

Original citation:

Lidbury, Ian, Fraser, Tandra, Murphy, Andrew, Scanlan, David J. , Bending, Gary D., Jones, Alexandra M., Moore, Jonathan D., Goodall, Andrew, Tibbett, Mark, Hammond, John P. and Wellington, E. M. H.. (2017) The 'known' genetic potential for microbial communities to degrade organic phosphorus is reduced in low-pH soils. *MicrobiologyOpen* . e00474.

Permanent WRAP URL:

<http://wrap.warwick.ac.uk/88920>

Copyright and reuse:

The Warwick Research Archive Portal (WRAP) makes this work of researchers of the University of Warwick available open access under the following conditions.

This article is made available under the Creative Commons Attribution 4.0 International license (CC BY 4.0) and may be reused according to the conditions of the license. For more details see: <http://creativecommons.org/licenses/by/4.0/>




A note on versions:

The version presented in WRAP is the published version, or, version of record, and may be cited as it appears here.

For more information, please contact the WRAP Team at: wrap@warwick.ac.uk

ORIGINAL RESEARCH

The 'known' genetic potential for microbial communities to degrade organic phosphorus is reduced in low-pH soils

Ian D. E. A. Lidbury¹  | Tandra Fraser² | Andrew R. J. Murphy¹ | David J. Scanlan¹ | Gary D. Bending¹ | Alexandra M. E. Jones¹ | Jonathan D. Moore³  | Andrew Goodall² | Mark Tibbett² | John P. Hammond^{2,4}  | Elizabeth M. H. Wellington¹

¹School of Life Sciences, University of Warwick, Coventry, West Midlands, United Kingdom

²School of Agriculture, Policy, and Development, University of Reading, Whiteknights, United Kingdom

³The Genome Analysis Centre, Norwich Research Park, Norwich, United Kingdom

⁴Southern Cross Plant Science, Southern Cross University, Lismore, Australia

Correspondence

Ian D. E. A. Lidbury, School of Life Sciences, University of Warwick, Coventry, West Midlands, United Kingdom.
Email: i.lidbury@warwick.ac.uk

Funding information

Biotechnology and Biological Sciences Research Council (BBSRC), Grant/Award Number: BB/L026074/1

Abstract

In soil, bioavailable inorganic orthophosphate is found at low concentrations and thus limits biological growth. To overcome this phosphorus scarcity, plants and bacteria secrete numerous enzymes, namely acid and alkaline phosphatases, which cleave orthophosphate from various organic phosphorus substrates. Using profile hidden Markov modeling approaches, we investigated the abundance of various non specific phosphatases, both acid and alkaline, in metagenomes retrieved from soils with contrasting pH regimes. This analysis uncovered a marked reduction in the abundance and diversity of various alkaline phosphatases in low-pH soils that was not counterbalanced by an increase in acid phosphatases. Furthermore, it was also discovered that only half of the bacterial strains from different phyla deposited in the Integrated Microbial Genomes database harbor alkaline phosphatases. Taken together, our data suggests that these 'phosphatase lacking' isolates likely increase in low-pH soils and future research should ascertain how these bacteria overcome phosphorus scarcity.

KEYWORDS

Acid phosphatase, alkaline phosphatase, metagenomics, microbial community, microbial diversity, soil

1 | INTRODUCTION

In soil, total phosphorus (P) concentrations typically vary between 100 and 3,000 mg kg⁻¹ (Hedley et al. 1995; Mengel 1997), of which 30%–60% is in the form of complex organic P esters (White & Hammond, 2008). However, plants can only acquire simple orthophosphate (Pi), which is frequently found at low concentrations in soils (<10 μmol L⁻¹) (White & Hammond, 2008). Therefore, P is often a limiting nutrient for global crop production and subsequently inorganic rock phosphate is applied in large quantities with deleterious economic and environmental consequences (Vance, Uhde-Stone, & Allan, 2003; Cordell, Drangert, & White, 2009; White & Hammond, 2008, 2009).

Microorganisms can have a beneficial effect on crop production, partly through the liberation of unavailable P in the rhizosphere and surrounding soil (Rodríguez & Fraga, 1999). Various studies have revealed that upon depletion of extracellular Pi bacteria can undergo a regulatory and physiological response resulting in the secretion of various exoproteins involved in liberating and binding Pi (Hirani, Suzuki, Murata, Hayashi, & Eaton-Rye, 2001; Rittmann, Sorger-Herrmann, & Wendisch, 2005; Monds, Newell, Schwartzman, & O'Toole, 2006; Antelmann, Scharf, & Hecker, 2000; Lidbury et al., 2016). Through this process, numerous reports have revealed that various phosphatases play an important role in the bioavailability of P in soils (Nannipieri, Newell, Giagnoni, Landi, & Renella, 2011). Usually, the most heavily

This is an open access article under the terms of the Creative Commons Attribution License, which permits use, distribution and reproduction in any medium, provided the original work is properly cited.

© 2017 The Authors. *MicrobiologyOpen* published by John Wiley & Sons Ltd.

secreted exoenzymes are alkaline phosphatases (APases), which cleave Pi from a plethora of complex organic P monoesters and diesters. The latter compounds account for the bulk (up to 90%) of organic P in soils (Condrón, Turner, & Cade-Menun, 2005; Lidbury et al., 2016; Santos-Beneit, 2015; Zaheer, Morton, Proudfoot, Yakunin, & Finan, 2009). APases encompass a wide genetic diversity including PhoA (Roy, Ghosh, & Das, 1982), PhoX (Sebastian & Ammerman, 2009) and PhoD (Eder, Shi, Jensen, Yamane, & Hulett, 1996) types. APases are secreted through various mechanisms, including the twin-arginine translocation (TAT) pathway (Putker et al., 2013) or *sec* pathway (PhoA) (Angelini et al., 2001). Mutagenesis of *phoX* almost entirely abolishes *para*-nitrophenyl phosphate degradation capacity (Monds et al., 2006), the substrate most frequently used to determine in situ soil phosphatase activity. Similarly, both PhoD and PhoA have been shown to be highly active against *para*-nitrophenyl phosphate (Rodríguez et al., 2014; Yang & Metcalf, 2004). APases are thought to be partly responsible for the 'P-fertilisation' effect seen when inoculating soils with plant growth-promoting bacteria (PGPB) (Condrón et al., 2005; Rodríguez & Fraga, 1999).

2 | EXPERIMENTAL PROCEDURES

2.1 | Bioinformatic analyses

For each protein analyzed, between 50 and 80 sequences from a range of phylogenetically distinct soil bacteria were downloaded from the Integrated Microbial Genomes database (IMG/JGI) (<https://img.jgi.doe.gov/>). To identify these homologs, a combination of BLASTP and function searches (IMG search option) using the conserved PFAM domains for each protein were performed. Downloaded sequences were aligned using MUSCLE and HMM profiles were constructed using the *hmmbuild* function in HMMER (Eddy, 1998).

The eight metagenomes were downloaded from the EBI metagenomics portal under the project code ERP001068 (Title: Functional diversity of soil microbes across environmental gradients). The eight sample IDs for each metagenome are as follows: ERS078132, ERS078133, ERS078134, ERS078135, ERS078136, ERS078137, ERS078138, ERS078139 corresponding to CS1, CS179, CS864, CS922, CS78, CS251, CS511, CS1053 sites of the UK countryside survey, respectively. Information regarding the taxonomic profile of each site (based on the 16S rRNA gene) can be obtained directly from the EBI metagenomics portal. For each site, both FASTA files for the 'predicted CDS' and 'predicted CDS without annotation' were downloaded.

Each site was screened for the abundance of each functional gene using the *hmmsearch* option in HMMER and using *easel* to generate output files. For determining the abundance of a given functional gene at each site, the method used by Howard, Sun, Biers, and Moran (2008) was adapted. Briefly, the raw number of hits at each site was normalized to RecA by taking the ratio of the length (amino acid) of each given functional protein against the length (amino acid) of RecA. Four single copy housekeeping genes, RpoB, AtpB, GyrB, and SucD were also used. The mean number of counts retrieved for each enzyme

was used to determine the 'average genome equivalent' for each site. Next, the ratio of the number of hits related to phosphorus-scavenging enzymes against each genome equivalent was calculated to provide the number of bacteria containing a given enzyme at each site.

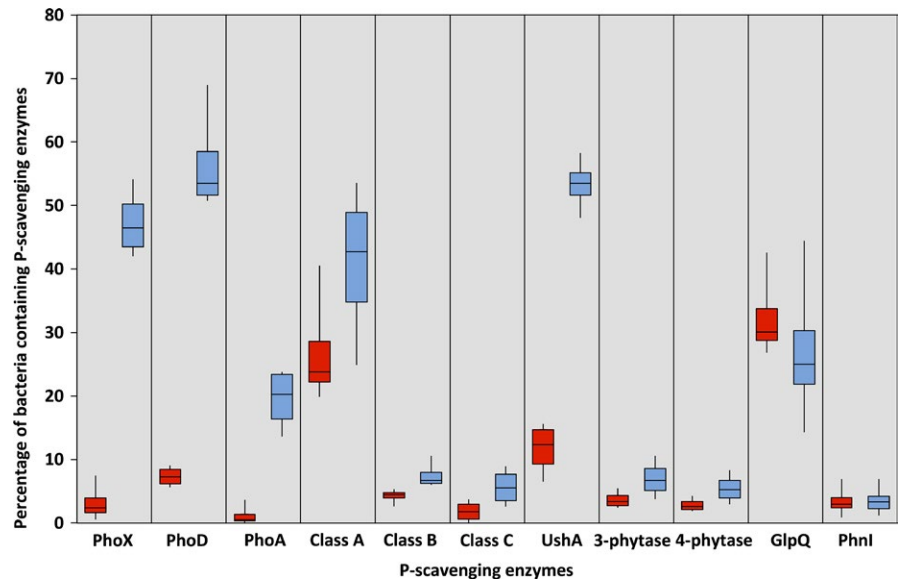
To assess the diversity of alkaline phosphatases and GyrB, manually curated databases were established for each gene by downloading all the known homologs present in the IMG/JGI database. A BLASTP search was performed using a relatively relaxed stringency (e^{-10}). To confirm the validity of the hits retrieved from *hmmsearches*, individual sequences retrieved by *hmmsearches* were used as queries in BLASTP searches using the *nr* database located on the National Centre for Biotechnology Information server (www.ncbi.nlm.nih.gov/BLAST.cgi). Higher taxonomic ranks were retrieved using the NCBI taxonomy tool. The data were visualized using the Krona tools package (Ondov et al. 2011).

3 | RESULTS AND DISCUSSION

To determine the 'genetic potential' of microbial communities to remineralize organic forms of P, we screened eight metagenomes (<https://www.ebi.ac.uk/metagenomics/>) from geographically distinct locations across the UK landscape (Griffiths et al., 2011). Four sites were considered low-pH soils (4.12–4.37), and four high-pH soils (8.04–8.46). Profile hidden Markov modeling (HMM) searches (Eddy, 1998) were conducted for each APase (PhoX, PhoD, PhoA), the nonspecific class A, class B, class C acid phosphatases (ACPases) (Gandhi & Chandra, 2012; Thaller, Berlutti, Schippa, Lombardi, & Rossolini, 1994), a number of other P-scavenging enzymes and several housekeeping genes (see Table S1). The number of bacteria containing P-scavenging genes was determined according to Howard et al. (2008). Based on 16S rRNA gene diversity, there were no significant changes (T -test, $p > .06$) in the dominant phyla (Actinobacteria, Proteobacteria, Firmicutes, Acidobacteria) between low-pH and high-pH soils (Figure S1). The diversity of GyrB at the phylum and class levels was also comparable (Figure S4) between low- and high-pH soils. Furthermore, there were no major differences in the broad functional capabilities (based on GO terms) between the high-pH and low-pH sites (Figure S2).

However, there was a significant reduction (T -test, unpaired, $p < .05$) in the various APases (PhoX, PhoD, PhoA) detected in low-pH soils compared to high-pH soils. In high-pH soils, 47%, 56%, and 20% of bacteria possessed genes encoding PhoX, PhoD, and PhoA, respectively, whereas in low-pH soils, only 3%, 7% and 1% of bacteria contained these genes, respectively (Figure 1A). Unexpectedly, the number of bacteria containing class A and C ACPases was similar (T -test, unpaired, $p > .05$) between all sites while the number of bacteria containing class B ACPases actually decreased (T -test, unpaired, $p = .046$) in low-pH soils (Figure 1A). Besides the nucleotidase, UshA (Rittmann et al., 2005), which also showed a significant increase in high-pH soils, the abundance of several other enzymes involved in organic P scavenging (phytases, phosphodiesterases) did not change (T -test, unpaired, $p > .05$) between low-pH and high-pH soils. The diversity of the three APases was comparable to that of GyrB

FIGURE 1 The abundance of various P-scavenging enzymes detected in the eight metagenomes retrieved from soils with contrasting pH values (red, low pH; blue, high pH). The percentage of bacteria containing phosphatases was calculated assuming a copy number of one per cell. A reduction in the three alkaline phosphatases was observed in low-pH soils while there was no concurrent increase in acid phosphatases. Abbreviations: PhoX, PhoD, PhoA, alkaline phosphatases; classA/B/C, nonspecific acid phosphatases; UshA, 5' nucleotidase, GlpQ, glycerolphosphodiester phosphodiesterase; PhnI, carbon-phosphorus lyase complex subunit I



and the 16S rRNA gene (Figure 2 and S3). For example, the majority of APase sequences (63%–73%) were related to the Proteobacteria, Actinobacteria, and Firmicutes.

Based on our curated databases for APases, the majority of sequences found in the soils analyzed here had a mean amino acid identity of 69% to those found in the Integrated Microbial Genomes database

at the Joint Genome Institute (IMG/JGI). However, due to a lack of biochemical and biophysical data on the majority of APases found in less-studied bacteria that make up our curated database, for example, members of the *Betaproteobacteria* or *Bacteroidetes*, it remains to be determined whether all of the sequences related to the various APases represent *bona fide* APases. Future work should ascertain whether or

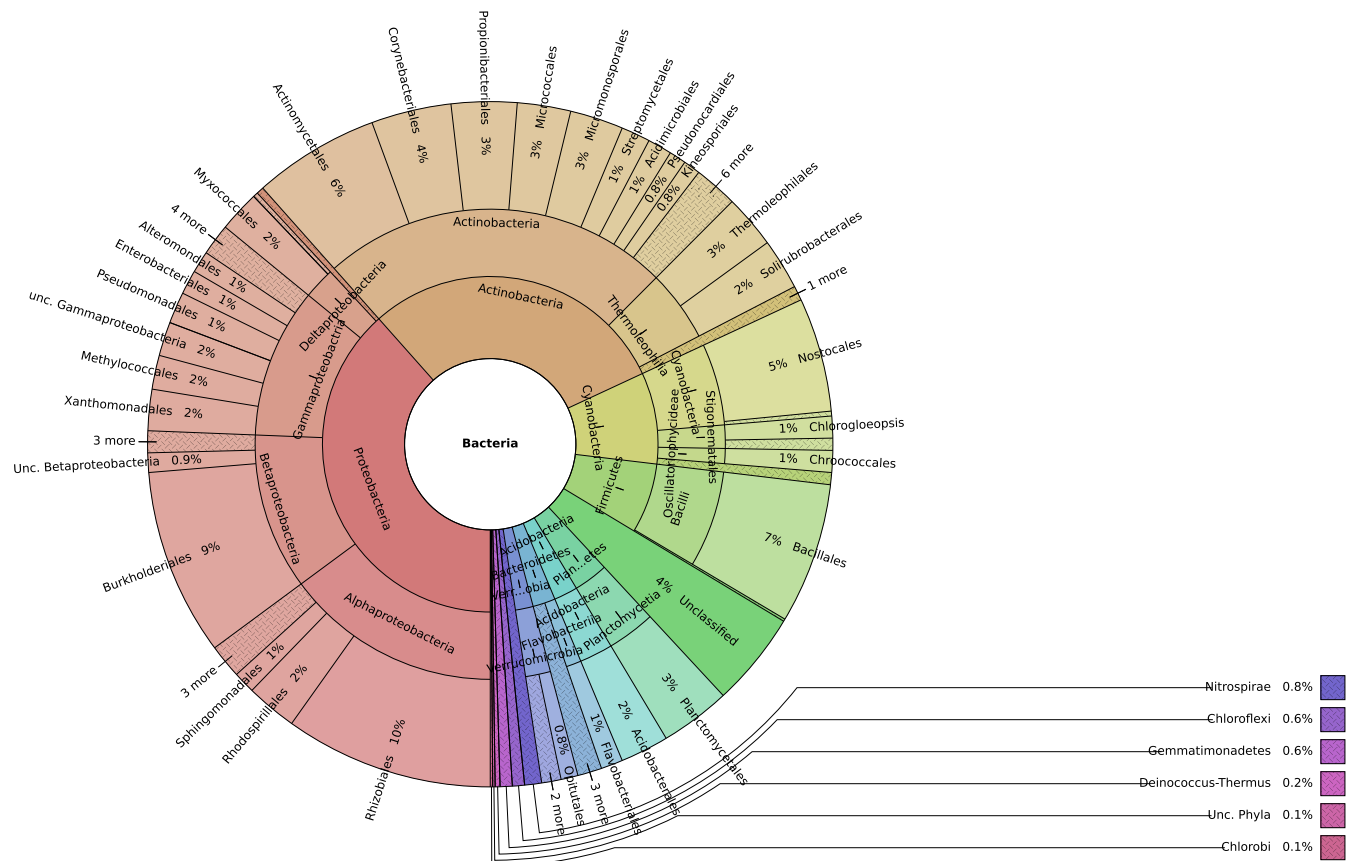


FIGURE 2 Combined taxonomic diversity of the three alkaline phosphatases (PhoX, PhoD, PhoA) in the metagenomes retrieved from all four high-pH sites (CS922, CS78, CS251, CS511). The chart was generated using the KronaTools software package (Ondov et al., 2011)

not predicted APases found in genomic databases, having significant sequence divergence from the relatively few characterized APases, function in a similar manner to those studied to date.

To determine the number of sequenced bacterial genomes possessing APases, we scrutinized all the genomes (status, 'finished') deposited in the IMG/JGI for the presence of each APase. It was discovered that approximately half of the strains related to the Proteobacteria (1510/2244), Actinobacteria (257/494), Firmicutes (471/1052), and Cyanobacteria (66/99) possess at least one of the three APases (PhoX, PhoA, PhoD). Furthermore, only 165/258 genomes (status, finished) tagged with 'soil' and 37/45 genomes (status, finished) tagged with 'rhizome/rhizoplane' under the filter 'ecosystem type' harbor one of the three APases, revealing that a number of bacteria exist that do not possess the typical phosphatases associated with overcoming Pi limitation (Lidbury et al., 2016; Santos-Beneit, 2015).

Together, this analysis revealed that low-pH soils have a marked reduction in their 'known genetic potential' to remineralize organic P. In a previous study, using a targeted amplicon approach, soil pH was also shown to be a driver of *phoD* diversity (Ragot, Kertesz, & Bünemann, 2015). Whether this reduction in genetic potential found in low-pH soils equates to a reduction in P remineralization capabilities warrants further investigation. Although APases have a pH optimum between 9 and 11, PhoX showed activity against 79 phosphomonoesters at pH 7.5 (Zaheer et al., 2009) and PhoD functions well at pH 8 (Rodríguez et al., 2014). However, it is likely that these promiscuous enzymes do not function in soils at such low-pH values. Considering that the pH optimum of ACPases is much lower (pH 4.8–7) (Rossolini et al., 1998), it was surprising that there was no observed increase in the genes encoding these enzymes in low-pH soils.

Low-pH soils may select for microbial communities containing bacteria lacking 'known' APases (PhoX, PhoD, PhoA), but which possess as-yet-unidentified phosphatases, rather than a total loss of non-specific phosphatases from bacterial genomes. This hypothesis is indirectly supported by evidence that numerous bacterial strains deposited in genome banks lack characterized APases. Given the fact that soils harbor a tremendous amount of genetic diversity (Torsvik & Øvreås, 2002) and that multiple Phyla inhabit the rhizosphere (Bulgarelli, Schlaeppi, Spaepen, van Themaat, & Schulze-Lefert, 2013; Bulgarelli et al., 2012), our knowledge of the P-scavenging abilities of soil bacteria remains relatively poor. Clearly our understanding of the microbial response to Pi limitation is limited to studies focusing on easily culturable strains. Future studies should focus on APase-lacking isolates to determine their P mineralization capabilities and thus their capacity to overcome P scarcity in soils. This should include the isolation and cultivation of various bacterial strains from low-pH soils to determine if they still elicit phosphatase activity or conversely, are reliant solely on various forms of inorganic P.

ACKNOWLEDGEMENTS

This research was funded by the Biotechnology and Biological Sciences Research Council (BBSRC) under grant BB/L026074/1.

CONFLICT OF INTEREST

The authors declare no conflict of interest.

REFERENCES

- Angelini, S., Moreno, R., Gouffi, K., Santini, C., Yamagishi, A., Berenguer, J., & Wu, L. (2001). Export of *Thermus thermophilus* alkaline phosphatase via the twin-arginine translocation pathway in *Escherichia coli*. *FEBS Letters*, 506, 103–107.
- Antelmann, H., Scharf, C., & Hecker, M. (2000). Phosphate starvation-inducible proteins of *Bacillus subtilis*: proteomics and transcriptional analysis. *Journal of Bacteriology*, 182, 4478–4490.
- Bulgarelli, D., Rott, M., Schlaeppi, K., Loren, V., van Themaat, E., Ahmadinejad, N., ... Schulze-Lefert, P. (2012). Revealing structure and assembly cues for *Arabidopsis* root-inhabiting bacterial microbiota. *Nature*, 488, 91–95.
- Bulgarelli, D., Schlaeppi, K., Spaepen, S., van Themaat, E. V. L., & Schulze-Lefert, P. (2013). Structure and functions of the bacterial microbiota of plants. *Annual Review of Plant Biology*, 64, 807–838.
- Condrón, L. M., Turner, B. L., & Cade-Menun, B. J. (2005). Chemistry and dynamics of soil organic phosphorus. In J. T. Sims, & A. N. Sharpley (Eds.), *Phosphorus: agriculture and the environment* (pp. 87–121). Soil Science Society of America Inc, Madison, WI: American Society of Agronomy.
- Cordell, D., Drangert, J.-O., & White, S. (2009). The story of phosphorus: Global food security and food for thought. *Global Environmental Change*, 19, 292–305.
- Eddy, S. R. (1998). Profile hidden Markov models. *Bioinformatics*, 14, 755–763.
- Eder, S., Shi, L., Jensen, K., Yamane, K., & Hulett, F. M. (1996). A *Bacillus subtilis* secreted phosphodiesterase/alkaline phosphatase is the product of a *Pho* regulon gene, *phoD*. *Microbiology*, 142, 2041–2047.
- Gandhi, N. U., & Chandra, S. B. (2012). A comparative analysis of three classes of bacterial non-specific acid phosphatases and archaeal phosphoesterases: Evolutionary perspective. *Acta Informatica Medica*, 20, 167–173.
- Griffiths, R. I., Thomson, B. C., James, P., Bell, T., Bailey, M., & Whiteley, A. S. (2011). The bacterial biogeography of British soils. *Environmental Microbiology*, 13, 1642–1654.
- Hirani, T. A., Suzuki, I., Murata, N., Hayashi, H., & Eaton-Rye, J. J. (2001). Characterization of a two-component signal transduction system involved in the induction of alkaline phosphatase under phosphate-limiting conditions in *Synechocystis* sp. PCC 6803. *Plant Molecular Biology*, 45, 133–144.
- Hedley, M. J., Mortvedt, J. J., Bolan, N. S., & Syers, J. K. (1995). Phosphorus Fertility Management In Agroecosystems. In H. Tiessen (Ed.), *Phosphorus in the Global Environment: Transfers, Cycles and Management* (pp. 59–92). USA: Wiley & Sons Ltd.
- Howard, E. C., Sun, S., Biers, E. J., & Moran, M. A. (2008). Abundant and diverse bacteria involved in DMSP degradation in marine surface waters. *Environmental Microbiology*, 10, 2397–2410.
- Lidbury, I. D. E. A., Murphy, A. R. J., Scanlan, D. J., Bending, G. D., Jones, A. M. E., Moore, J. D., ... Wellington, E. M. H. (2016). Comparative genomic, proteomic and exoproteomic analyses of three *Pseudomonas* strains reveals novel insights into the phosphorus scavenging capabilities of soil bacteria. *Environmental Microbiology*, 18, 3535–3549. doi: 10.1111/1462-2920.13390
- Mengel, K. (1997). Agronomic measures for better utilization of soil and fertilizer phosphates. *European Journal of Agronomy*, 7, 221–233.
- Monds, R. D., Newell, P. D., Schwartzman, J. A., & O'Toole, G. A. (2006). Conservation of the *Pho* regulon in *Pseudomonas fluorescens* Pf0-1. *Applied and Environment Microbiology*, 72, 1910–1924.
- Nannipieri, P., Newell, P. D., Giagnoni, L., & Landi, L. Renella, G. (2011). Role of phosphatase enzymes in soil. In E. K. Bünemann (Eds.), *et al.*,

- Phosphorus in action, soil biology* (pp. 215–243) Berlin Heidelberg, Springer-Verlag.
- Ondov, B. D., Bergman, N. H., & Phillippy, A. M. (2011). Interactive metagenomic visualization in a web browser. *BMC Informatics*, 12, 385. doi: 10.1186/1471-2105-12-385
- Putker, F., Tommassen-van Boxtel, R., Stork, M., Rodríguez-Herva, J. J., Koster, M., & Tommassen, J. (2013). The type II secretion system (Xcp) of *Pseudomonas putida* is active and involved in the secretion of phosphatases. *Environmental Microbiology*, 15, 2658–2671.
- Ragot, S. A., Kertesz, M. A., & Bünemann, E. K. (2015). *phoD* alkaline phosphatase gene diversity in soil. *Applied and Environment Microbiology*, 81, 7281–7289.
- Rittmann, D., Sorger-Herrmann, U., & Wendisch, V. F. (2005). Phosphate starvation-inducible gene *ushA* encodes a 5' nucleotidase required for growth of *Corynebacterium glutamicum* on media with nucleotides as the phosphorus source. *Applied and Environment Microbiology*, 71, 4339–4344.
- Rodríguez, H., Fraga, R. (1999). Phosphate solubilizing bacteria and their role in plant growth promotion. *Advanced Biotech*, 17, 319–339.
- Rodriguez, F., Lillington, J., Johnson, S., Timmel, C. R., Lea, S. M., & Berks, B. C. (2014). Crystal structure of the *Bacillus subtilis* phosphodiesterase PhoD reveals an iron and calcium-containing active site. *Journal of Biological Chemistry*, 289, 30889–30899.
- Rossolini, M. G., Schippa, S., Riccio, L. M., Berlutti, F., Macaskie, E. L., & Thaller, C. M. (1998). Bacterial nonspecific acid phosphohydrolases: Physiology, evolution and use as tools in microbial biotechnology. *Cellular and Molecular Life Sciences*, 54, 833–850.
- Roy, N. K., Ghosh, R. K., & Das, J. (1982). Monomeric alkaline phosphatase of *Vibrio cholerae*. *Journal of Bacteriology*, 150, 1033–1039.
- Santos-Beneit, F. (2015). The Pho regulon: A huge regulatory network in bacteria. *Frontiers in Microbiology*, 6, 402.
- Sebastian, M., & Ammerman, J. W. (2009). The alkaline phosphatase PhoX is more widely distributed in marine bacteria than the classical PhoA. *ISME Journal*, 3, 563–572.
- Thaller, M. C., Berlutti, F., Schippa, S., Lombardi, G., & Rossolini, G. M. (1994). Characterization and sequence of PhoC, the principal phosphate-irrepressible acid phosphatase of *Morganella morganii*. *Microbiology*, 140, 1341–1350.
- Torsvik, V., & Øvreås, L. (2002). Microbial diversity and function in soil: From genes to ecosystems. *Current Opinion in Microbiology*, 5, 240–245.
- Vance, C. P., Uhde-Stone, C., & Allan, D. L. (2003). Phosphorus acquisition and use: Critical adaptations by plants for securing a nonrenewable resource. *New Phytologist*, 157, 423–447.
- White, P. J., & Hammond, J. P. (2008). Phosphorus nutrition of terrestrial plants. In P. J. White, & J. P. Hammond (Eds.), *The ecophysiology of plant-phosphorus interactions* (pp. 51–81). Dordrecht: Springer, Netherlands.
- White, P. J., & Hammond, J. P. (2009). The sources of phosphorus in the waters of Great Britain. *Journal of Environmental Quality*, 38, 13–26.
- Yang, K., & Metcalf, W. W. (2004). A new activity for an old enzyme: *Escherichia coli* bacterial alkaline phosphatase is a phosphite-dependent hydrogenase. *Proceedings of the National Academy of Sciences of the United States of America*, 101, 7919–7924.
- Zaheer, R., Morton, R., Proudfoot, M., Yakunin, A., & Finan, T. M. (2009). Genetic and biochemical properties of an alkaline phosphatase PhoX family protein found in many bacteria. *Environmental Microbiology*, 11, 1572–1587.

SUPPORTING INFORMATION

Additional Supporting Information may be found online in the supporting information tab for this article.

How to cite this article: Lidbury IDEA, Fraser T, Murphy ARJ, et al. The 'known' genetic potential for microbial communities to degrade organic phosphorus is reduced in low-pH soils. *MicrobiologyOpen*. 2017;00:e474. <https://doi.org/10.1002/mbo3.474>