Cell Biol* 2012; 32:1733-44; PMID:22354992; <http://dx.doi.org/10.1128/MCB.06717-11>
- [24] Pankiv S, Alemu EA, Brech A, Bruun JA, Lamark T, Overvatn A, Bjorkoy G, Johansen T. FYCO1 is a Rab7 effector that binds to LC3 and PI3P to mediate microtubule plus end-directed vesicle transport. *J Cell Biol* 2010; 188:253-69; PMID:20100911; <http://dx.doi.org/10.1083/jcb.200907015>
- [25] Gao C, Cao W, Bao L, Zuo W, Xie G, Cai T, Fu W, Zhang J, Wu W, Zhang X, et al. Autophagy negatively regulates Wnt signalling by promoting Dishevelled degradation. *Nat Cell Biol* 2010; 12:781-90; PMID:20639871; <http://dx.doi.org/10.1038/ncb2082>
- [26] Zhang Y, Wang F, Han L, Wu Y, Li S, Yang X, Wang Y, Ren F, Zhai Y, Wang D, et al. GABARAP1 negatively regulates Wnt/beta-catenin signaling by mediating Dvl2 degradation through the autophagy pathway. *Cell Physiol Biochem* 2011; 27:503-12; PMID:21691068; <http://dx.doi.org/10.1159/000329952>
- [27] Petherick KJ, Williams AC, Lane JD, Ordonez-Moran P, Huelsken J, Collard TJ, Smartt HJ, Batson J, Malik K, Paraskeva C, et al. Autolysosomal beta-catenin degradation regulates Wnt-autophagy-p62 crosstalk. *EMBO J* 2013; 32:1903-16; PMID:23736261; <http://dx.doi.org/10.1038/emboj.2013.123>
- [28] Behrends C, Sowa ME, Gygi SP, Harper JW. Network organization of the human autophagy system. *Nature* 2010; 466:68-76; PMID:20562859; <http://dx.doi.org/10.1038/nature09204>
- [29] Altschul SF, Madden TL, Schaffer AA, Zhang J, Zhang Z, Miller W, Lipman DJ. Gapped BLAST and PSI-BLAST: a new generation of protein database search programs. *Nucleic Acids Res* 1997; 25:3389-402; PMID:9254694; <http://dx.doi.org/10.1093/nar/25.17.3389>
- [30] Birgisdottir AB, Lamark T, Johansen T. The LIR motif - crucial for selective autophagy. *J Cell Sci* 2013; 126:3237-47; PMID:23908376
- [31] Garcia-Marcos M, Ear J, Farquhar MG, Ghosh P. A GDI (AGS3) and a GEF (GIV) regulate autophagy by balancing G protein activity and growth factor signals. *Mol Biol Cell* 2011; 22:673-86; PMID:21209316; <http://dx.doi.org/10.1091/mbc.E10-08-0738>
- [32] Pattingre S, De Vries L, Bauvy C, Chantret I, Cluzeaud F, Ogier-Denis E, Vandewalle A, Codogno P. The G-protein regulator AGS3 controls an early event during macroautophagy in human intestinal HT-29 cells. *J Biol Chem* 2003; 278:20995-1002; PMID:12642577; <http://dx.doi.org/10.1074/jbc.M300917200>
- [33] Mancias JD, Wang X, Gygi SP, Harper JW, Kimmelman AC. Quantitative proteomics identifies NCOA4 as the cargo receptor mediating ferritinophagy. *Nature* 2014; 509:105-9; PMID:24695223; <http://dx.doi.org/10.1038/nature13148>
- [34] Fu MM, Nirschl JJ, Holzbaur EL. LC3 binding to the scaffolding protein JIP1 regulates processive dynein-driven transport of autophagosomes. *Dev Cell* 2014; 29:577-90; PMID:24914561; <http://dx.doi.org/10.1016/j.devcel.2014.04.015>
- [35] Tian Y, Chang JC, Fan EY, Flajolet M, Greengard P. Adaptor complex AP2/PICALM, through interaction with LC3, targets Alzheimer's APP-CTF for terminal degradation via autophagy. *Proc Natl Acad Sci U S A* 2013; 110:17071-6; PMID:24067654; <http://dx.doi.org/10.1073/pnas.1315110110>
- [36] Tang Z, Lin MG, Stowe TR, Chen S, Zhu M, Stearns T, Franco B, Zhong Q. Autophagy promotes primary ciliogenesis by removing OFD1 from centriolar satellites. *Nature* 2013; 502:254-7; PMID:24089205; <http://dx.doi.org/10.1038/nature12606>
- [37] Bourke LT, Knight RA, Latchman DS, Stephanou A, McCormick J. Signal transducer and activator of transcription-1 localizes to the mitochondria and modulates mitophagy. *Jak-Stat* 2013; 2:e25666; PMID:24470977; <http://dx.doi.org/10.4161/jkst.25666>
- [38] Chang YP, Tsai CC, Huang WC, Wang CY, Chen CL, Lin YS, Kai JJ, Hsieh CY, Cheng YL, Choi PC, et al. Autophagy facilitates IFN-gamma-induced Jak2-STAT1 activation and cellular inflammation. *J Biol Chem* 2010; 285:28715-22; PMID:20592027; <http://dx.doi.org/10.1074/jbc.M110.133355>
- [39] N'Diaye EN, Kajihara KK, Hsieh I, Morisaki H, Debnath J, Brown EJ. PLIC proteins or ubiquilins regulate autophagy-dependent cell survival during nutrient starvation. *EMBO Rep* 2009; 10:173-9; PMID:19148225; <http://dx.doi.org/10.1038/embor.2008.238>
- [40] Rothenberg C, Srinivasan D, Mah L, Kaushik S, Peterhoff CM, Ugoilino J, Fang S, Cuervo AM, Nixon RA, Monteiro MJ. Ubiquilin functions in autophagy and is degraded by chaperone-mediated autophagy. *Hum Mol Genet* 2010; 19:3219-32; PMID:20529957; <http://dx.doi.org/10.1093/hmg/ddq231>
- [41] Buraschi S, Neill T, Goyal A, Poluzzi C, Smythies J, Owens RT, Schaefer L, Torres A, Iozzo RV. Decorin causes autophagy in endothelial cells via Peg3. *Proc Natl Acad Sci U S A* 2013; 110:E2582-91; PMID:23798385; <http://dx.doi.org/10.1073/pnas.1305732110>
- [42] Poluzzi C, Casulli J, Goyal A, Mercer TJ, Neill T, Iozzo RV. Endorepellin evokes autophagy in endothelial cells. *J Biol Chem* 2014; 289:16114-28; PMID:24737315; <http://dx.doi.org/10.1074/jbc.M114.556530>
- [43] Ochaba J, Lukacsovich T, Csikos G, Zheng S, Margulis J, Salazar L, Mao K, Lau AL, Yeung SY, Humbert S, et al. Potential function for the Huntingtin protein as a scaffold for selective autophagy. *Proc Natl Acad Sci U S A* 2014; PMID:25385587
- [44] Ulbricht A, Eppler FJ, Tapia VE, van der Ven PF, Hampe N, Hersch N, Vakeel P, Stadel D, Haas A, Saftig P, et al. Cellular mechanotransduction relies on tension-induced and chaperone-assisted autophagy. *Curr Biol* 2013; 23:430-5; PMID:23434281; <http://dx.doi.org/10.1016/j.cub.2013.01.064>
- [45] Tasaki T, Kim ST, Zakrzewska A, Lee BE, Kang MJ, Yoo YD, Chamolstad HJ, Hwang J, Soung NK, Sung KS, et al. UBR box N-recognin-4 (UBR4), an N-recognin of the N-end rule pathway, and its role in yolk sac vascular development and autophagy. *Proc Natl Acad Sci U S A* 2013; 110:3800-5; PMID:23431188; <http://dx.doi.org/10.1073/pnas.1217358110>
- [46] Yamano K, Youle RJ. PINK1 is degraded through the N-end rule pathway. *Autophagy* 2013; 9:1758-69; PMID:24121706; <http://dx.doi.org/10.4161/auto.24633>
- [47] Xie R, Nguyen S, McKeehan K, Wang F, McKeehan WL, Liu L. Microtubule-associated protein 1S (MAP1S) bridges autophagic components with microtubules and mitochondria to affect autophagosomal biogenesis and degradation. *J Biol Chem* 2011; 286:10367-77; PMID:21262964; <http://dx.doi.org/10.1074/jbc.M110.206532>
- [48] Paul S, Kashyap AK, Jia W, He YW, Schaefer BC. Selective autophagy of the adaptor protein Bcl10 modulates T cell receptor activation of NF-kappaB. *Immunity* 2012; 36:947-58; PMID:22658522; <http://dx.doi.org/10.1016/j.immuni.2012.04.008>
- [49] Dunlop EA, Seifan S, Claessens T, Behrends C, Kamps MA, Rozycka E, Kemp AJ, Nookala RK, Blenis J, Coull BJ, et al. FLCN, a novel autophagy component, interacts with GABARAP and is regulated by ULK1 phosphorylation. *Autophagy* 2014; 10:1749-60; PMID:25126726; <http://dx.doi.org/10.4161/auto.29640>
- [50] Sarkar C, Zhao Z, Aungst S, Sabirzhanov B, Faden AI, Lipinski MM. Impaired autophagy flux is associated with neuronal cell death after traumatic brain injury. *Autophagy* 2014; 10:2208-22; PMID:25484084; <http://dx.doi.org/10.4161/15548627.2014.981787>
- [51] Colleran A, Ryan A, O'Gorman A, Mureau C, Liprot C, Dockery P, Fearnhead H, Egan LJ. Autophagosomal I kappaB alpha degradation

- plays a role in the long term control of tumor necrosis factor-alpha-induced nuclear factor-kappaB (NF-kappaB) activity. *J Biol Chem* 2011; 286:22886-93; PMID:21454695; <http://dx.doi.org/10.1074/jbc.M110.199950>
- [52] Li P, Du Q, Cao Z, Guo Z, Evankovich J, Yan W, Chang Y, Shao L, Stolz DB, Tsung A, et al. Interferon-gamma induces autophagy with growth inhibition and cell death in human hepatocellular carcinoma (HCC) cells through interferon-regulatory factor-1 (IRF-1). *Cancer Lett* 2012; 314:213-22; PMID:22056812; <http://dx.doi.org/10.1016/j.canlet.2011.09.031>
- [53] Zhang L, Cardinal JS, Bahar R, Evankovich J, Huang H, Nace G, Biliari TR, Rosengart MR, Pan P, Tsung A. Interferon regulatory factor-1 regulates the autophagic response in LPS-stimulated macrophages through nitric oxide. *Mol Med* 2012; 18:201-8; PMID:22105605; <http://dx.doi.org/10.2119/molmed.2011.00094>
- [54] Chikh A, Sanza P, Raimondi C, Akinduro O, Warnes G, Chiorino G, Byrne C, Harwood CA, Bergamaschi D. iASPP is a novel autophagy inhibitor in keratinocytes. *J Cell Sci* 2014; 127:3079-93; PMID:24777476; <http://dx.doi.org/10.1242/jcs.144816>
- [55] Niso-Santano M, Shen S, Adjemian S, Malik SA, Marino G, Lachkar S, Senovilla L, Kepp O, Galluzzi L, Maiuri MC, et al. Direct interaction between STAT3 and EIF2AK2 controls fatty acid-induced autophagy. *Autophagy* 2013; 9:415-7; PMID:23221979; <http://dx.doi.org/10.4161/auto.22910>
- [56] Chang CP, Su YC, Hu CW, Lei HY. TLR2-dependent selective autophagy regulates NF-kappaB lysosomal degradation in hepatoma-derived M2 macrophage differentiation. *Cell Death Differ* 2013; 20:515-23; PMID:23175187; <http://dx.doi.org/10.1038/cdd.2012.146>
- [57] Ryu HJ, Kim JE, Yeo SI, Kang TC. p65/RelA-Ser529 NF-kappaB subunit phosphorylation induces autophagic astroglial death (Clasmatodendrosis) following status epilepticus. *Cell Mol Neurobiol* 2011; 31:1071-8; PMID:21598036; <http://dx.doi.org/10.1007/s10571-011-9706-1>
- [58] Esteve JM, Armengod ME, Knecht E. BRCA1 negatively regulates formation of autophagic vacuoles in MCF-7 breast cancer cells. *Exp Cell Res* 2010; 316:2618-29; PMID:20599945; <http://dx.doi.org/10.1016/j.yexcr.2010.06.019>
- [59] Fan S, Meng Q, Saha T, Sarkar FH, Rosen EM. Low concentrations of diindolylmethane, a metabolite of indole-3-carbinol, protect against oxidative stress in a BRCA1-dependent manner. *Cancer Res* 2009; 69:6083-91; PMID:19622773; <http://dx.doi.org/10.1158/0008-5472.CAN-08-3309>
- [60] Balakumaran BS, Porrello A, Hsu DS, Glover W, Foye A, Leung JY, Sullivan BA, Hahn WC, Loda M, Febbo PG. MYC activity mitigates response to rapamycin in prostate cancer through eukaryotic initiation factor 4E-binding protein 1-mediated inhibition of autophagy. *Cancer Res* 2009; 69:7803-10; PMID:19773438; <http://dx.doi.org/10.1158/0008-5472.CAN-09-0910>
- [61] Tsuneoka M, Umata T, Kimura H, Koda Y, Nakajima M, Kosai K, Takahashi T, Takahashi Y, Yamamoto A. c-myc induces autophagy in rat 3Y1 fibroblast cells. *Cell Struct Funct* 2003; 28:195-204; PMID:12951440; <http://dx.doi.org/10.1247/csf.28.195>
- [62] Jiang H, Martin V, Gomez-Manzano C, Johnson DG, Alonso M, White E, Xu J, McDonnell TJ, Shinjima N, Fueyo J. The RB-E2F1 pathway regulates autophagy. *Cancer Res* 2010; 70:7882-93; PMID:20807803; <http://dx.doi.org/10.1158/0008-5472.CAN-10-1604>
- [63] Biasoli D, Kahn SA, Cornelio TA, Furtado M, Campanati L, Chneiweiss H, Moura-Neto V, Borges HL. Retinoblastoma protein regulates the crosstalk between autophagy and apoptosis, and favors glioblastoma resistance to etoposide. *Cell Death Dis* 2013; 4:e767; PMID:23949216; <http://dx.doi.org/10.1038/cddis.2013.283>
- [64] Yu J, Parkhitko A, Henske EP. Autophagy: an 'Achilles' heel of tumorigenesis in TSC and LAM. *Autophagy* 2011; 7:1400-1; PMID:21997371; <http://dx.doi.org/10.4161/auto.7.11.17652>
- [65] Zhao Y, Yang J, Liao X, Liu X, Zhang H, Wang S, Wang D, Feng J, Yu L, Zhu WG. Cytosolic FoxO1 is essential for the induction of autophagy and tumour suppressor activity. *Nat Cell Biol* 2010; 12:665-75; PMID:20543840; <http://dx.doi.org/10.1038/ncb2069>
- [66] Huang X, Wu Z, Mei Y, Wu M. XIAP inhibits autophagy via XIAP-Mdm2-p53 signalling. *EMBO J* 2013; 32:2204-16; PMID:23749209; <http://dx.doi.org/10.1038/emboj.2013.133>
- [67] Lin F, Ghislat G, Luo S, Renna M, Siddiqi F, Rubinsztein DC. XIAP and cIAP1 amplifications induce Beclin 1-dependent autophagy through NFkappaB activation. *Hum Mol Genet* 2015; PMID: AMBIGUOUS
- [68] Chong-Kopera H, Inoki K, Li Y, Zhu T, Garcia-Gonzalo FR, Rosa JL, Guan KL. TSC1 stabilizes TSC2 by inhibiting the interaction between TSC2 and the HERC1 ubiquitin ligase. *J Biol Chem* 2006; 281:8313-6; PMID:16464865; <http://dx.doi.org/10.1074/jbc.C500451200>
- [69] Mashimo T, Hadjei O, Amair-Pinedo F, Tsurumi T, Langa F, Serikawa T, Sotelo C, Guenet JL, Rosa JL. Progressive Purkinje cell degeneration in tambaleante mutant mice is a consequence of a missense mutation in HERC1 E3 ubiquitin ligase. *PLoS Genet* 2009; 5:e1000784; PMID:20041218; <http://dx.doi.org/10.1371/journal.pgen.1000784>
- [70] Boletta A. Emerging evidence of a link between the polycystins and the mTOR pathways. *PathoGenetics* 2009; 2:6; PMID:19863783; <http://dx.doi.org/10.1186/1755-8417-2-6>
- [71] Cebotaru V, Cebotaru L, Kim H, Chiaravalli M, Boletta A, Qian F, Guggino WB. Polycystin-1 negatively regulates Polycystin-2 expression via the aggressive/autophagosome pathway. *J Biol Chem* 2014; 289:6404-14; PMID:24459142; <http://dx.doi.org/10.1074/jbc.M113.501205>
- [72] Li L, Guan KL. Microtubule-associated protein/microtubule affinity-regulating kinase 4 (MARK4) is a negative regulator of the mammalian target of rapamycin complex 1 (mTORC1). *J Biol Chem* 2013; 288:703-8; PMID:23184942; <http://dx.doi.org/10.1074/jbc.C112.396903>
- [73] Yang FC, Tan BC, Chen WH, Lin YH, Huang JY, Chang HY, Sun HY, Hsu PH, Liou GG, Shen J, et al. Reversible acetylation regulates salt-inducible kinase (SIK2) and its function in autophagy. *J Biol Chem* 2013; 288:6227-37; PMID:23322770; <http://dx.doi.org/10.1074/jbc.M112.431239>
- [74] Hoyer-Hansen M, Bastholm L, Szyniarowski P, Campanella M, Szabadkai G, Farkas T, Bianchi K, Fehrenbacher N, Elling F, Rizzuto R, et al. Control of macroautophagy by calcium, calmodulin-dependent kinase kinase-beta, and Bcl-2. *Mol Cell* 2007; 25:193-205; PMID:17244528; <http://dx.doi.org/10.1016/j.molcel.2006.12.009>
- [75] Kang ZH, Wang CY, Zhang WL, Zhang JT, Yuan CH, Zhao PW, Lin YY, Hong S, Li CY, Wang L. Histone deacetylase HDAC4 promotes gastric cancer SGC-7901 cells progression via p21 repression. *PLoS One* 2014; 9:e98894; PMID:24896240; <http://dx.doi.org/10.1371/journal.pone.0098894>
- [76] Artal-Martinez de Narvajás A, Gomez TS, Zhang JS, Mann AO, Taoda Y, Gorman JA, Herreros-Villanueva M, Gress TM, Ellenrieder V, Bujanda L, et al. Epigenetic regulation of autophagy by the methyltransferase G9a. *Mol Cell Biol* 2013; 33:3983-93; PMID:23918802; <http://dx.doi.org/10.1128/MCB.00813-13>
- [77] Kim Y, Kim YS, Kim DE, Lee JS, Song JH, Kim HG, Cho DH, Jeong SY, Jin DH, Jang SJ, et al. BIX-01294 induces autophagy-associated cell death via EHMT2/G9a dysfunction and intracellular reactive oxygen species production. *Autophagy* 2013; 9:2126-39; PMID:24322755; <http://dx.doi.org/10.4161/auto.26308>
- [78] Kuchay S, Duan S, Schenkein E, Peschiaroli A, Saraf A, Florens L, Washburn MP, Pagano M. FBXL2- and PTPL1-mediated degradation of p110-free p85beta regulatory subunit controls the PI(3)K signalling cascade. *Nat Cell Biol* 2013; 15:472-80; PMID:23604317; <http://dx.doi.org/10.1038/ncb2731>
- [79] Wild P, McEwan DG, Dikic I. The LC3 interactome at a glance. *J Cell Sci* 2014; 127:3-9; PMID:24345374; <http://dx.doi.org/10.1242/jcs.140426>
- [80] von Muhlinen N, Akutsu M, Ravenhill BJ, Foeglein A, Bloor S, Rutherford TJ, Freund SM, Komander D, Randow F. LC3C, bound selectively by a noncanonical LIR motif in NDP52, is required for antibacterial autophagy. *Mol Cell* 2012; 48:329-42; PMID:23022382; <http://dx.doi.org/10.1016/j.molcel.2012.08.024>
- [81] Stamatis DH. *Typical Sampling Techniques. Essential Statistical Concepts for the Quality Professional*. Boca Raton, FL: CRC Press, 2012; 147-8.