

Original citation:

Tatton, Andrew S., Blade, Helen, Brown, Steven P., Hodgkinson, Paul, Hughes, Leslie P., Nilsson Lill, Sten and Yates, Jonathan R. (2018) Improving confidence in crystal structure solutions using NMR crystallography : the case of β -piroxicam. *Crystal Growth & Design* . doi:10.1021/acs.cgd.8b00022 (In Press)

Permanent WRAP URL:

<http://wrap.warwick.ac.uk/101322>

Copyright and reuse:

The Warwick Research Archive Portal (WRAP) makes this work by researchers of the University of Warwick available open access under the following conditions. Copyright © and all moral rights to the version of the paper presented here belong to the individual author(s) and/or other copyright owners. To the extent reasonable and practicable the material made available in WRAP has been checked for eligibility before being made available.

Copies of full items can be used for personal research or study, educational, or not-for profit purposes without prior permission or charge. Provided that the authors, title and full bibliographic details are credited, a hyperlink and/or URL is given for the original metadata page and the content is not changed in any way.

Publisher's statement:

This document is the Accepted Manuscript version of a Published Work that appeared in final form in *Crystal Growth & Design*, copyright © American Chemical Society after peer review and technical editing by the publisher.

To access the final edited and published work see

<http://dx.doi.org/10.1021/acs.cgd.8b00022>

A note on versions:

The version presented here may differ from the published version or, version of record, if you wish to cite this item you are advised to consult the publisher's version. Please see the 'permanent WRAP url' above for details on accessing the published version and note that access may require a subscription.

For more information, please contact the WRAP Team at: wrap@warwick.ac.uk

1 **Improving confidence in crystal structure solutions using NMR crystallography: the case of β -**
2 **piroxicam**

3
4 **Andrew S. Tatton^a, Helen Blade^b, Steven P. Brown^c, Paul Hodgkinson^d, Leslie P. Hughes^b, Sten Nilsson Lill^e,**
5 **and Jonathan R. Yates^a**

6 ^a Department of Materials, University of Oxford, Oxford, OX1 3PH, UK

7 E-mail: Andrew.Tatton@materials.ox.ac.uk, Jonathan.Yates@materials.ox.ac.uk

8 ^b Pharmaceutical Development, AstraZeneca, Macclesfield, SK10 2NA, UK

9 E-mail: Les.Hughes2@astrazeneca.com

10 ^c Department of Physics, University of Warwick, Coventry, CV4 7AL, UK

11 E-mail: S.P.Brown@warwick.ac.uk

12 ^d Department of Chemistry, Durham University, Durham, DH1 3LE, UK

13 E-mail: Paul.Hodgkinson@durham.ac.uk

14 ^e Early Product Development, Pharmaceutical Sciences, IMED Biotech Unit, AstraZeneca Gothenburg, Mölndal, SE-431 83, Sweden

15
16 **Abstract**

17 NMR crystallographic techniques are used to validate a structure of β -piroxicam determined from powder
18 X-ray diffraction (PXRD) with a relatively poor R-factor. Geometry optimisation of PXRD- and single-
19 crystal XRD- derived structures results in convergence to the same energy of the structures, with minimal
20 atomic displacements, and good agreement of gauge-included projector augmented wave (GIPAW)
21 calculated and experimentally determined NMR ¹H, ¹³C and ¹⁵N chemical shifts and ¹⁴N quadrupolar
22 parameters. Calculations on isolated molecules combined with 2D magic-angle spinning (MAS) ¹H double-
23 quantum (DQ) and ¹⁴N-¹H NMR experiments confirm the 3D packing arrangement of β -piroxicam. NMR
24 crystallography is shown to be an effective means of validating crystal structures that might otherwise be
25 considered sceptically on the basis of diffraction data alone.

26 **Introduction**

27 A comprehensive understanding of the different crystal structures adopted by an active pharmaceutical
28 ingredient (API) is critical throughout the development process of a new medicine for both characterisation

1 and regulatory purposes. A detailed knowledge of the specific crystal structures facilitates an improved
2 understanding of the polymorph landscape, reliable property predictions, both physical and chemical, and
3 the ability to perform better solid-state risk assessments.¹ In a pharmaceutical context, a solid-state risk
4 assessment is performed to aid in selection of the final commercial solid form and manufacturing control
5 required.

6 Structural elucidation of a material using single crystal X-ray diffraction (SCXRD) is typically the preferred
7 structure solution technique, but growth of a single crystal of sufficient size and quality for SCXRD is often
8 not possible, and alternative methods must be considered. PXRD remains a more challenging, although
9 increasingly feasible,² pathway for obtaining a structure solution relative to SCXRD. For example, light
10 elements, such as hydrogen, have a low electron density and therefore interact only weakly with X-ray
11 radiation. This is a limitation where accurate determination of a hydrogen position is crucial, for example
12 when distinguishing between a pharmaceutical salt or co-crystal.

13 A structural model from a diffraction study is accompanied by a discrepancy value, which quantifies the fit
14 of the proposed structural model to experimental data. Although a number of discrepancy values have been
15 presented and discussed,³⁻⁵ the mostly widely used is the conventional R-factor,⁶ which describes agreement
16 between the amplitude of structure factors determined from the proposed structural model and
17 corresponding experimental X-ray diffraction data. The R-factor is an important indicator of the confidence
18 of a crystal structure model, but it is important to note that a low R-factor does not always correspond to a
19 good crystal structure description.⁷ There is no single source of a high R-factor, rather a multitude of
20 potential uncertainties exist. These can include high-temperature acquisition, a small number of sampling
21 angles, low quality data (weak intensity, poor resolution) and somewhat paradoxically, higher quality data
22 (increased intensity and/or resolution) that may amplify the influence of any systematic errors.⁵ In the case
23 where a structural model is solved with a high R-factor, or a degree of uncertainty in the structural model
24 exists, then a methodology that complements the diffraction data to increase confidence in the structural
25 model is highly desirable.

1 In recent years, NMR crystallography,⁸⁻¹⁴ which utilises a combination of solid-state NMR experimental
2 data and calculated NMR parameters, has emerged as a powerful tool for structural refinement and
3 elucidation of various materials, such as pharmaceutical salts,¹⁵⁻¹⁶ co-crystals,¹⁷⁻²⁰ and hydrates.^{16, 21-22}
4 Furthermore, the wide range of NMR-active nuclei potentially included within a NMR crystallography
5 study increases the range of systems that can be studied. The accuracy of a proposed model can be gauged
6 from a comparison between chemical shift values,²³⁻²⁴ and for spin $>1/2$ nuclei, the quadrupolar interaction
7 can also be determined, using experiment and calculation.²⁵⁻²⁷ Insight into intermolecular packing
8 configurations can be provided by 2D correlation experiments, where an observed correlation between
9 atoms is due to through-space or through-bond couplings. Experimentally observed interactions between
10 atoms in neighbouring molecules can then be compared to the expected intermolecular configuration from
11 a geometry-optimised structural model.²⁸ NMR crystallography has also been used extensively to probe
12 intermolecular interactions that determine the 3D packing arrangement, such as hydrogen bonding and
13 aromatic dispersive forces.²⁹⁻³¹ Quantitative analysis of intermolecular interactions has been demonstrated
14 from comparison between calculated chemical shift values obtained from a full crystal calculation relative
15 to chemical shifts of an isolated molecule.³²

16 To demonstrate that NMR crystallography can act as a complementary methodology for diffraction studies
17 to increase the confidence in a structural model, we present an NMR crystallography study for different
18 structural models of piroxicam, with emphasis upon a structural model solved from PXRD with a high R-
19 factor. Piroxicam (IUPAC name: 4-hydroxy-2-methyl-*N*-(2-pyridinyl)-2*H*-1,2-benzothiazine-3-
20 carboxamide-1,1-dioxide) is a non-steroidal, analgesic, anti-inflammatory drug.³³ Piroxicam does exhibit
21 polymorphism, although only β -piroxicam, which is also referred to as form I by Sheth et al.,³⁴ is discussed
22 within this article. Recently, Shi et al.³⁵ deposited a β -piroxicam structure in the CSD solved from SCXRD
23 (CSD reference code: BIYSEH13), with an associated R-factor of 2.4%. Previously β -piroxicam has been
24 solved from PXRD (CSD reference code: BIYSEH03) with a reported R-factor of 19%.³⁶ These particular
25 values may imply that the former model is a significantly more accurate structure solution than the latter,

1 and is explored in this work. Note that the cause of the high R-factor for BIYSEH03 is not discussed by
2 Sheth et al.³⁶

3 These two structures provide an ideal case study owing to their different R-factors and respective
4 methodologies used to solve the structure. Initially, the structural models (BIYSEH03 and BIYSEH13) are
5 geometry optimised by applying Density Functional Theory (DFT) using the CASTEP code.³⁷ Using the
6 geometry-optimised structures, NMR parameters are calculated using the GIPAW (gauge-including
7 projector augmented wave) method.³⁸⁻³⁹ 1D and 2D magic-angle spinning (MAS) solid-state NMR
8 experiments are performed to determine ¹H, ¹³C and ¹⁵N chemical shifts for comparison to calculated NMR
9 chemical parameters derived from the geometry optimised structural model. Additionally interatomic
10 distances from the geometry-optimised structural model are linked to experimental data from 2D MAS
11 NMR correlation experiments.

12 **Experimental and calculation details**

13 β -piroxicam was purchased from Sigma Aldrich and was used as supplied. PXRD confirmed that correct
14 form I was supplied (see supporting information, Fig. S1). ¹³C ramped cross-polarisation magic-angle
15 spinning (CPMAS)⁴⁰ and 2D ¹³C-¹H CP Heteronuclear Correlation (HETCOR)⁴¹ solid-state NMR
16 experiments were performed using a Bruker Avance III HD spectrometer operating at a ¹H Larmor
17 frequency of 399.9 MHz using a Bruker 4 mm double resonance probe at spinning frequencies of 12.5 kHz
18 and 10 kHz, respectively. For ¹H-¹³C cross-polarisation, SPINAL-64⁴²⁻⁴³ ¹H decoupling was applied during
19 acquisition at a nutation frequency of 100 kHz for 35 ms. The ¹H 90° pulse duration was 2.5 μ s, and ¹³C
20 magnetisation was created using a CP ramp of magnitude 90% to 100% for a duration of 2 ms. A recycle
21 delay of 10 s was used, and a total of 1024 transients were co-added. During ¹³C-¹H 2D HETCOR
22 experiments, SPINAL-64 ¹H decoupling was applied during t_2 at a nutation frequency of 100 kHz for a
23 duration of 20 ms, and ¹H homonuclear decoupling during t_1 was achieved using the Frequency Switched
24 Lee-Goldburg (FSLG)⁴⁴ method at a nutation frequency of 100 kHz. A total of 38 t_1 FIDs were recorded at
25 increments of 75.53 μ s using the States-TPPI method for sign discrimination in the indirect dimension. A

1 recycle delay of 5 s was used, and 320 transients were co-added for each slice. ^1H - ^{13}C magnetisation transfer
2 was achieved using a magnitude 50% to 100% ramp applied on the proton channel. ^{13}C chemical shifts are
3 referenced to TMS at 0 ppm, using the CH_2 peak in adamantane at 38.5 ppm as a secondary reference.⁴⁵

4 ^1H experiments were performed using a Bruker Avance III HD spectrometer operating at a ^1H Larmor
5 frequency of 399.9 MHz using a Bruker 1.3 mm double resonance probe. Single pulse experiments were
6 performed using a spinning frequency of 65 kHz, 8 co-added transients and a recycle delay of 30 s. ^1H DQ-
7 SQ Back-to-Back (BABA)⁴⁶⁻⁴⁷ rotor synchronised (in F_1) spectra were obtained using a spinning frequency
8 of 60 kHz, 16 co-added transients per each t_1 slice and a recycle delay of 30 s. ^1H chemical shifts are
9 referenced to TMS at 0 ppm, using the CH_2 peak in adamantane at 1.8 ppm as a secondary reference.⁴⁸

10 ^{15}N cross-polarisation experiments were performed using a Bruker Avance II spectrometer operating at a
11 ^1H Larmor frequency of 500.5 MHz and a Bruker 4 mm double resonance probe at a spinning frequency of
12 10 kHz. SPINAL-64 ^1H decoupling was applied during acquisition at a nutation frequency of 100 kHz for
13 40 ms. The ^1H 90° pulse duration was 2.5 μs , and ^{15}N magnetisation was created using a CP ramp of
14 magnitude 50% to 100% for a duration of 5 ms. A total of 7168 co-added transients were acquired using a
15 recycle delay of 10 s. ^{15}N chemical shifts are referenced to liquid nitromethane at 0 ppm,⁴⁹ using the NH_3^+
16 peak in glycine at -346.9 ppm as a secondary reference. To convert to the chemical shift scale frequently
17 used in protein NMR, where the alternative IUPAC reference (see Appendix 1 of Ref. 50) is liquid ammonia
18 at 50°C , it is necessary to add 379.5 ppm to the given values.⁵¹

19

20 ^{14}N - ^1H Heteronuclear Multiple Quantum (HMQC) experiments⁵² were performed using a Bruker Avance
21 III spectrometer operating at a ^1H Larmor frequency of 700.1 MHz using a Bruker 1.3 mm triple resonance
22 probe, operating in double resonance mode, at a spinning frequency of 60 kHz. Rotary resonance recoupling
23 (R^3) with a $n=2$ resonance condition⁵³ with an x $-x$ phase inversion⁵⁴⁻⁵⁵ of individual blocks lengths of
24 duration 16.7 μs was implemented. A total of 16 and 32 co-added transients were acquired for shorter and
25 longer recoupling time experiments, respectively, and a recycle delay of 32 s was used. States-TPPI was

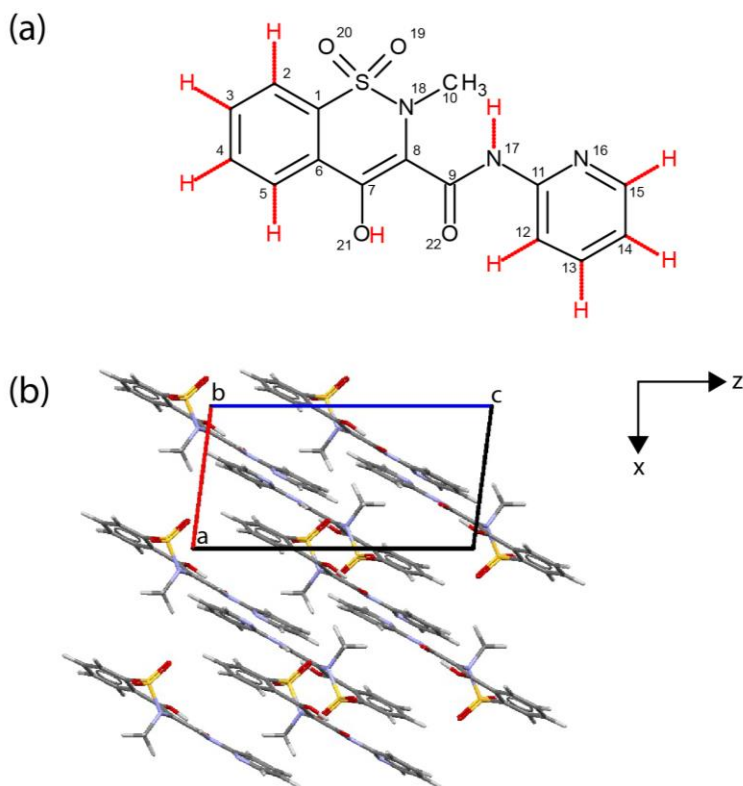
1 used to provide sign discrimination, and a total of 52 t_1 FIDs were recorded at increments of 16.7 μs . ^{14}N
2 shifts are referenced to liquid nitromethane at 0 ppm, using the NH_4 peak in a saturated aqueous solution
3 of NH_4Cl at -352.9 ppm as a secondary reference (see Table 2 of Ref. 49).

4 First-principle calculations were performed using CASTEP³⁷ version 17.0.8 in the Materials Studio
5 platform. All calculations employed the Perdew-Burke-Ernzerhof (PBE)⁵⁶ functional with the Tkatchenko
6 and Scheffler (TS) dispersion scheme,⁵⁷ and on-the-fly generated ultrasoft pseudopotentials (OTFG). For
7 all geometry optimisation calculations, a plane-wave cut-off energy of 600 eV was used with a Monkhorst-
8 Pack grid with a minimum k-point sampling space of 0.05 \AA^{-1} , which corresponds to a k-point grid with
9 dimensions (3, 1, 1). Crystal structures of BIYSEH03 and BIYSEH13 were downloaded from the
10 Cambridge Structural Database. Note that the lattice parameters stated for BIYSEH03 in Table 1 of Ref.
11 36 differ slightly from those of the deposited structure. All 144 atoms in the unit cell ($Z = 4$, $Z' = 1$) were
12 allowed to relax, unless otherwise stated, and calculations were performed with unit cell parameters fixed,
13 unless otherwise stated. Distances and angles presented are for geometry-optimised structures. The space
14 group $P2_1/c$ was preserved during all relaxations. NMR shielding values were calculated using the
15 GIPAW³⁸⁻³⁹ method for the geometry optimised structures, with a plane-wave cut-off energy of 700 eV,
16 and the same k-point grid and pseudopotentials as for the relaxations. Visualisation of GIPAW NMR results
17 was performed using MagresView.⁵⁸

18 **Results and Discussion**

19 *Effect of geometry optimisation on different β -piroxicam structure solutions*

20 The chemical structure of β -piroxicam is shown with numbering in Fig. 1a. Different numbering schemes
21 for piroxicam are found in the literature.^{35, 59-61} The carbon atom labels in Fig. 1a agree with those used in
22 Sheth et al.⁵⁹ The hydrogen atoms take the number of their respective through-bond heteroatom (methyl
23 protons are labelled H10a, H10b, and H10c).



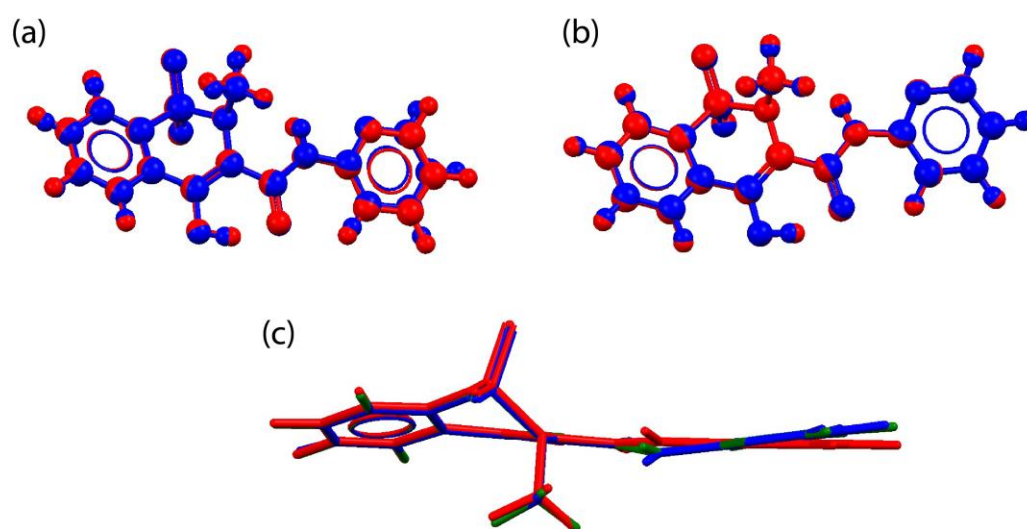
1
 2 **Figure 1.** (a) Chemical structure of β -piroxicam. (b) View along crystallographic axis b showing the stacking
 3 arrangement in the BIYSEH03 structure.

4 Both the β -piroxicam structures studied are modelled in the monoclinic space group $P2_1/c$, one molecule
 5 per asymmetric unit cell ($Z' = 1$) and four molecules in the conventional unit cell ($Z = 4$). Inspection of both
 6 β -piroxicam structural models indicate a short distance between N17 and O19 that suggests a putative
 7 intermolecular N17-H17...O19 hydrogen bond, which may direct the formation of dimers along the
 8 crystallographic axis b, as suggested in previous studies.^{34, 61-62} Intramolecular hydrogen bonding has also
 9 been indicated between N17-H17...N18³⁴ and O21-H21...O22.⁶⁰⁻⁶¹

10 Visualisations of the structural models, before and after geometry optimisation (unit cell parameters fixed,
 11 all atoms optimised), are presented in Fig. 2a and Fig. 2b, respectively. After geometry optimisation, there
 12 is a lengthening of heteronuclear X-H bond lengths for both structural models, and, for BIYSEH03 only, a
 13 slight rotation of the pyridine ring and a change in direction of the NH bond. The displacement in atomic

1 position of each proton and carbon due to geometry optimisation is presented in the supporting information
 2 (Tables S1-S3).

3 In Fig. 2c, an additional BIYSEH03 structural overlay is included, shown in green, obtained from a
 4 geometry optimisation where only hydrogen positions were allowed to relax. It is evident that allowing
 5 only hydrogen positions to relax prohibits the rotation of the pyridine ring, since the carbon atoms are fixed.
 6 Furthermore, the direction of the N17-H17 bond angle is now located at an intermediate point between the
 7 original, non-geometry-optimised structure, and the all-atom geometry-optimised structure. This indicates
 8 that the structure solution from PXRD did not locate N17 at its lowest energy position, and as can be seen
 9 in Table S2, the largest distance between hydrogen positions between fully and partially (hydrogen only)
 10 geometry optimised structures is for H17.



11
 12 **Figure 2.** Ball and stick representations before (blue) and after (red) geometry optimisation, of (a) BIYSEH03 and (b)
 13 BIYSEH13. Unit cell parameters were fixed during geometry optimisation. (c) Overlay of BIYSEH03 before
 14 geometry optimisation (blue), geometry optimisation of proton positions only (green) and full-atom geometry-
 15 optimisation (red).

16 **Table 1.** Selected output parameters from all atom geometry optimisations for BIYSEH03 and BIYSEH13, with and
 17 without relaxation of unit cell parameters.

Structure code	Unit cell parameters	RMSD displacement ^a / Å		Lattice parameters				Volume
		H	C	<i>a</i> / Å	<i>b</i> / Å	<i>c</i> / Å	β / °	<i>V</i> / Å ³

BIYSEH03	Fixed	0.036	0.015	7.16	15.18	13.99	97.4	1504.5
BIYSEH03	Relaxed ^b	0.033	0.016	7.07	15.41	13.75	95.3	1492.4
BIYSEH13	Fixed ^b	0.010	0.005	7.03	14.98	13.89	96.4	1453.5
BIYSEH13	Relaxed	0.009	0.006	7.05	15.31	13.84	95.7	1486.7

1 ^a Displacement relative to non-geometry optimised structural model, determined as an average of the displacement in atomic co-
2 ordinates of the respective nuclei within 1 molecule. ^b Starting unit cell parameters were the same as those for the respective
3 geometry optimisation with fixed unit cell parameters. ^c Symmetry group $P2_1/c$ was preserved, therefore $\alpha = \gamma = 90^\circ$ was maintained.

4 As shown in Table 1, it is evident that there is a significantly larger shift in atomic positions, reflected in
5 the RMSD values, after a geometry optimisation for the crystal structure derived from PXRD. This structure
6 was solved using DASH,⁶³ which by default does not include the hydrogen atoms in the structure factor
7 calculation, and hydrogen atoms are simply added at standard distances.³⁶ For the structure solution of
8 BIYSEH13 determined from SCXRD, the electron density difference map has been used to assign the
9 hydrogen positions.³⁵

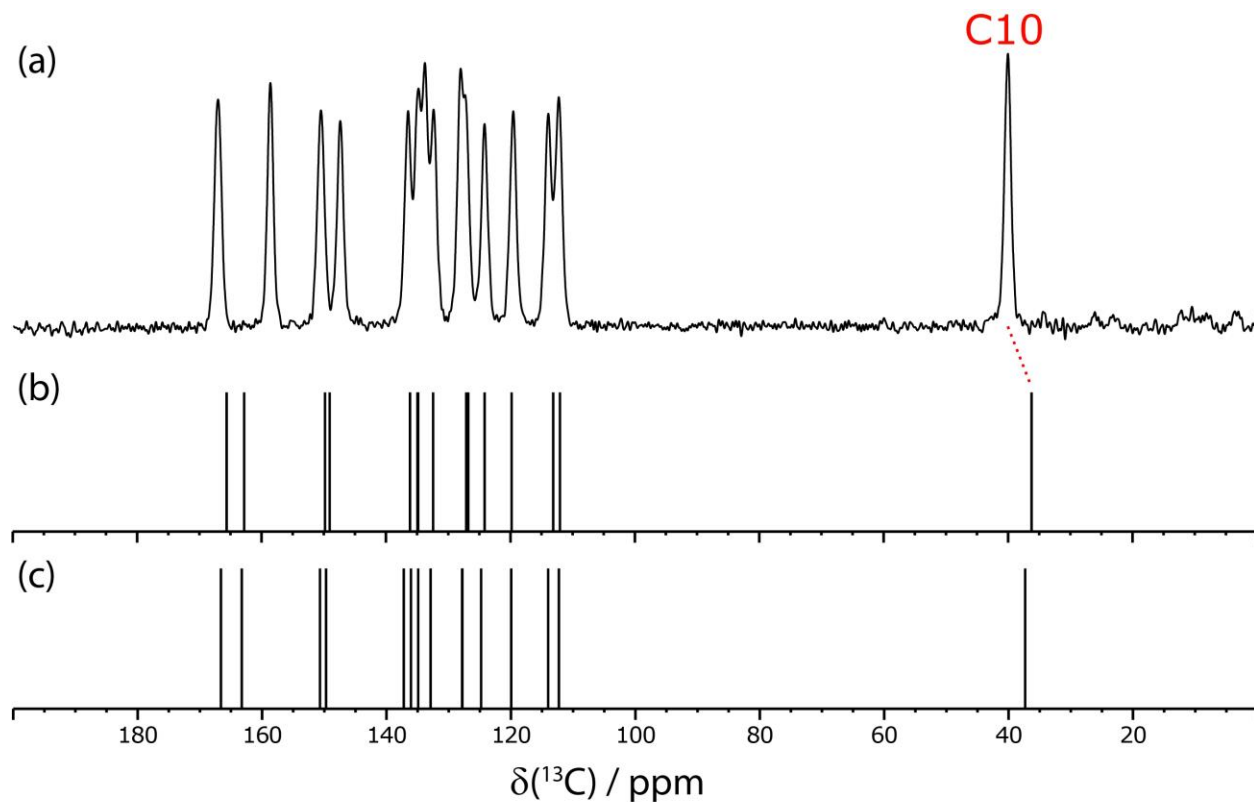
10 Allowing the unit cell parameters to relax during geometry optimisation does not have a significant effect
11 upon the atomic positions relative to the geometry-optimised structure with unit cell parameters fixed,
12 although unit cell parameters and cell volume differ when relaxation of the unit cell parameters is permitted.
13 After unit cell varying optimisation, the energies are the same (less than 0.1 kJ/ mol difference), whereas,
14 with unit cells fixed, one energy is 4.8 kJ / mol lower in energy. Predicted PXRD patterns for the original
15 structural models and subsequent geometry-optimised structures included in Table 1 can be found in the
16 supporting information (Fig. S2). Unsurprisingly, there is a larger difference between predicted powder
17 patterns of BIYSEH03, before and after geometry optimisation, relative to BIYSEH13. After a full atom
18 geometry optimisation, the RMSD value between atomic positions relative to the SCXRD structure solution
19 is 0.19 Å when considered over 15 molecules, which is the default molecular cluster size when comparing
20 crystal packing when using Mercury.

21
22 Conversion to an alternate polymorphic form is unlikely as geometry optimisation is always a refinement
23 to a local energy minimum, and therefore will only occur if the starting structure is closer to another form.

1 In the case of β -piroxicam, no significant change is noted between final structures determined using a
2 geometry optimisation with unit cell parameters allowed to relax, and full atom relaxation, relative to fixed
3 unit cell parameters and protons only allowed to relax during geometry optimisation.

4
5 The convergence of the two structures indicates that BIYSEH03, which owing to a high R-factor would
6 normally be treated with a degree of caution in a solid-state risk assessment, is a reasonably sound structural
7 model. Unit cell parameters are slightly different between the two structures, which will account for some
8 of this difference. An overlay of the respective unit cells is included in the Supporting Information (Fig.
9 S3). Note, that these are after a full atom geometry optimisation, with fixed unit cell parameters. Without
10 access to the original diffraction datasets any possible reduction in R-factor from the optimised structure of
11 BIYSEH03 is not determinable.

12 ***Comparison between experimental and GIPAW calculated NMR chemical shift values***



13

1 **Figure 3.** (a) A ^{13}C CPMAS spectrum of β -piroxicam obtained at 9.4 T and a MAS frequency of 12.5 kHz. Stick
2 spectra representing ^{13}C chemical shift GIPAW NMR values calculated for the (b) BIYSEH03 and (c) BIYSEH13
3 geometry-optimised structures. To convert to a scale comparable to TMS = 0 ppm, $\delta_{\text{iso}} = -(\sigma_{\text{iso}} - \sigma_{\text{ref}})$, where $\sigma_{\text{ref}} =$
4 170.4 ppm (b) and 170.2 ppm (c). σ_{ref} was calculated from averaging the $\delta(^{13}\text{C})_{\text{expt}}$ and σ_{iso} values.

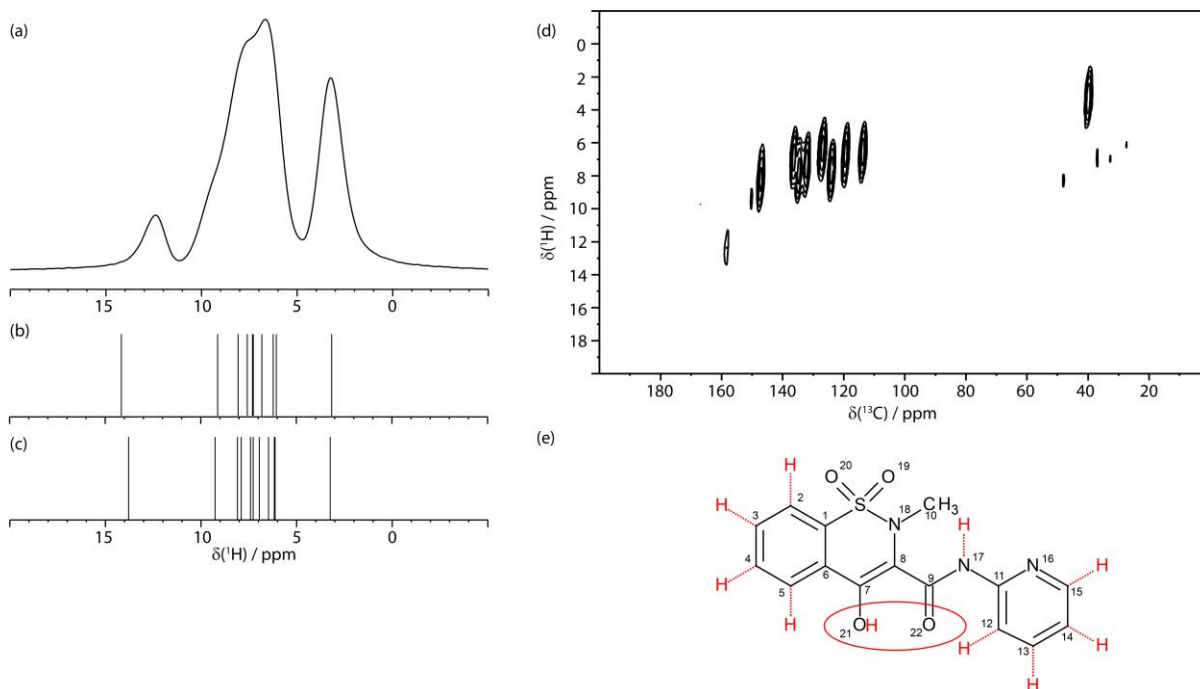
5 Particularly if the only structure solution available for a solid-state risk assessment is from PXRD with a
6 high R-factor, then it remains desirable to verify a structure solution with a methodology that is not
7 diffraction based, ideally with a direct link to experiment, NMR crystallography can provide this additional
8 confidence. Furthermore, the RMSD displacement range of the carbon atoms falls below the 0.25 Å limit
9 suggested by van de Streek et al. as an incorrect structure or revealing of interesting structural features,⁶⁴
10 and therefore there is an incentive to investigate if NMR crystallography is sensitive to any finer structural
11 differences.

12 Figure 3 compares (a) ^{13}C experimental isotropic chemical shift values, determined from a ^{13}C CPMAS
13 experiment, and (b, c) chemical shift values determined from a GIPAW NMR calculation for BIYSEH03
14 and BIYSEH13, respectively, after geometry optimisation of all atomic positions and fixed unit cell
15 parameters. Experimental assignment was assisted by performing a ^{13}C non-quaternary suppression
16 experiment (see Supporting information, Fig. S4), and is in agreement with previous ^{13}C solid-state NMR
17 studies of piroxicam.⁶⁵⁻⁶⁷ Previous studies indicate that the associated error in the ^{13}C chemical shift for a
18 GIPAW NMR calculation using the PBE functional is approximately 2 ppm, i.e., approximately 1% of the
19 chemical shift range.¹² Broad agreement is noted between all sites (complete chemical shift tables can be
20 found in the Supporting information, Tables S5-S6) except for C10, as indicated by the red dashed line
21 included in Fig. 3. The RMSD values between experiment and calculated ^{13}C chemical shifts are 1.7 ppm
22 (BIYSEH03) and 1.6 ppm (BIYSEH13), however if C10 is discounted the ^{13}C RMSD value is 1.3 ppm for
23 both models.

24 The discrepancy in C10 chemical shifts is most likely a consequence of the referencing system used for
25 GIPAW NMR calculation. The challenges in overcoming such a referencing discrepancy are discussed in
26 the supporting information. Additionally, the influence of nuclear delocalisation in methyl proton positions,
27 and their subsequent effect upon the ^{13}C chemical shift,⁶⁸ may account for this discrepancy between

1 experiment and calculation. An alternative approach for the referencing system is to calculate an averaged
2 value of σ_{ref} for a series of well assigned materials,⁶⁹⁻⁷⁰ or for a single well defined reference compound,⁷¹⁻
3 ⁷² and apply this value to all further calculations. This approach was not chosen herein as β -piroxicam is a
4 highly aromatic compound, and therefore as discussed by Webber et al.,⁷³ ^{13}C σ_{ref} values are likely to be
5 skewed by the absence of peaks in the lower ppm ^{13}C region.

6 As shown in Fig. 2c, a geometry optimisation of BIYSEH03 with only hydrogen positions allowed to relax
7 prevents rotation of the pyridine ring as compared to the case of a full atom geometry optimisation. However,
8 the effect upon ^{13}C GIPAW calculated chemical shieldings is less clear. The ^{13}C magnetic shieldings (σ_{iso})
9 for BIYSEH03 structures (proton only relaxation versus all atom relaxation) differ by an average of 2.7
10 ppm. This also influences, however, the determined value of σ_{ref} , which is used to convert magnetic
11 shieldings to a scale comparable to experimental chemical shift values. Consequently, the calculated δ_{iso}
12 values are converted to values that closely match geometry-optimisation conditions. By comparison, the
13 average difference between ^{13}C σ_{iso} values for the BIYSEH13 geometry-optimised structures is 0.7 ppm.
14 This observation is unsurprising, given that the atomic coordinates of BIYSEH13 move less after geometry
15 optimisation.



1
 2 **Figure 4.** (a) A ^1H one pulse spectrum obtained for β -piroxicam at 9.4 T and a MAS frequency of 65 kHz. (b, c) Stick
 3 spectra representing ^1H chemical shift GIPAW NMR values calculated for the (b) BIYSEH03 and (c) BIYSEH13
 4 geometry-optimised crystal structures. To convert to a scale comparable to TMS = 0 ppm, $\delta_{\text{iso}} = -(\sigma_{\text{iso}} - \sigma_{\text{ref}})$, where
 5 $\sigma_{\text{ref}} = 30.7$ ppm for both structures. σ_{ref} was calculated from averaging the $\delta(^1\text{H})_{\text{expt}}$ and σ_{iso} values. (d) A ^{13}C - ^1H
 6 HETCOR ($\nu_r = 10$ kHz, $B_0 = 9.4$ T) spectrum obtained for β -piroxicam using a cross-polarization contact time of 200
 7 μs and FSLG homonuclear decoupling in F_1 . (e) Chemical structure of β -piroxicam with intramolecular O21-
 8 H21...O22 hydrogen bonding motif highlighted.

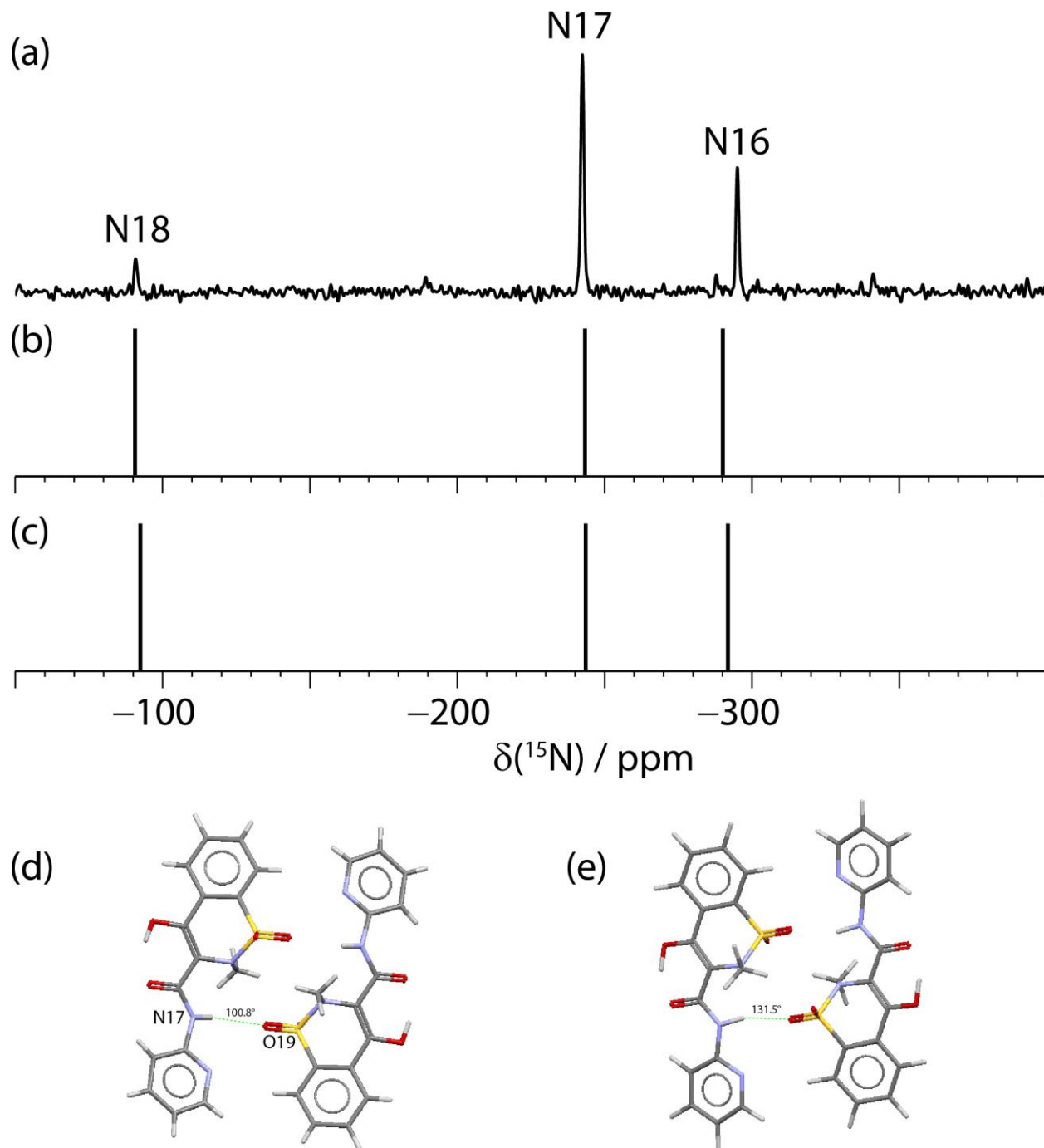
9 As protons often directly participate in the intermolecular packing arrangement, for example via hydrogen
 10 bonding or aromatic dispersive forces, any inaccuracies in the structural model will be clearly seen as
 11 discrepancies between experimental and calculated GIPAW NMR ^1H chemical shift values. Figure 4, parts
 12 (a-c), show a comparison between a single-pulse ^1H spectrum obtained using fast MAS and stick spectra
 13 corresponding to calculated GIPAW NMR chemical shifts of BIYSEH03 and BIYSEH13, respectively.
 14 Owing to the presence of strong homonuclear dipolar couplings, ^1H solid-state NMR spectra usually yield
 15 broad peaks that prevent individual identification of each individual ^1H chemical shift. Modest
 16 improvement in resolution can be achieved by applying fast MAS; here a MAS rotation frequency of 65
 17 kHz was applied and consequently spectral regions (methyl, aromatic, carboxylic acid), although not
 18 individual peaks, are distinguishable. The peaks at approximately 12 and 3 ppm correspond to the
 19 carboxylic acid proton and the methyl protons, respectively. However, owing to the large number of

1 aromatic protons present in piroxicam, it is not possible to individually assign aromatic protons in the region
2 between 5 and 10 ppm using the 1D MAS NMR spectrum.

3 Individual proton chemical shifts can be identified indirectly using 2D NMR experiments, where the
4 introduction of an indirect dimension allows for improved spectral separation of protons via the
5 heteronucleus. In this case the higher resolution in the ^{13}C dimension is used to indirectly probe proton
6 environments using a ^{13}C - ^1H HETCOR experiment,⁴¹ which is typically performed at more modest spinning
7 rates so as not to inhibit magnetisation transfer between ^{13}C and ^1H nuclei. Improvement of the resolution
8 in the proton dimension can be achieved by application of homonuclear ^1H dipolar decoupling (FSLG)
9 during t_1 . Fig. 4d presents a ^{13}C - ^1H HETCOR spectrum obtained using a short contact time of 200 μs . The
10 short contact time means that predominantly only C-H pairs in close-proximity (i.e., through-bond CH
11 pairs) are observed, and thus the ^{13}C and ^1H resonances can be directly assigned. The tilt observed in the
12 cross-peaks is attributed to Anisotropic Bulk Magnetic Susceptibility (ABMS) broadening.⁷⁴⁻⁷⁵ Table S7
13 and Table S8 shows a full comparison between experimental ^1H isotropic chemical shifts, derived using the
14 ^{13}C - ^1H HETCOR spectrum shown in Fig. 4d, and calculated GIPAW NMR chemical shift values. The
15 RMSD difference between experiment and calculation for ^1H chemical shift values are 0.6 ppm and 0.5
16 ppm for BIYSEH03 and BIYSEH13 when all hydrogen atoms are considered. These values indicate
17 structural agreement for both models,^{11, 76-78} providing further evidence that BIYSEH03, despite its high R-
18 factor, is an accurate structural model.

19
20 When comparing different geometry optimisation criteria (full atom versus hydrogens only), the same
21 general trend is noted for ^1H σ_{ref} values as was noted for ^{13}C σ_{ref} values: BIYSEH03 magnetic shieldings
22 differ on average by a larger value relative to BIYSEH13 σ_{ref} values, although the effect is reduced relative
23 to ^{13}C . The value of the pyridine ^1H GIPAW calculated NMR chemical shifts determined for BIYSEH03
24 do not significantly vary between the two geometry optimisation cases described. Relative to experimental
25 data, the ^1H GIPAW NMR shift of H17 calculated from the hydrogen-only geometry-optimised BIYSEH03

1 structural model differs by 0.9 ppm compared to experiment, as opposed to 0.5 ppm for a full atom geometry
2 optimisation (see H17 values in Table S7). This appears to agree with the observation that the largest proton
3 displacement extracted from the structural models was H17 (see Table S2).



4
5 **Figure 5.** (a) A ^{15}N CPMAS spectrum obtained for β -piroxicam at 11.7 T and a MAS frequency of 10 kHz. (b, c)
6 Stick spectra representing calculated ^{15}N chemical shift GIPAW NMR values calculated for the (b) BIYSEH03 and
7 (c) BIYSEH13 geometry optimised crystal structures. To convert to a scale comparable to liquid nitromethane at 0

1 ppm, $\delta_{\text{iso}} = -(\sigma_{\text{iso}} - \sigma_{\text{ref}})$, where $\sigma_{\text{ref}} = -160.1$ ppm (BIYSEH03) and -161.1 ppm (BIYSEH13). σ_{ref} was calculated
2 from averaging the $\delta(^{15}\text{N})_{\text{expt}}$ and σ_{iso} values. (d, e) Extracts of the BIYSEH03 structural model showing putative N17-
3 H17...O19 interactions (d) before and (e) after geometry optimisation.

4 Fig. 5 compares ^{15}N experimental chemical shifts and calculated GIPAW NMR chemical shifts. As for ^{13}C
5 and ^1H , there is good agreement between chemical shift values for both structural models. It is noted that
6 for the case of the putative intermolecular hydrogen bond,⁷⁹ N17-H17...O19, bond distances and angles
7 changed significantly in BIYSEH03 after geometry optimisation, as is presented in Fig 5, parts (d, e). As
8 N17 directly participates in this interaction, the absence of a significant difference between the ^{15}N
9 experimental chemical shifts relative to the calculated GIPAW NMR chemical shift values indicates that
10 both structures are correct after geometry optimisation.

11 Further insight into intermolecular packing in the structural model can be obtained from the comparison of
12 chemical shifts calculated for the full crystal relative to those in an isolated molecule. To simulate an
13 isolated molecule in a GIPAW calculation, a molecule is taken from the related crystal structure and placed
14 in a vacuum supercell, i.e. a unit cell with sufficiently large lattice parameters to effectively remove any
15 intermolecular interactions between neighbouring molecules.^{24, 32} Any protons participating in strong
16 hydrogen bonding are no longer deshielded for an isolated molecule calculation, and therefore have a lower
17 ppm value, whereas chemical shifts of protons subject to the effects of a ring current increase in the isolated
18 molecule calculation relative to a full crystal. From inspection of Table 2, H5, H12 and H21 produce
19 significant (magnitude greater than 1 ppm) chemical shift differences between the isolated molecule and
20 the full crystal. H5 and H12 are aromatic protons, and therefore the change in chemical shift for an isolated
21 molecule calculation is attributed to the absence of ring currents associated with aromatic moieties, whereby
22 aromatic dispersive forces are responsible for driving the intermolecular arrangement, that are otherwise
23 present in the full crystal. Interestingly, H17 does not display any significant change in proton chemical
24 shift, showing that this does not participate in an intermolecular hydrogen bond, which is unsurprising from
25 inspection of the non-planar extracted N17-H17...O19 bond angle. Table S4 in the supporting information
26 describes the N17-H17...O19 arrangement for both starting structural models and their respective

1 geometry-optimised models. Rather, NMR crystallography demonstrates that intermolecular packing
 2 arrangement for both structural models is predominately driven by aromatic dispersive forces, as opposed
 3 to a N17-H17...O19 hydrogen bonding motif.

4
 5

6 **Table 2.** GIPAW NMR calculated ^1H chemical shifts for a full crystal and for an isolated molecule in a vacuum for
 7 β -piroxicam (BIYSEH03 and 13), together with experimental ^1H chemical shifts.

Site	$\delta(^1\text{H})_{\text{expt}} / \text{ppm}$	BIYSEH03			BIYSEH13		
		$\delta(^1\text{H})_{\text{calc}} / \text{ppm}$ Full crystal	$\delta(^1\text{H})_{\text{calc}} / \text{ppm}$ Single molecule ^a	Difference / ppm	$\delta(^1\text{H})_{\text{calc}} / \text{ppm}$ Full crystal	$\delta(^1\text{H})_{\text{calc}} / \text{ppm}$ Single molecule ^a	Difference / ppm
H2	7.8	7.4	7.6	-0.2	7.4	7.6	-0.2
H3	7.4	7.4	7.4	0.0	7.4	7.4	0.0
H4	7.7	7.6	7.4	0.2	7.6	7.4	0.2
H5	6.4	6.1	8.0	-1.9	6.1	7.9	-1.8
H10a	3.2	3.5	3.2	0.3	3.4	3.3	0.1
H10b	3.2	2.6	2.2	0.4	2.4	2.2	0.3
H10c	3.2	3.6	2.8	0.8	3.8	2.8	1.0
H12	6.4	6.3	8.5	-2.2	6.2	8.4	-2.2
H13	6.6	6.8	7.6	-0.8	6.9	7.6	-0.7
H14	6.8	6.4	6.8	-0.4	6.4	6.8	-0.4
H15	8.3	8.2	8.2	0.0	8.1	8.2	-0.1
H17	9.6	9.2	9.0	0.2	9.3	9.0	0.3
H21	12.4	14.0	15.5	-1.5	13.6	15.3	-1.7

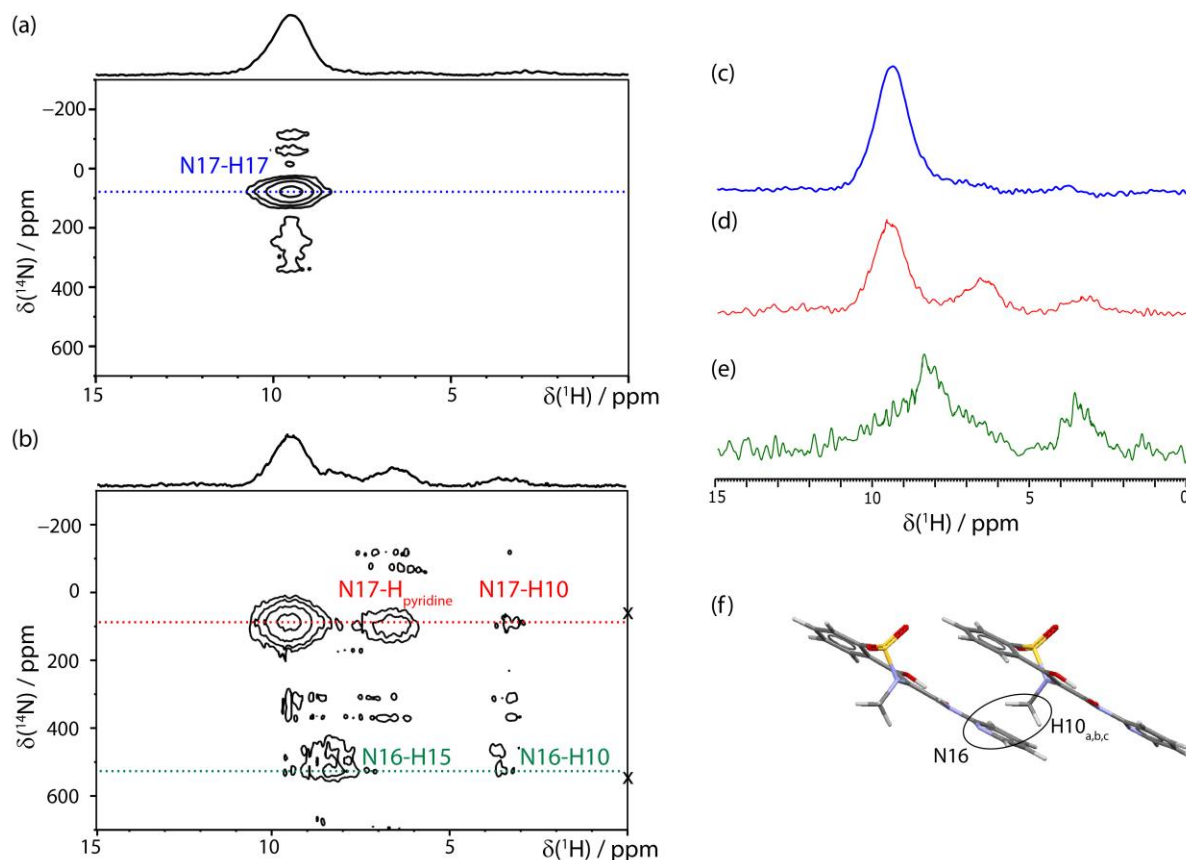
^a The same σ_{ref} values were used as specified in Fig. 4.

8

9 ***Using 2D MAS NMR to verify extracted intermolecular proximities from geometry-optimised structural***
 10 ***models***

11 An important confirmation of the intermolecular packing arrangement of the structural models is obtained
 12 from comparison between intermolecular correlations observed in 2D MAS NMR spectra relative to
 13 extracted intermolecular distances from the geometry-optimised structural models. The 2D experiments
 14 presented here provide a more qualitative form of structural confirmation. In the following experiments,
 15 magnetisation transfer is achieved when a sufficiently strong dipolar coupling between nuclei exists, which
 16 is only the case for closer proximity spin-pairs. Consequently, cross-peaks are only observed between nuclei
 17 within a proximity of up to approximately 4 Å, although no precise distance measurements are inferred.

1 Further information concerning the nitrogen environments can be inferred from ^{14}N experiments. ^{14}N is a
2 spin $I = 1$ nucleus, which is typically more experimentally challenging relative to nuclei with $I = 1/2$ owing
3 to the presence of the quadrupolar interaction (see SI for further information).⁸⁰⁻⁸¹ While experimental NMR
4 techniques for quadrupolar nuclei are more challenging compared to spin $I = 1/2$ nuclei, the quadrupolar
5 interaction does provide further information concerning the local electronic environment, and has been
6 demonstrated to be a powerful probe of intermolecular hydrogen bonding.^{52, 82-85} Additionally, whereas ^{15}N
7 experiments are severely limited by a low natural abundance (0.4%), ^{14}N has a high abundance (99.6%). In
8 recent years ^{14}N - ^1H Heteronuclear Multiple Quantum Coherence (HMQC) experiments performed at fast
9 MAS frequencies^{52, 83, 86-88} have been demonstrated as a powerful probe of pharmaceuticals.^{81, 89} These
10 experiments observe magnetisation upon the more sensitive proton spin, and polarisation is transferred from
11 protons to nitrogen via through-space couplings using a heteronuclear dipolar recoupling scheme. Fast
12 MAS is required to improve resolution in the proton dimension, and to increase the ^1H coherence lifetime
13 during the application of *rf* recoupling.^{86, 90}

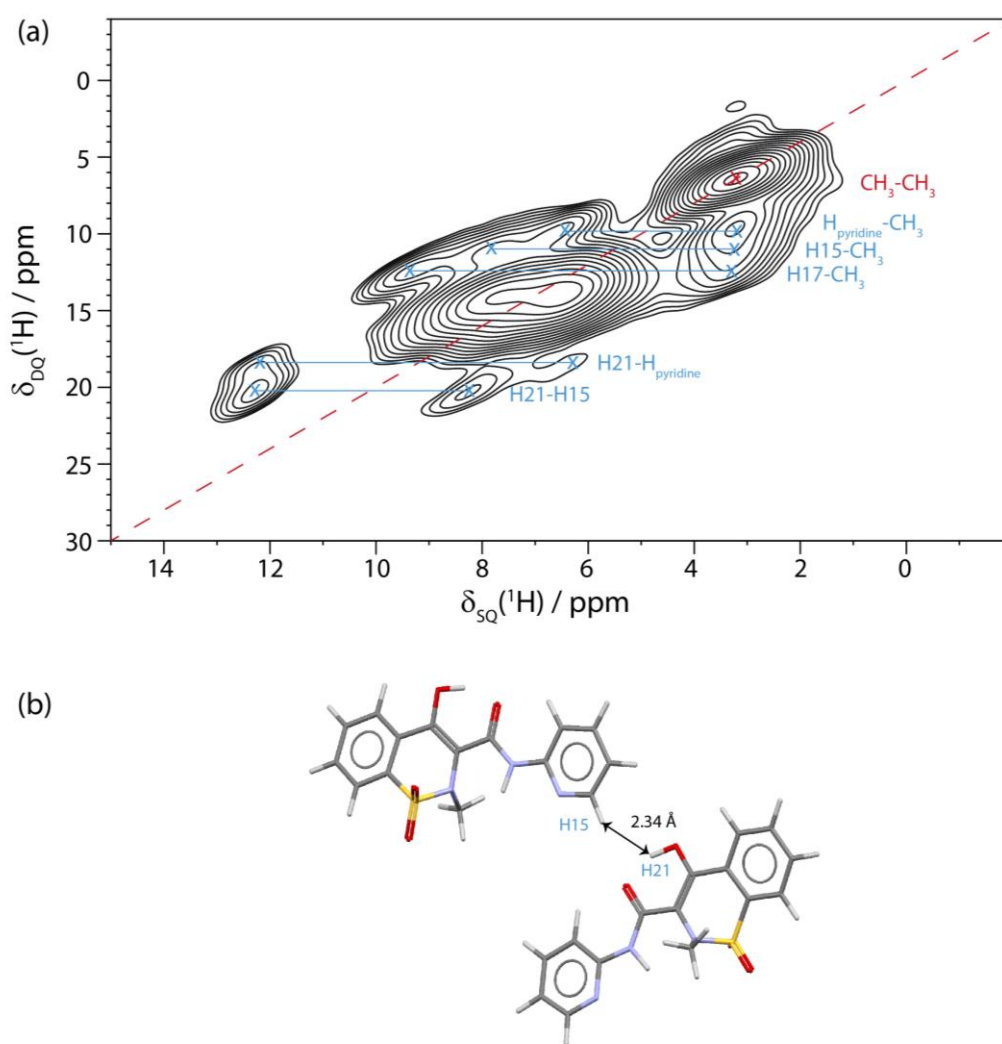


1
 2 **Figure 6.** (a, b) 2D ^{14}N - ^1H HMQC spectra with ^1H skyline projections of β -piroxicam ($\nu_r = 60$ kHz, $B_0 = 16.4$ T) using
 3 $n = 2$ rotary resonance recoupling of ^{14}N - ^1H heteronuclear dipolar couplings for (a) $\tau_{\text{RCPL}} = 266.7$ μs and (b) $\tau_{\text{RCPL}} =$
 4 533.3 μs . The crosses indicate the predicted centre-of-gravity of the N16 and N17 resonances determined from
 5 GIPAW NMR calculations. (c, d, e) ^1H individual sub-spectra taken through the ^{14}N resonance of corresponding colour
 6 to demonstrate which ^{14}N - ^1H heteronuclear dipolar couplings are recoupled owing to close through-space proximity.
 7 (f) Stacking arrangement of β -piroxicam from the geometry-optimised BIYSEH03 structural model.

8 In Fig. 6, ^{14}N - ^1H HMQC spectra recorded with different recoupling times are presented. These spectra were
 9 obtained at $B_0 = 16.4$ T. Spectra obtained at 9.4 T can be found in the supporting information (Fig. S6).
 10 Only the N17 site, which has the smallest C_Q , is visible at lower field, whereas N-H correlations between
 11 N16 and neighbouring protons are also visible at 16.4 T. Extracted distances referred to herein include
 12 hydrogen positions as hydrogen directly participates in the experimental observations shown.

13 At short recoupling times, only the directly-bonded NH correlation is observed, see Fig. 6a. A slice of the
 14 proton dimension, taken through the isotropic shift of N17 shows a single peak corresponding to the NH
 15 proton (H17), Fig. 6c. At longer recoupling times, see Fig. 6b, longer-range N-H proximities are visible.⁸⁶

1 A slice taken through N17 for a longer recoupling time is presented in Fig. 6d, and the intramolecular
2 correlations are labelled accordingly in Fig. 6b. Correlations from N16 to methyl protons and a further
3 correlation to a resonance at approximately 8.3 ppm are visible in Fig. 6e. The N16 correlation to the methyl
4 protons must be an intermolecular correlation; inspection of the geometry optimised structural model shows
5 the closest intramolecular N16-methyl distance is approximately 4.3 Å, whereas the shortest N16-H10
6 intermolecular distance is 3.1 Å. In Fig. 6f, an extract from the geometry optimised BIYSEH03 structural
7 model shows the stacking arrangement between molecules, with this methyl-N16 proximity circled.



8
9 **Figure 7.** (a) A ^1H DQ-SQ MAS spectrum of β -piroxicam recorded at $\nu_r = 60$ kHz and $\nu_0(^1\text{H}) = 400$ MHz using one
10 rotor period of Back-to-Back dipolar recoupling. The diagonal red line at $\delta_{\text{DQ}} = 2\delta_{\text{SQ}}$ is included as a visual aid, and

1 connected cross-peaks are indicated by blue horizontal dashed lines. (b) Structural extract from BIYSEH03
2 highlighting the close proximity (2.34 Å) of H15 and H21 as a consequence of the intermolecular packing arrangement.
3 Further details of through-space correlations that can be directly linked to the geometry optimised structural
4 model are determinable from a ^1H - ^1H DQ MAS experiment⁹¹⁻⁹² using BABA recoupling.^{46-47, 91} Performed
5 at fast MAS to improve spectral resolution, the homonuclear dipolar coupling is selectively recoupled
6 during the DQ excitation and reconversion periods. Consequently, correlations between close-proximity
7 through-space protons produce a pair of peaks (connected by blue dashed lines in Fig. 7) horizontally
8 equidistant from a diagonal line $\delta_{\text{DQ}} = 2\delta_{\text{SQ}}$ (included as a visual aid in red). Cross peaks on the diagonal
9 correspond to protons close enough in space to produce a through-space correlation to the same proton in
10 a neighbouring molecule, or a neighbouring proton with the same chemical shift value, e.g. the on-diagonal
11 peak for the methyl protons.⁹²

12 A clear cross peak between H21 (OH proton at 12.4 ppm) and H15 (at 8.3 ppm) is visible in Fig. 7a. As
13 H21 and H15 are the only proton resonances at these chemical shift values then this cross-peak can be
14 assigned with confidence. Inspection of the geometry optimised structural model (see Fig. 7b) indicates that
15 this corresponds to an intermolecular distance of approximately 2.4 Å (the intramolecular distance between
16 H21 and H15 is 7.7 Å). A cross-peak between H21 and a resonance at 6.4 ppm is also visible. This is
17 potentially an intermolecular correlation to a neighbouring pyridine proton, H14, although owing to
18 overlapping proton resonances in this spectral region this assignment is not unambiguous. A cross-peak
19 between H17 at 9.6 ppm and methyl protons at 3.2 ppm is observed, which is consistent with the correlation
20 between N17 and the methyl protons in the ^{14}N - ^1H HMQC experiment (see Fig. 6d).

21 **Conclusions**

22 We have demonstrated that NMR crystallography can act as a valuable complementary tool to diffraction
23 studies and associated metrics, such as the R-factor, to build confidence for two structural models solved
24 for β -piroxicam using PXRD and SCXRD, respectively. In particular the structure solution solved from
25 PXRD, BIYSEH03, has a high R-factor and therefore any associated solid-state risk assessment performed

1 for this model, or a model with an equivalently high R-factor, would benefit from any methodology that
2 increases confidence in the model, especially when the only structural model available has been solved
3 using PXRD.

4 We observe a convergence between the two structure solutions after geometry optimisation, which both
5 give good agreement between calculated and experimental NMR parameters, and are therefore
6 demonstrably more accurate structural models of β -piroxicam, relative to both starting structures. The
7 change in atomic positions after a geometry optimisation step clearly demonstrates the value in performing
8 a geometry optimisation as a routine step in any diffraction solved structural model,⁶⁴ particularly for a
9 powder diffraction. The results herein also indicate that in most cases all atoms in organic materials should
10 be relaxed during a geometry optimisation,⁹³ at least for a structural model solved using PXRD data. Herein,
11 this was demonstrated by the deviation in N17 and the pyridine ring between full-atom and hydrogen only
12 geometry optimisation. We demonstrate that NMR crystallography is a valuable tool for the verification of
13 different crystal structure solutions and as an additional indicator of the accuracy of a proposed structural
14 model to be used in conjunction with the existing R-factor.

15 Agreement between experimental chemical shifts and calculated GIPAW NMR chemical shifts determined
16 for ^{13}C , ^1H and ^{15}N verifies this convergence. The two structure solutions have the same GIPAW NMR
17 chemical shift values within reasonable error, and crucially both have a broad agreement with experiment.
18 Additional insight into the intermolecular arrangement of β -piroxicam is evident from comparison between
19 full crystal and isolated molecule calculations. These demonstrate that the intermolecular packing
20 arrangement of both models, which we consider accurate from chemical shift comparisons detailed, is
21 predominately via aromatic dispersive forces, as opposed to intermolecular hydrogen bonding. Furthermore,
22 we also note that experimentally observed intermolecular correlations in ^1H DQ and ^{14}N - ^1H MAS NMR
23 spectra between close-proximity spin-pairs agree with predicted short-range intermolecular proximities
24 extracted for both structural models, which provides further evidence of the accuracy of both structural
25 models.

1 **Associated Content**

2 The calculated and experimental data for this study are provided as a supporting dataset from WRAP, the
3 Warwick Research Archive Portal at http://wrap.warwick.ac.uk/**. The supporting information (PDF)
4 contains: (i) experimental verification of β -piroxicam form using PXRD, (ii) atomic displacement tables,
5 (iii) comparison of geometry optimised structures of BIYSEH03 and BIYSEH13, (iv) a ^{13}C CP non-
6 quaternary suppression spectrum, (v) a ^{13}C - ^1H HETCOR spectrum obtained with a longer contact time, (vi)
7 tables comparing experimental and calculated NMR parameters, and (vii) ^{14}N - ^1H spectra obtained at $B_0 =$
8 9.4 T. Geometry optimised structural models (cif format) of BIYSEH03 and BIYSEH13 calculated with
9 different relaxation conditions are also available to freely download.

10 **Acknowledgements**

11 We acknowledge support from the Collaborative Computing Project for NMR Crystallography (CCP-NC)
12 funded by EPSRC (EP/M022501/1). PH acknowledges support from EPSRC grant EP/L012243/1. The
13 European Research Council (ERC Starting Grant 639907 awarded to Jozef Lewandowski) and the
14 University of Warwick funded the 700 MHz Bruker Avance III spectrometer used for ^{14}N - ^1H experiments.
15 We acknowledge assistance from Anjali Menakath with the recording of the ^{14}N - ^1H spectra (700 MHz).

16

17 **References**

- 18 (1) Feeder, N.; Pidcock, E.; Reilly, A. M.; Sadiq, G.; Doherty, C. L.; Back, K. R.; Meenan, P.; Docherty,
19 R., The integration of solid-form informatics into solid-form selection. *J. Pharm. Pharmacol.* **2015**, *67*,
20 857-868.
- 21 (2) Hughes, C. E.; Reddy, G. N. M.; Masiero, S.; Brown, S. P.; Williams, P. A.; Harris, K. D. M.,
22 Determination of a complex crystal structure in the absence of single crystals: analysis of powder X-ray
23 diffraction data, guided by solid-state NMR and periodic DFT calculations, reveals a new 2'-
24 deoxyguanosine structural motif. *Chem. Sci.* **2017**, *8*, 3971-3979.
- 25 (3) Flack, H., On enantiomorph-polarity estimation. *Acta Crystallogr. Sect. A: Found. Crystallogr.* **1983**,
26 *39*, 876-881.
- 27 (4) Flack, H. D.; Bernardinelli, G., Reporting and evaluating absolute-structure and absolute-
28 configuration determinations. *J. Appl. Crystallogr.* **2000**, *33*, 1143-1148.
- 29 (5) Toby, B. H., R factors in Rietveld analysis: How good is good enough? *Powder Diffr.* **2012**, *21*, 67-
30 70.
- 31 (6) Groom, C. R.; Bruno, I. J.; Lightfoot, M. P.; Ward, S. C., The Cambridge Structural Database. *Acta*
32 *Crystallogr. Sect. B: Struct. Sci.* **2016**, *72*, 171-179.

- 1 (7) Buchsbaum, C.; Schmidt, M. U., Rietveld refinement of a wrong crystal structure. *Acta Crystallogr.*
2 *Sect. B: Struct. Sci.* **2007**, *63*, 926-932.
- 3 (8) Harris, R. K., NMR crystallography: the use of chemical shifts. *Solid State Sci.* **2004**, *6*, 1025-1037.
- 4 (9) Elena, B.; Pintacuda, G.; Mifsud, N.; Emsley, L., Molecular Structure Determination in Powders by
5 NMR Crystallography from Proton Spin Diffusion. *J. Am. Chem. Soc.* **2006**, *128*, 9555-9560.
- 6 (10) Taulelle, F., Fundamental Principles of NMR Crystallography. In *eMagRes*, John Wiley & Sons,
7 Ltd: 2007.
- 8 (11) Salager, E.; Stein, R. S.; Pickard, C. J.; Elena, B.; Emsley, L., Powder NMR crystallography of
9 thymol. *Phys. Chem. Chem. Phys.* **2009**, *11*, 2610-2621.
- 10 (12) Bonhomme, C.; Gervais, C.; Babonneau, F.; Coelho, C.; Pourpoint, F.; Azaïs, T.; Ashbrook, S. E.;
11 Griffin, J. M.; Yates, J. R.; Mauri, F.; Pickard, C. J., First-Principles Calculation of NMR Parameters
12 Using the Gauge Including Projector Augmented Wave Method: A Chemist's Point of View. *Chem. Rev.*
13 **2012**, *112*, 5733-5779.
- 14 (13) Ashbrook, S. E.; McKay, D., Combining solid-state NMR spectroscopy with first-principles
15 calculations - a guide to NMR crystallography. *Chem. Commun.* **2016**, *52*, 7186-7204.
- 16 (14) Bryce, D., NMR crystallography: structure and properties of materials from solid-state nuclear
17 magnetic resonance observables. *IUCrJ* **2017**, *4*, 350-359.
- 18 (15) Namespetra, A. M.; Hirsh, D. A.; Hildebrand, M. P.; Sandre, A. R.; Hamaed, H.; Rawson, J. M.;
19 Schurko, R. W., ³⁵Cl solid-state NMR spectroscopy of HCl pharmaceuticals and their polymorphs in bulk
20 and dosage forms. *CrystEngComm* **2016**, *18*, 6213-6232.
- 21 (16) Abraham, A.; Apperley, D. C.; Byard, S. J.; Ilott, A. J.; Robbins, A. J.; Zorin, V.; Harris, R. K.;
22 Hodgkinson, P., Characterising the role of water in sildenafil citrate by NMR crystallography.
23 *CrystEngComm* **2016**, *18*, 1054-1063.
- 24 (17) Chierotti, M. R.; Gobetto, R., NMR crystallography: the use of dipolar interactions in polymorph
25 and co-crystal investigation. *CrystEngComm* **2013**, *15*, 8599-8612.
- 26 (18) Fernandes, J. A.; Sardo, M.; Mafra, L.; Choquesillo-Lazarte, D.; Masciocchi, N., X-ray and NMR
27 Crystallography Studies of Novel Theophylline Cocrystals Prepared by Liquid Assisted Grinding. *Cryst.*
28 *Growth Des.* **2015**, *15*, 3674-3683.
- 29 (19) Luedeker, D.; Gossmann, R.; Langer, K.; Brunklaus, G., Crystal Engineering of Pharmaceutical Co-
30 crystals: "NMR Crystallography" of Niclosamide Co-crystals. *Cryst. Growth Des.* **2016**, *16*, 3087-3100.
- 31 (20) Kerr, H. E.; Softley, L. K.; Suresh, K.; Hodgkinson, P.; Evans, I. R., Structure and physicochemical
32 characterization of a naproxen-picolinamide cocrystal. *Acta Crystallogr. Sect. C: Cryst. Struct. Commun.*
33 **2017**, *73*, 168-175.
- 34 (21) Mafra, L.; Santos, S. M.; Siegel, R.; Alves, I.; Almeida Paz, F. A.; Dudenko, D.; Spiess, H. W.,
35 Packing Interactions in Hydrated and Anhydrous Forms of the Antibiotic Ciprofloxacin: a Solid-State
36 NMR, X-ray Diffraction, and Computer Simulation Study. *J. Am. Chem. Soc.* **2012**, *134*, 71-74.
- 37 (22) Reddy, G. N. M.; Cook, D. S.; Iuga, D.; Walton, R. I.; Marsh, A.; Brown, S. P., An NMR
38 crystallography study of the hemihydrate of 2', 3'-O-isopropylidineguanosine. *Solid State Nucl. Magn.*
39 *Reson.* **2015**, *65*, 41-48.
- 40 (23) Watts, A. E.; Maruyoshi, K.; Hughes, C. E.; Brown, S. P.; Harris, K. D. M., Combining the
41 Advantages of Powder X-ray Diffraction and NMR Crystallography in Structure Determination of the
42 Pharmaceutical Material Cimetidine Hydrochloride. *Cryst. Growth Des.* **2016**, *16*, 1798-1804.
- 43 (24) Zilka, M.; Sturniolo, S.; Brown, S. P.; Yates, J. R., Visualising crystal packing interactions in solid-
44 state NMR: Concepts and applications. *J. Chem. Phys.* **2017**, *147*, 144203.
- 45 (25) Perras, F. A.; Bryce, D. L., Multinuclear Magnetic Resonance Crystallographic Structure
46 Refinement and Cross-Validation Using Experimental and Computed Electric Field Gradients:
47 Application to Na₂Al₂B₂O₇. *J. Phys. Chem. C* **2012**, *116*, 19472-19482.
- 48 (26) Romao, C. P.; Perras, F. A.; Werner-Zwanziger, U.; Lussier, J. A.; Miller, K. J.; Calahoo, C. M.;
49 Zwanziger, J. W.; Bieringer, M.; Marinkovic, B. A.; Bryce, D. L.; White, M. A., Zero Thermal Expansion
50 in ZrMgMo₃O₁₂: NMR Crystallography Reveals Origins of Thermoelastic Properties. *Chem. Mater.* **2015**,
51 *27*, 2633-2646.

- 1 (27) Holmes, S. T.; Schurko, R. W., Refining Crystal Structures with Quadrupolar NMR and Dispersion-
2 Corrected Density Functional Theory. *J. Phys. Chem. C* **2018**, *122*, 1809-1820.
- 3 (28) Dudenko, D. V.; Yates, J. R.; Harris, K. D. M.; Brown, S. P., An NMR crystallography DFT-D
4 approach to analyse the role of intermolecular hydrogen bonding and π - π interactions in driving
5 cocrystallisation of indomethacin and nicotinamide. *CrystEngComm* **2013**, *15*, 8797-8807.
- 6 (29) Bradley, J. P.; Velaga, S. P.; Antzutkin, O. N.; Brown, S. P., Probing Intermolecular Crystal Packing
7 in γ -Indomethacin by High-Resolution ^1H Solid-State NMR Spectroscopy. *Cryst. Growth Des.* **2011**, *11*,
8 3463-3471.
- 9 (30) Filip, X.; Grosu, I.-G.; Miclaus, M.; Filip, C., NMR crystallography methods to probe complex
10 hydrogen bonding networks: application to structure elucidation of anhydrous quercetin. *CrystEngComm*
11 **2013**, *15*, 4131-4142.
- 12 (31) Carignani, E.; Borsacchi, S.; Bradley, J. P.; Brown, S. P.; Geppi, M., Strong Intermolecular Ring
13 Current Influence on ^1H Chemical Shifts in Two Crystalline Forms of Naproxen: a Combined Solid-State
14 NMR and DFT Study. *J. Phys. Chem. C* **2013**, *117*, 17731-17740.
- 15 (32) Yates, J. R.; Pham, T. N.; Pickard, C. J.; Mauri, F.; Amado, A. M.; Gil, A. M.; Brown, S. P., An
16 Investigation of Weak $\text{CH}\cdots\text{O}$ Hydrogen Bonds in Maltose Anomers by a Combination of Calculation
17 and Experimental Solid-State NMR Spectroscopy. *J. Am. Chem. Soc.* **2005**, *127*, 10216-10220.
- 18 (33) Brogden, R. N.; Heel, R. C.; Speight, T. M.; Avery, G. S., Piroxicam: A Review of its
19 Pharmacological Properties and Therapeutic Efficacy. *Drugs* **1981**, *22*, 165-187.
- 20 (34) Sheth, A. R.; Bates, S.; Muller, F. X.; Grant, D. J. W., Polymorphism in Piroxicam. *Cryst. Growth*
21 *Des.* **2004**, *4*, 1091-1098.
- 22 (35) Shi, X.; El Hassan, N.; Ikni, A.; Li, W.; Guiblin, N.; Spasojevic de-Bire, A.; Ghermani, N. E.,
23 Experimental electron densities of neutral and zwitterionic forms of the drug piroxicam. *CrystEngComm*
24 **2016**, *18*, 3289-3299.
- 25 (36) Sheth, A. R.; Bates, S.; Muller, F. X.; Grant, D. J. W., Local Structure in Amorphous Phases of
26 Piroxicam from Powder X-ray Diffractometry. *Cryst. Growth Des.* **2005**, *5*, 571-578.
- 27 (37) Clark, S. J.; Segall, M. D.; Pickard, C. J.; Hasnip, P. J.; Probert, M. I. J.; Refson, K.; Payne, M. C.,
28 First principles methods using CASTEP. *Z. Kristallogr.* **2005**, *220*, 567.
- 29 (38) Pickard, C. J.; Mauri, F., All-electron magnetic response with pseudopotentials: NMR chemical
30 shifts. *Phys. Rev. B* **2001**, *63*, 245101.
- 31 (39) Yates, J. R.; Pickard, C. J.; Mauri, F., Calculation of NMR chemical shifts for extended systems
32 using ultrasoft pseudopotentials. *Phys. Rev. B* **2007**, *76*, 024401.
- 33 (40) Metz, G.; Wu, X. L.; Smith, S. O., Ramped-Amplitude Cross Polarization in Magic-Angle-Spinning
34 NMR. *J. Magn. Reson., Ser A* **1994**, *110*, 219-227.
- 35 (41) van Rossum, B. J.; Förster, H.; de Groot, H. J. M., High-Field and High-Speed CP-MAS ^{13}C NMR
36 Heteronuclear Dipolar-Correlation Spectroscopy of Solids with Frequency-Switched Lee-Goldburg
37 Homonuclear Decoupling. *J. Magn. Reson.* **1997**, *124*, 516-519.
- 38 (42) Khitritin, A.; Fung, B. M., Design of heteronuclear decoupling sequences for solids. *J. Chem. Phys.*
39 **2000**, *112*, 2392-2398.
- 40 (43) Fung, B. M.; Khitritin, A. K.; Ermolaev, K., An Improved Broadband Decoupling Sequence for
41 Liquid Crystals and Solids. *J. Magn. Reson.* **2000**, *142*, 97-101.
- 42 (44) Bielecki, A.; Kolbert, A. C.; Levitt, M. H., Frequency-switched pulse sequences: Homonuclear
43 decoupling and dilute spin NMR in solids. *Chem. Phys. Lett.* **1989**, *155*, 341-346.
- 44 (45) Morcombe, C. R.; Zilm, K. W., Chemical shift referencing in MAS solid state NMR. *J. Magn.*
45 *Reson.* **2003**, *162*, 479-486.
- 46 (46) Sommer, W.; Gottwald, J.; Demco, D. E.; Spiess, H. W., Dipolar Heteronuclear Multiple-Quantum
47 NMR Spectroscopy in Rotating Solids. *J. Magn. Reson., Ser A* **1995**, *113*, 131-134.
- 48 (47) Schnell, I.; Lupulescu, A.; Hafner, S.; Demco, D. E.; Spiess, H. W., Resolution Enhancement in
49 Multiple-Quantum MAS NMR Spectroscopy. *J. Magn. Reson.* **1998**, *133*, 61-69.
- 50 (48) Hayashi, S.; Hayamizu, K., Chemical Shift Standards in High-Resolution Solid-State NMR (^1H , ^{13}C ,
51 ^{29}Si , and ^1H Nuclei. *Bull. Chem. Soc. Jpn.* **1991**, *64*, 685-687.

- 1 (49) Hayashi, S.; Hayamizu, K., Chemical Shift Standards in High-Resolution Solid-State NMR (2) ¹⁵N
2 Nuclei. *Bull. Chem. Soc. Jpn.* **1991**, *64*, 688-690.
- 3 (50) Harris, R. K.; Becker, E. D.; De Menezes, S. M. C.; Granger, P.; Hoffman, R. E.; Zilm, K. W.,
4 Further Conventions for NMR Shielding and Chemical Shifts (IUPAC Recommendations 2008). *Magn.*
5 *Reson. Chem.* **2008**, *46*, 582-598.
- 6 (51) Martin, G. E.; Hadden, C. E., Long-Range ¹H-¹⁵N Heteronuclear Shift Correlation at Natural
7 Abundance. *J. Nat. Prod.* **2000**, *63*, 543-585.
- 8 (52) Gan, Z.; Amoureux, J. P.; Trébosc, J., Proton-detected ¹⁴N MAS NMR using homonuclear
9 decoupled rotary resonance. *Chem. Phys. Lett.* **2007**, *435*, 163-169.
- 10 (53) Oas, T. G.; Griffin, R. G.; Levitt, M. H., Rotary resonance recoupling of dipolar interactions in
11 solid - state nuclear magnetic resonance spectroscopy. *J. Chem. Phys.* **1988**, *89*, 692-695.
- 12 (54) Costa, P. R.; Gross, J. D.; Hong, M.; Griffin, R. G., Solid-state NMR measurement of Ψ in peptides:
13 a NCCN 2Q-heteronuclear local field experiment. *Chem. Phys. Lett.* **1997**, *280*, 95-103.
- 14 (55) Webber, A. L.; Masiero, S.; Pieraccini, S.; Burley, J. C.; Tatton, A. S.; Iuga, D.; Pham, T. N.; Spada,
15 G. P.; Brown, S. P., Identifying Guanosine Self Assembly at Natural Isotopic Abundance by High-
16 Resolution ¹H and ¹³C Solid-State NMR Spectroscopy. *J. Am. Chem. Soc.* **2011**, *133*, 19777-19795.
- 17 (56) Perdew, J. P.; Burke, K.; Ernzerhof, M., Generalized Gradient Approximation Made Simple. *Phys.*
18 *Rev. Lett.* **1996**, *77*, 3865-3868.
- 19 (57) Tkatchenko, A.; Scheffler, M., Accurate Molecular Van Der Waals Interactions from Ground-State
20 Electron Density and Free-Atom Reference Data. *Phys. Rev. Lett.* **2009**, *102*, 073005.
- 21 (58) Sturniolo, S.; Green, T. F. G.; Hanson, R. M.; Zilka, M.; Refson, K.; Hodgkinson, P.; Brown, S. P.;
22 Yates, J. R., Visualization and processing of computed solid-state NMR parameters: MagresView and
23 MagresPython. *Solid State Nucl. Magn. Reson.* **2016**, *78*, 64-70.
- 24 (59) Sheth, A. R.; Lubach, J. W.; Munson, E. J.; Muller, F. X.; Grant, D. J. W., Mechanochromism of
25 Piroxicam Accompanied by Intermolecular Proton Transfer Probed by Spectroscopic Methods and Solid-
26 Phase Changes. *J. Am. Chem. Soc.* **2005**, *127*, 6641-6651.
- 27 (60) Kojic-Prodic, B.; Ruzic-Toros, Z., Structure of the anti-inflammatory drug 4-hydroxy-2-methyl-N-2-
28 pyridyl-2H-1λ6,2-benzothiazine-3-carboxamide 1,1-dioxide (piroxicam). *Acta Crystallogr. Sect. B:*
29 *Struct. Sci.* **1982**, *38*, 2948-2951.
- 30 (61) Taddei, P.; Torreggiani, A.; Simoni, R., Influence of environment on piroxicam polymorphism:
31 Vibrational spectroscopic study. *Biopolymers* **2001**, *62*, 68-78.
- 32 (62) Redenti, E.; Zanol, M.; Ventura, P.; Fronza, G.; Comotti, A.; Taddei, P.; Bertoluzza, A., Raman and
33 solid state ¹³C-NMR investigation of the structure of the 1 : 1 amorphous piroxicam : β-cyclodextrin
34 inclusion compound. *Biospectroscopy* **1999**, *5*, 243-251.
- 35 (63) David, W. I. F.; Shankland, K.; van de Streek, J.; Pidcock, E.; Motherwell, W. D. S.; Cole, J. C.,
36 DASH: a program for crystal structure determination from powder diffraction data. *J. Appl. Crystallogr.*
37 **2006**, *39*, 910-915.
- 38 (64) van de Streek, J.; Neumann, M. A., Validation of experimental molecular crystal structures with
39 dispersion-corrected density functional theory calculations. *Acta Crystallogr. Sect. B: Struct. Sci.* **2010**,
40 *66*, 544-558.
- 41 (65) Vrečer, F.; Vrbinc, M.; Meden, A., Characterization of piroxicam crystal modifications. *Int. J.*
42 *Pharm.* **2003**, *256*, 3-15.
- 43 (66) Vogt, F. G.; Clawson, J. S.; Strohmeier, M.; Edwards, A. J.; Pham, T. N.; Watson, S. A., Solid-State
44 NMR Analysis of Organic Cocrystals and Complexes. *Cryst. Growth Des.* **2009**, *9*, 921-937.
- 45 (67) Liu, W.; Wang, W. D.; Wang, W.; Bai, S.; Dybowski, C., Influence of Structure on the
46 Spectroscopic Properties of the Polymorphs of Piroxicam. *J. Phys. Chem. B* **2010**, *114*, 16641-16649.
- 47 (68) Dračinský, M.; Hodgkinson, P., Effects of Quantum Nuclear Delocalisation on NMR Parameters
48 from Path Integral Molecular Dynamics. *Chem. Eur. J.* **2014**, *20*, 2201-2207.
- 49 (69) Bryce, D. L.; Bultz, E. B.; Aebi, D., Calcium-43 Chemical Shift Tensors as Probes of Calcium
50 Binding Environments. Insight into the Structure of the Vaterite CaCO₃ Polymorph by ⁴³Ca Solid-State
51 NMR Spectroscopy. *J. Am. Chem. Soc.* **2008**, *130*, 9282-9292.

- 1 (70) Gervais, C.; Jones, C.; Bonhomme, C.; Laurencin, D., Insight into the local environment of
2 magnesium and calcium in low-coordination-number organo-complexes using ^{25}Mg and ^{43}Ca solid-state
3 NMR: a DFT study. *Acta Crystallogr. Sect. C: Cryst. Struct. Commun.* **2017**, *73*, 208-218.
- 4 (71) Burgess, K. M. N.; Korobkov, I.; Bryce, D. L., A Combined Solid-State NMR and X-ray
5 Crystallography Study of the Bromide Ion Environments in Triphenylphosphonium Bromides. *Chem.*
6 *Eur. J.* **2012**, *18*, 5748-5758.
- 7 (72) Li, X.; Tapmeyer, L.; Bolte, M.; van de Streek, J., Crystallographic and Dynamic Aspects of Solid-
8 State NMR Calibration Compounds: Towards ab Initio NMR Crystallography. *ChemPhysChem* **2016**, *17*,
9 2496-2502.
- 10 (73) Webber, A. L.; Emsley, L.; Claramunt, R. M.; Brown, S. P., NMR Crystallography of Campho[2,3-
11 c]pyrazole ($Z' = 6$): Combining High-Resolution ^1H - ^{13}C Solid-State MAS NMR Spectroscopy and
12 GIPAW Chemical-Shift Calculations. *J. Phys. Chem. A* **2010**, *114*, 10435-10442.
- 13 (74) Harris, R. K.; Hodgkinson, P.; Zorin, V.; Dumez, J.-N.; Elena-Herrmann, B.; Emsley, L.; Salager,
14 E.; Stein, R. S., Computation and NMR crystallography of terbutaline sulfate. *Magn. Reson. Chem.* **2010**,
15 *48*, S103-S112.
- 16 (75) Hanrahan, M. P.; Venkatesh, A.; Carnahan, S. L.; Calahan, J. L.; Lubach, J. W.; Munson, E. J.;
17 Rossini, A. J., Enhancing the Resolution of ^1H and ^{13}C Solid-State NMR Spectra by Reduction of
18 Anisotropic Bulk Magnetic Susceptibility Broadening. *Phys. Chem. Chem. Phys.* **2017**, *19*, 28153-28162.
- 19 (76) Salager, E.; Day, G. M.; Stein, R. S.; Pickard, C. J.; Elena, B.; Emsley, L., Powder Crystallography
20 by Combined Crystal Structure Prediction and High-Resolution ^1H Solid-State NMR Spectroscopy. *J.*
21 *Am. Chem. Soc.* **2010**, *132*, 2564-2566.
- 22 (77) Widdifield, C. M.; Robson, H.; Hodgkinson, P., Furosemide's one little hydrogen atom: NMR
23 crystallography structure verification of powdered molecular organics. *Chem. Commun.* **2016**, *52*, 6685-
24 6688.
- 25 (78) Widdifield, C. M.; Nilsson Lill, S. O.; Broo, A.; Lindkvist, M.; Pettersen, A.; Svensk Ankarberg, A.;
26 Aldred, P.; Schantz, S.; Emsley, L., Does Z' equal 1 or 2? Enhanced powder NMR crystallography
27 verification of a disordered room temperature crystal structure of a p38 inhibitor for chronic obstructive
28 pulmonary disease. *Phys. Chem. Chem. Phys.* **2017**, *19*, 16650-16661.
- 29 (79) Wood, P. A.; Allen, F. H.; Pidcock, E., Hydrogen-bond directionality at the donor H atom-analysis
30 of interaction energies and database statistics. *CrystEngComm* **2009**, *11*, 1563-1571.
- 31 (80) Samoson, A., Satellite transition high-resolution NMR of quadrupolar nuclei in powders. *Chem.*
32 *Phys. Lett.* **1985**, *119*, 29-32.
- 33 (81) Tatton, A. S.; Pham, T. N.; Vogt, F. G.; Iuga, D.; Edwards, A. J.; Brown, S. P., Probing
34 intermolecular interactions and nitrogen protonation in pharmaceuticals by novel ^{15}N -edited and 2D ^{14}N -
35 ^1H solid-state NMR. *CrystEngComm* **2012**, *14*, 2654-2659.
- 36 (82) Gervais, C.; Dupree, R.; Pike, K. J.; Bonhomme, C.; Profeta, M.; Pickard, C. J.; Mauri, F.,
37 Combined First-Principles Computational and Experimental Multinuclear Solid-State NMR Investigation
38 of Amino Acids. *J. Phys. Chem. A* **2005**, *109*, 6960-6969.
- 39 (83) Cavadini, S.; Lupulescu, A.; Antonijevic, S.; Bodenhausen, G., Nitrogen-14 NMR Spectroscopy
40 Using Residual Dipolar Splittings in Solids. *J. Am. Chem. Soc.* **2006**, *128*, 7706-7707.
- 41 (84) Kwan, I. C. M.; Mo, X.; Wu, G., Probing Hydrogen Bonding and Ion-Carbonyl Interactions by
42 Solid-State ^{17}O NMR Spectroscopy: G-Ribbon and G-Quartet. *J. Am. Chem. Soc.* **2007**, *129*, 2398-2407.
- 43 (85) O'Dell, L. A.; Schurko, R. W., Static solid-state ^{14}N NMR and computational studies of nitrogen
44 EFG tensors in some crystalline amino acids. *Phys. Chem. Chem. Phys.* **2009**, *11*, 7069-7077.
- 45 (86) Tatton, A. S.; Bradley, J. P.; Iuga, D.; Brown, S. P., ^{14}N - ^1H Heteronuclear Multiple-Quantum
46 Correlation Magic-Angle Spinning NMR Spectroscopy of Organic Solids. *Z. Phys. Chem.* **2012**, *226*,
47 1187.
- 48 (87) Cavadini, S.; Abraham, A.; Bodenhausen, G., Proton-detected nitrogen-14 NMR by recoupling of
49 heteronuclear dipolar interactions using symmetry-based sequences. *Chem. Phys. Lett.* **2007**, *445*, 1-5.
- 50 (88) Cavadini, S., Indirect detection of nitrogen-14 in solid-state NMR spectroscopy. *Prog. Nucl. Magn.*
51 *Reson. Sp.* **2010**, *56*, 46-77.

- 1 (89) Maruyoshi, K.; Iuga, D.; Antzutkin, O. N.; Alhalaweh, A.; Velaga, S. P.; Brown, S. P., Identifying
2 the intermolecular hydrogen-bonding supramolecular synthons in an indomethacin-nicotinamide cocrystal
3 by solid-state NMR. *Chem. Commun.* **2012**, *48*, 10844-10846.
- 4 (90) Nishiyama, Y.; Endo, Y.; Nemoto, T.; Utsumi, H.; Yamauchi, K.; Hioka, K.; Asakura, T., Very fast
5 magic angle spinning ^1H - ^{14}N 2D solid-state NMR: Sub-micro-liter sample data collection in a few
6 minutes. *J. Magn. Reson.* **2011**, *208*, 44-48.
- 7 (91) Geen, H.; Titman, J. J.; Gottwald, J.; Spiess, H. W., Solid-state proton multiple-quantum NMR
8 spectroscopy with fast magic angle spinning. *Chem. Phys. Lett.* **1994**, *227*, 79-86.
- 9 (92) Brown, S. P., Applications of high-resolution ^1H solid-state NMR. *Solid State Nucl. Magn. Reson.*
10 **2012**, *41*, 1-27.
- 11 (93) Harris, R. K.; Hodgkinson, P.; Pickard, C. J.; Yates, J. R.; Zorin, V., Chemical shift computations on
12 a crystallographic basis: some reflections and comments. *Magn. Reson. Chem.* **2007**, *45*, S174-S186.

13
14
15
16
17
18
19
20
21
22
23
24
25
26
27
28
29
30
31
32
33
34

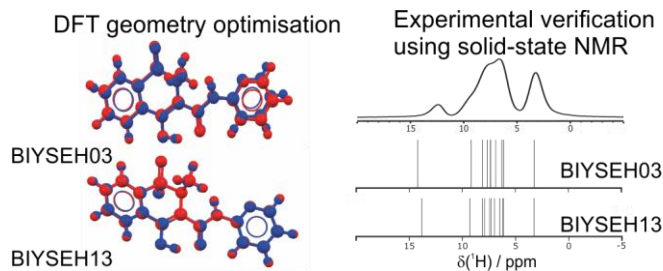
1 **“For Table of Contents Use Only”**

2 **Improving confidence in crystal structure solutions using NMR crystallography: the case of β -**
3 **piroxicam**

4

5 **Andrew S. Tatton^{a, *}, Helen Blade^b, Steven P. Brown^c, Paul Hodgkinson^d, Leslie P. Hughes^b, Sten Nilsson Lill^e,**
6 **and Jonathan R. Yates^a**

7



8

9

10 **Synopsis**

11 An NMR crystallography study is presented for two structural models of β -piroxicam, where one model
12 has a relatively poor R-factor. NMR crystallography verifies that these are comparable after geometry
13 optimisation and agree well with experiment. This demonstrates the capability of NMR crystallography to
14 verify and increase confidence in structural models with a degree of uncertainty in the precision and
15 accuracy.

16

17

18

19

20

21