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Hydroxypropyl methylcellulose and hydroxypropyl starch: Rheological and gelation effects on the phase structure of their mixed hydrocolloid system

Yanfei Wang, Long Yu, Qingjie Sun, Fengwei Xie

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- 1 Hydroxypropyl methylcellulose and hydroxypropyl starch:
- 2 Rheological and gelation effects on the phase structure of their
- 3 mixed hydrocolloid system
- 5 Yanfei Wang^{1,2,3}, Long Yu^{3,*}, Qingjie Sun^{1,2}, Fengwei Xie^{4,†}
- 6

- ⁷ ¹College of Food Science and Engineering, Qingdao Agricultural University, Qingdao, Shandong 266109,
- 8 China
- 9 ²Qingdao Special Food Research Institute, Qingdao, Shandong 266109, China
- ³College of Food Science and Engineering, South China University of Technology, Guangzhou, Guangdong
- 11 510640, China
- 12 ⁴International Institute for Nanocomposites Manufacturing (IINM), WMG, University of Warwick, Coventry
- 13 CV4 7AL, United Kingdom
- 14

^{*}Corresponding author. *E-mail address*: <u>felyu@scut.edu.cn</u> (L. Yu),

[†]Corresponding author. *E-mail addresses*: <u>d.xie.2@warwick.ac.uk</u>; <u>fwhsieh@gmail.com</u> (F. Xie)

15 Abstract

16 It is common to hybridize biopolymers for developing materials with combined properties or 17 functionality. However, biopolymers are usually not fully compatible despite their chemical 18 similarity, posing challenges to create mixed systems. Herein, we investigated how the gelation 19 behavior and rheological properties of hydroxypropyl methylcellulose (HPMC), a thermal gel, and 20 hydroxypropyl starch (HPS), a cooling gel, affect their miscibility and the phase structure of their mixed system. The dependence of the zero-shear viscosity for HPMC/HPS paste on biopolymer 21 concentration in a double-logarithmic coordinate can be divided into two parts with the slopes being 22 23 11.9 and 2.8 respectively, indicating different degrees of intermolecular entanglement. A typical "sea-island" morphology was shown in the blends, and the phase structure (continuous or discrete) 24 25 changed with varying HPMC/HPS blend ratio and temperature. This phase structure change can be 26 well correlated to rheological parameters such as zero-shear viscosity, loss tangent, complex 27 viscosity, and the Arrhenius equation correction coefficient (α). Biopolymer concentration, 28 HPMC/HPS ratio, and temperature together controlled rheological properties and phase distribution 29 for the mixed system. The relationship between rheological behavior and phase structure for 30 HPMC/HPS understood from this work provides an insight into designing mixed biopolymer 31 systems with desirable processability, structure, and properties. 32 *Keywords:* Hydroxypropyl methylcellulose; Hydroxypropyl starch; Rheological properties; 33 Biopolymer phase structure; Biopolymer blends

34

35 **1** Introduction

53

36	As a modified biopolymer, hydroxypropyl methylcellulose (HPMC) has been widely applied in
37	different applications such as edible packaging, medicinal capsules, and drug delivery systems due to
38	its water-solubility and biodegradability and excellent film-forming, mechanical and barrier
39	properties (Al-Tabakha, 2010; Muhammad-Javeed & Mohammed, 2018; Siepmann & Peppas, 2012).
40	However, the low oxygen-barrier property (Ghadermazi, Hamdipour, Sadeghi, Ghadermazi, & Asl,
41	2019; Hay, et al., 2018; Y. F. Wang, Yu, et al., 2016), high production energy consumption (Zhang,
42	et al., 2013), and high price of HPMC (Allenspach, Timmins, Sharif, & Minko, 2020) pose strong
43	limits for its applications. To cope with the property limitations of individual polymers, achieve
44	enhanced and/or new material properties, reduce costs, and to expand applications, mixing different
45	polymers is one of the most cost-effective methods (Aghjeh, Khonakdar, & Jafari, 2015; Botaro, de
46	Freitas, do Carmo, & Raimundo, 2020; Jéssica Bassi da Silva, Michael Thomas Cook, & Bruschi,
47	2020; Xu, Wang, & Shi, 2020). Hydroxypropyl starch (HPS), a typical chemically modified starch
48	with some original hydroxyl groups replaced with hydroxypropyl groups, is cheaper than HPMC,
49	show excellent gas barrier property, and has been widely used in the food industry, therefore being
50	an ideal substitute for part of HPMC (X. Chen, et al., 2019; J. Liu, Lai, Wang, Wang, & Liu, 2020;
51	Qin, et al., 2019; W. W. Wang, Sun, & Shi, 2019).
52	The production of edible materials such as pre-formed films (medicinal capsules) or edible

54 (Cuq, Gontard, & Guilbert, 1998; Suhag, Kumar, Petkoska, & Upadhyay, 2020). Specifically,

coating generally relies on wet processes that are based on film-forming solutions or dispersions

55 polymers are first dissolved or dispersed in a liquid and then dried. The design of such processes

56	(typically comprising pumping, casting, dipping, brushing, spraying, and drying) requires accurate
57	data on the rheological properties of film-forming solutions or dispersions (Q. Wang, Yang, Chen, &
58	Shao, 2012). In turn, rheological properties also play an important role in controlling the quality of
59	coating films (Peressini, Bravin, Lapasin, Rizzotti, & Sensidoni, 2003; Xiao, Tong, & Lim, 2012).
60	For thermodynamic reasons, most polymer blends are immiscible on a molecular scale.
61	Nevertheless, immiscible blends often turn out to be useful. For example, while polypropylene and
62	polystyrene are immiscible, a polypropylene 90/polystyrene 10 blend, which showed a phase
63	separation structure where polystyrene droplets were scattered in the polypropylene matrix, showed
64	the maximum impact strength that blends at other ratios (R. Y. Chen, Liu, Han, Zhang, & Li, 2020).
65	The degree of polymer miscibility largely influences the rheological properties of the mixed systems
66	(Tanaka, Sako, Hiraoka, Yamaguchi, & Yamaguchi, 2020; Wongphan & Harnkarnsujarit, 2020).
67	And, the rheological properties of polymer blends determine material phase structure (Ilyin,
68	Makarova, Polyakova, & Kulichikhin, 2020; Xiao, Tong, Zhou, & Deng, 2015).
69	Considering that both HPMC and HPS are water-soluble polysaccharides and consist of the same
70	original repeat unit (glucose with hydroxyl groups), there should be good compatibility between
71	them. However, HPMC and HPS are hydrogels with opposite temperature-induced gelation behavior.
72	Specifically, HPMC dissolves in water at a low temperature and congeals at a high temperature,
73	whereas HPS undergoes gelation on cooling but solates upon heating (Polamaplly, et al., 2019; Qin,
74	et al., 2019; Zhong, et al., 2020). The difference in gelation behavior between HPMC and HPS
75	makes it challenging to achieve a blend of these two biopolymers with high miscibility and fine
76	phase distribution. Moreover, the phase structure of HPMC and HPS should influence the mixed gel

77 properties. The rheological properties of HPMC/HPS blends depend on several factors such as 78 biopolymer concentration, mixing ratio, temperature, and shear stress applied (Y. F. Wang, Zhang, et 79 al., 2016). The manipulation of these factors can be used to further explore the relationship between 80 microstructure and properties for thermal-/cooling-gel mixed systems. In this work, the effects of solution concentration, mixing ratio, and temperature on the 81 82 rheological properties and morphology of HPMC/HPS blends were investigated. A schematic model 83 is proposed to describe temperature-induced phase structure changes for HPMC/HPS blends. The 84 knowledge obtained from this work could also help in understanding the relationship between 85 microstructure and processability for mixed systems based on biopolymers with different gelation behavior. The results could also be insightful for the design of similar biopolymer composite 86 87 materials with tailored structure and properties.

88 2 Materials and method

89 2.1 Materials

A commercially available pharmaceutical-grade HPMC (HT-E15; viscosity at 2% concentration:
6.3 mPas; pH 6.0; methoxyl content on dry basis: 29%; hydroxypropyl oxygen content on dry basis
8.4%) was purchased from Huzhou Hopetop Pharmaceutical Co., Ltd (China). A food-grade maize
HPS with a degree of substitution (DS) of 0.11 was supplied by Penford (Australia).

94 **2.2 Sample preparation**

HPMC/HPS solutions with different total biopolymer concentration (5–20 wt%) and weight ratio
(0:10 to 10:0, w/w) were prepared. Specifically, HPMC and HPS in the form of dry powder were

97	firstly mixed and then dispersed in hot water (70 $^{\circ}$ C) with stirring for 30 min to ensure their proper
98	dispersion. Then, using a water bath, the solutions were heated to 95 °C and maintained for 1 h with
99	stirring to gelatinize HPS. Afterwards, the solutions were cooled to room temperature to dissolve
100	HPMC under stirring before testing.
101	2.3 Rheological measurement
102	The rheological properties of HPMC/HPS pastes were investigated using a Discovery HR-2
103	rheometer (TA Instruments, New Castle, DE, USA). A parallel-plate geometry (40 mm diameter)
104	with a gap of 0.5 mm was used for measurement.
105	To study the stability of HPMC/HPS pastes under shear, the samples were subjected to steady
106	shear at a constant high shear rate of 800 s ^{-1} for 2500 s at 25 °C and the change in viscosity was
107	recorded.
108	Under steady shear, the viscosity of a polymer may change with time before a stable value is
109	achieved (Tajuddin, Xie, Nicholson, Liu, & Halley, 2011). As a result, pre-shearing with a shear rate
110	of 800 s ⁻¹ at room temperature (25 °C) for 1000 s was performed to ensure all samples to achieve a
111	stable rheological state before immediate measurement. Shear viscosity was measured as a function
112	of shear rate in the range of 10^{-2} – 10^3 s.
113	Three-interval thixotropy testing was performed to investigate the stability of HPMC/HPS pastes
114	This test was carried out at 25 °C at a low-shear stage with the shear rate kept at 1 s ^{-1} for 50 s, then a
115	high-shear stage with the shear rate kept at 1000 s^{-1} for 20 s, and finally, a structural recovery stage

116 with the shear rate kept at 1 s^{-1} for 250 s.

117	Temperature sweeps were carried out from 5 °C to 85 °C with a heating rate of 2 °C/min under
118	dynamic mode. The frequency was set at 1 Hz and the strain at 0.1% (to be in the linear range of
119	viscoelasticity). The sample was placed between the parallel plates, and then a small amount of
120	silicone oil was applied to the periphery of the sample to prevent moisture evaporation.
121	2.4 Microscopy observation and method of dying HPS
122	An Olympus BHZ-UMA optical microscope was used to image the morphology of HPMC/HPS
123	pastes. The HPMC/HPS (5:5, w/w) solution of a concentration of 3 wt% was prepared using the
124	same method mentioned above. The glass and the solution were kept at the same testing temperature
125	and then the films were prepared by casting the solutions on glass at different temperatures (25 °C,
126	45 °C, and 85 °C). HPS was dyed with 1% iodine alcohol solution (prepared by mixing 1g of iodine
127	and 10 g of potassium iodine solution in a 100 mL flask, with alcohol subsequently added) and dried
128	at the same testing temperature.

129 **3** Results and discussions

130 **3.1 Time-dependence of HPMC/HPS paste viscosity**

Fig. 1 shows the viscosity *vs*. time curves for HPMC/HPS blends with different selected blending ratio and concentrations at a constant shear rate of 800 s⁻¹ (not all results presented here, as the trend was quite similar). For all blends, the viscosity decreased with time until a stable value was achieved. The time needed for achieving a stable state varied with HPMC/HPS ratio and total biopolymer concentration. After 1000 s, all the samples reached constant viscosity. Therefore, pre-shearing at 800 s⁻¹ for 1000 s was performed for rheological tests for all the samples.



Fig. 1 Viscosity as a function of time for HPMC/HPS pastes. The shear rate was fixed at 800 s⁻¹.

140

141 **3.2** Effect of biopolymer concentration on HPMC/HPS paste viscosity

- Fig. 2(A) shows the viscosity *vs.* shear rate curves for HPMC/HPS pastes of different concentration. At a certain shear rate, increasing biopolymer concentration resulted in higher viscosity, which is as expected. For all HPMC/HPS pastes, the viscosity decreased with increasing shear rate, suggesting shear-thinning behavior. Moreover, the dependence of viscosity on shear rate depended on total biopolymer concentration. The shear-thinning behavior was more evident with a higher biopolymer concentration.
- 148



150 Fig. 2 A) Flow curves for 50:50 (w/w) HPMC/HPS pastes of different total biopolymer

151 concentration at 25 °C; B) Zero-shear viscosity as a function of total biopolymer concentration, with

152 the molecular conformation of HPMC/HPS schematically shown (the HPMC/HPS ratio was 50:50

- 153 (w/w), red lines represent HPMC chains and grey lines HPS).
- 154

155 The viscosity *vs.* shear rate curves for HPMC/HPS pastes were fitted with the Carreau model 156 (Carreau, Pierre, & J., 1972) and then extrapolated to obtain zero-shear viscosity, η_0 (0.9969 < R^2 < 157 0.9997). **Fig. 2(B)** shows zero-shear viscosity as a function of total biopolymer concentration. It is 158 noticeable that η_0 had a power-law dependence on biopolymer concentration as represented by the 159 equation below:

 $\eta_0 = kC^m$

161 Where η_0 is the zero-shear viscosity, C is the total biopolymer concentration, k and m are constants. The dependence of η_0 on biopolymer concentration in a double-logarithmic coordinate can be 162 163 divided into two parts with the slopes (*m*) being 11.9 and 2.8, respectively. The critical concentration (C^*) that divides the two regions was about 8 wt%. According to the general relationship between 164 165 biopolymer concentration dependence and the polymer state in solution (Colby, 2010), the likely 166 molecular conformation of HPMC/HPS blends was proposed schematically in the inset of Fig. 2(B). 167 As the lowest concentration (5 wt%) in our work is still much higher than those used in previous studies (0.05–1.3 wt%) (Morris, Cutler, Ross-Murphy, Rees, & Price, 1981; Pakravan, Heuzey, & 168 169 Ajji, 2011; Tan, Li, Chen, & Xie, 2016), likely, the samples in both regions were in a concentrated 170 state. In the first region ($C < C^*$), gelatinized HPS thickened at a low temperature and forms 171 microgel, which interacts with HPMC chains at the microgel particle edge. HPS microgel particles

172	and HPMC chains tended to interact or entangle with each other due to the space-block effect and		
173	hydrogen bonding (inset a of Fig. 2B). The η_0 of HPMC/HPS paste increased rapidly with increasing		
174	concentration. In the second region ($C > C^*$), the interactions and entanglements were already denes		
175	enough (inset b of Fig. 2B) and their enhancement by increasing concentration was not as effective		
176	as in the first region. Thus, the slope for the second region was not as high as the first region.		
177	For a polymer solution, the relationship between shear stress (τ) and shear rate (γ) can be		
178	described by the Ostwald-de Waele equation:		
179	$\tau = K\gamma^n \tag{2}$		
180	Where K is the fluid consistency index and n is the flow behavior index. The values of K and n		
181	calculated for HPMC/HPS pastes of different concentration are shown in Table 1.		
182	For a Newtonian fluid, <i>n</i> equals 1. Pseudoplastic fluids have $n < 1$, and greater deviation of <i>n</i>		
183	from 1 indicates stronger pseudoplastic (shear-thinning) behavior. Table 1 shows that for all the		
184	samples, n is less than 1, indicating they were all pseudoplastic (shear-thinning). For HPMC/HPS		
185	pastes of low concentration (e.g. 5 wt%), the <i>n</i> values are close to 1, meaning they were more like a		
186	Newtonian fluid as HPMC chains and HPS microgel particles are mostly separated in the solution.		
187	Increasing concentration led to lower <i>n</i> , suggesting stronger shear-thinning behavior. The flow		
188	behavior of high-concentration samples is attributed to the interaction and entanglement among		
189	HPMC/HPS chains.		
190			
191	Table 1 Flow behavior index (<i>n</i>) and fluid consistency index (<i>K</i>) for 50:50 (w/w) HPS/HPMC pastes		
192	of different concentration at 25 °C.		

 R^2 $K(\operatorname{Pa} \cdot \mathbf{s}^n)$ Concentration (%) п

5	0.922 ± 0.004 ^a	0.06±0.00	1.0000
6	0.909±0.001	0.09±0.00	1.0000
7	0.859 ± 0.002	0.20±0.01	1.0000
8	0.842±0.001	0.28±0.01	0.9999
10	0.747±0.003	1.15±0.05	0.9981
12	0.735±0.005	2.34±0.07	0.9988
15	0.681±0.006	6.27±0.94	0.9993
18	0.678 ± 0.006	13.37±0.88	0.9991
20	0.656 ± 0.000	18.06±0.54	0.9984
22	0.653±0.012	24.40±2.31	0.9979

193

^a Mean \pm standard deviation.

194

For low-concentration samples, *K* was small. Higher concentration resulted in higher *K*, whichcan be linked to higher viscosity, as expected.

197 3.3 Effect of HPMC/HPS ratio on HPMC/HPS paste viscosity

Fig. 3(A) shows the effect of HPMC/HPS ratio on the viscosity of HPMC/HPS pastes. At a low HPS ratio (< 20%), the viscosity was not significantly influenced by shear rate, which could be ascribed to dispersed biopolymer chains. In contrast, the samples with a high HPS ratio showed significantly lower viscosity with increasing shear rate, a typical shear thinning behavior. At a certain shear rate, a higher HPS ratio led to higher viscosity. This corresponds to the fact that HPS is a gel with high viscosity at a low temperature (25 °C).



Fig. 3 A) Viscosity *vs.* shear rate curves for HPMC/HPS pastes with different HPS ratio at 25 °C; B) Zero-shear viscosity η_0 , with the solid line indicating measured values and the dashed line predicted values; C) Flow behavior index *n*; and D) flow consistency index *K* for HPMC/HPS pastes with different HPS ratio. The total concentration of HPMC/HPS was 15 wt%.

213

Table 2 lists the *n* and *K* values for HPS/HPMC pastes with different mixing ratio. With
increasing HPS ratio, *n* reduced gradually, while *K* showed a rising trend, implying that the addition
of HPS made the HPMC paste more viscous and difficult to flow. This result is consistent with a
previous study (Zhang, et al., 2015).

218

Table 2 Flow behavior index (*n*) and fluid consistency index (*K*) of HPS/HPMC solutions with
different HPS/HPMC ratio at 25 °C. The total biopolymer concentration was fixed at 15 wt%.

HPMC/HPS	п	K (Pa·s ⁿ)	R^2
10:0	0.9511±0.0023	0.84±0.01	0.9999
9:1	0.9280±0.0096	1.09±0.10	0.9998
8:2	0.8915±0.0146	1.41±0.16	0.9994
7:3	0.8423±0.0135	1.94±0.31	0.9998
6:4	0.7170±0.0094	4.76±0.20	0.9996
5:5	0.6807±0.0063	6.27±0.94	0.9993
4:6	0.6666±0.0032	8.33±0.41	0.9969
3:7	0.5835±0.0028	14.91±0.40	0.9942
2:8	0.5068±0.0037	26.93±1.39	0.9869
1:9	0.4694±0.0008	41.76±0.64	0.9897
0:10	0.4687±0.0019	45.01±0.72	0.9935

221

For a homogeneous system, the relationship among the rheological properties of the blend and its individual components follow a logarithmic sum rule (Utracki, 1983). For a binary system, this rule can be expressed as:

225

 $\log F = \phi_1 \log F_1 + \phi_2 \log F_2 \tag{3}$

where F, F_1 , and F_2 are the rheological parameter of the mixture, component 1, and component 2,

227 respectively; ϕ_1 and ϕ_2 are the mass fractions of component 1 and component 2, respectively,

228 with $\phi_1 + \phi_2 = 1$.

229 The measured and predicted curves of η_0 as a function of HPS ratio are shown in **Fig. 3(B)**. The 230 zero-shear viscosity increased with HPS ratio and their relationship generally follows the log-

231	additivity rule, implying good compatibility of this system. Nonetheless, the measured values deviate
232	from the mixing rule either positively or negatively, depending on HPS ratio. This suggests the
233	mixed system had a continuous-discrete type of phase structure and the change of continuous phase
234	occurred (Yao, Mukuze, Zhang, & Wang, 2013) at an HPS ratio of 60%. Regarding the negative
235	deviation at a low HPS ratio, HPMC chain existed as a continuous phase in which HPS microgel was
236	scattered. Regarding positive deviation at a high HPS ratio, HPMC became a separated phase
237	scattered in the HPS continuous phase. These results are confirmed by microscopy observation with
238	a previous study (Y. F. Wang, Zhang, et al., 2016).
239	Fig. $3(C)$ shows that <i>n</i> decreased progressively with increasing HPS ratio and the relationship
240	between n and HPS ratio follows the linear-additivity rule. This suggests that addition of HPS to
241	HPMC imparts stronger pseudoplastic (shear-thinning) behavior. The linear regression analysis was
242	achieved with $R^2 = 0.98062$, denoting good compatibility between the two biopolymers.
243	Fig. $3(D)$ shows that increasing HPS ratio resulted in higher K, related to the gelation behavior of
244	HPS at a low temperature. For HPMC/HPS pastes with an extreme blend ratio (HPS ratio < 20 wt%
245	or > 70 wt%), the K value for a blend paste is closer to that for the biopolymer with a high ratio in
246	that mixture. In this regard, the viscosity of a mixture paste is mainly determined by the continuous
247	phase. On the other hand, for HPMC/HPS pastes with an intermediate ratio (20 wt% < HPS ratio <
248	70 wt%), although HPMC with low viscosity still existed as a continuous phase, K rose rapidly with
249	increasing HPS ratio, indicating that HPS as the separated phase made a major contribution to the
250	viscosity of the blends. This implies that the continuous phase and the separated phase make
251	different contributions to the viscosity of a blend depending on the ratio of the two components.

252 **3.4** Thixotropic behavior of HPMC/HPS pastes

- 253 Thixotropy describes the structural stability of a material against shear applied (Krystyjan, et al.,
- 254 2016; Shakeel, Kirichek, & Chassagne, 2020; Marek Sikora, Adamczyk, & Krystyjan, 2011).
- 255 Thixotropy can be linked to time and the history of shearing that results in microstructural changes
- 256 (Czaikoski, da Cunha, & Menegalli, 2020; Mewis & Wagner, 2009; M. Sikora, et al., 2015). Fig. 4
- shows the results of the three-interval thixotropic behavior of HPMC/HPS pastes with different
- 258 mixing ratio. All the samples were thixotropic. The viscosity at a low shear rate (1 s^{-1} , in the first and
- third stages) increased significantly with a higher HPS ratio.

260



261



²⁶³ °C. The total biopolymer concentration was 15 wt%.

264

265 The structural recovery ratio (DSR) can be defined by the equation:

$$266 \qquad \text{DSR} = \frac{\eta_t}{n} \times 100\% \qquad (4)$$

267	where η_t is the viscosity in the structural recovery stage (the third stage) at a certain time (<i>t</i>) and η is
268	the final viscosity in the first stage (Mewis & Wagner, 2011; Toker, Karasu, Yilmaz, & Karaman,
269	2015).
270	With $DSR < 100\%$, it can be assumed that the recovered paste was less resistant to shear and the
271	material is thixotropic. In contrast, DSR > 100% indicates that the sample is anti-thixotropic. In a
272	word, for thixotropic material, a higher DSR value means weaker thixotropic behavior (higher
273	shear-resistance) and higher rheological stability.
274	The DSR data from three-interval thixotropic tests were listed in Table 3. The pure-HPMC
275	solution showed a very high DSR value, close to 100%. In this regard, HPMC chains are rigid with a
276	short relaxation time, which means the structure can recover quickly. In contrast, the pure-HPS
277	solution presented a low DSR value. This indicates that the structural recovery of HPS is slow and
278	can be explained by HPS chains being flexible and their long relaxation time.
• - •	

279

Table 3 Degree of structure recovery (DSR) at certain recovery time for HPMC/HPS solutions with
 different HPMC/HPS ratio at 25 °C. The total biopolymer content was 15 wt%.

DSR (%) HPMC/HPS ratio 10 s 60 s 250 s $91.29{\pm}1.70^{\ a}$ 10:0 93.57±0.65 98.98 ± 0.37 9:1 71.66±1.66 63.26 ± 2.12 66.67 ± 2.18 8:2 39.32 ± 0.95 42.94 ± 0.95 45.78 ± 1.30 7:3 20.43 ± 0.62 22.87±0.75 24.76 ± 0.77 $30.81{\pm}0.27$ 24.68 ± 0.43 31.83 ± 0.53 6:4

0:10	38.29±0.34	45.31±0.45	38.65±0.19
1:9	67.60±0.16	72.78±0.76	72.74±1.01
2:8	57.58±0.97	58.86±0.98	59.40±0.59
3:7	59.98±1.90	53.23±1.84	50.36±1.67
4:6	44.19±0.90	41.75±0.78	38.98±0.81
5:5	37.00±1.44	36.97±1.03	34.03±0.91

282	^a Mean ± standard deviation
283	
284	While the DSR values for mixture pastes were lower than that for the pure-HPMC sample but
285	higher than that for the pure-HPS sample, the change in DSR did not follow a consistent trend. For
286	mixtures with a low HPS ratio (< 30 wt%), DSR decreased with increasing HPS ratio, indicating
287	enhanced thixotropic behavior. In this regard, flexible HPS chains in the system counteracted the
288	effect of rigid HPMC chains to some extent, thus decreasing the capability of the structure to recover
289	rapidly from shear-induced deformation and prolonging the relaxation time. However, further
290	increasing HPS content (> 30 wt%) led to an increase in DSR, suggesting weakened thixotropic
291	behavior. In this regard, the interaction between HPMC and HPS chains might result in improvement
292	in overall chain rigidity and thus a reduction in relaxation time.
293	The DSR values with different recover time are also listed in Table 3. HPMC/HPS pastes almost
294	finished the structural recovery within 10 s as the DSR were very close to the final DSR (at 250 s).
295	For some samples, the DSR values at 60 s were even high than the final DSR. Given this, these
296	samples were more unstable as their structure could be destroyed even at very low shear rate.

297	3.5 Effect of temperature on the dynamic rheological properties and phase structure of
298	HPMC/HPS pastes
299	As a wide range of temperature could be applied during the processing and storage of
300	biopolymers, it is important to understand the effect of temperature on rheological properties (Park,
301	Chung, & Yoo, 2004; Rao, 2014). Both HPMC and HPS are temperature-sensitive hydrogels and
302	their opposite gelling behavior make it difficult to achieve a blend with high miscibility, despite
303	these polysaccharides are chemically akin. Nevertheless, at a suitable mixing ratio and under a
304	certain temperature, HPMC and HPS could interact with each other to the maximum extent.
305	3.5.1 Gelation behavior of HPMC/HPS pastes studied by dynamic viscoelasticity
306	The viscoelastic properties of HPMC/HPS pastes with different mixing ratio are shown in Fig. 5
307	(A). The pure-HPMC paste showed liquid-like behavior at a low temperature as G' was below G'' ,
308	while it congealed at a high temperature with G' surpassing G'' . For the pure-HPS sample, the three-
309	dimensional gel network at a low temperature solated at a high temperature, inferred from $G' < G''$.
310	The cross point of G' and G'' was at about 49 °C for the pure-HPMC sample and 70 °C for the
311	pure-HPS sample, confirming HPMC being a thermal gel and HPS being a cooling-gel.
312	



Fig. 5 A) Storage modulus (G'), loss modulus (G'') and loss tangent (tan δ) vs. temperature curves for 314 315 the pure-HPMC paste, the 5:5 (w/w) HPMC/HPS paste, and the HPS paste, respectively, all having a 316 total biopolymer concentration of 15 wt%; B) Complex viscosity vs. temperature curves for the 317 HPMC paste, the 5:5 (w/w) HPMC/HPS paste, and the HPS paste, respectively, all having a total biopolymer concentration of 15 wt%; C) Light-microscopic images of the dyed 5:5 (w/w) 318 319 HPMC/HPS paste of 3 wt% total biopolymer concentration at 25°C, 45°C, and 85 °C, respectively. 320

For the 5:5 (w/w) HPMC/HPS paste, the G' and G'' curves were roughly parallel to those for the 321 322 pure-HPS paste at a low temperature and to those for the pure-HPMC paste at a high temperature.

Given this, HPS had a major influence on the viscoelastic properties of the mixture at a low temperature because HPS mainly contributed to the gelation at a low temperature, whereas HPMC dominated the mixture behavior at a high temperature when HPMC was responsible for gelation. The blend had intermediate moduli between those for individual HPS and HPMC. Besides, *G'* was always higher than *G''* during the whole temperature range, indicating solid-like behavior contributed by biopolymer chain interactions. Furthermore, the 5:5 (w/w) HPMC/HPS paste showed a tan δ peak at about 45 °C, at which point a change of the continuous phase occurred.

330 3.5.2 Effect of temperature on the complex viscosity of HPMC/HPS pastes

For mixed systems, the viscosity of individual components is important. Normally, when two components with largely different viscosity were mixed, the component with a higher viscosity tends to form a scattered phase in the continuous phase of the lower-viscosity component.

334 Fig. 5(B) shows the effect of temperature on the complex viscosity (η^*) in the range of 5–85 °C of HPMC/HPS pastes. With increasing temperature, the η^* of the pure-HPS sample showed a slight 335 336 decrease followed by a sharp decrease. However, the η^* of the pure-HPMC sample first experienced 337 a slight decline and then a strong rise starting at 45 °C. For the 5:5 (w/w) HPMC/HPS paste, η^* first 338 decreased slightly and then increased moderately, a changing pattern close to that for HPS at a low 339 temperature but similar to that for HPMC at a high temperature. This result should be ascribed to the 340 different gelation behavior of HPS (cooling gel) and HPMC (thermal gel). The rapid change in η^* of 341 the 5:5 (w/w) mixture at about45 °C might be due to the change in the phase distribution of 342 HPMC/HPS.

343 The temperature dependence of η^* for HPMC/HPS pastes at a certain temperature can be 344 evaluated using an Arrhenius-type equation:

345
$$\eta^* = \eta_i \exp\left(\alpha \frac{E}{RT}\right)$$
 (5)

346	where η^* is complex viscosity, η_i is a constant describing the viscosity coefficient at a reference
347	temperature (Pa·s) (Laity & Holland, 2017; Qing, Jinsong, Changjiang, Jianpeng, & Shouqin, 2014a),
348	<i>T</i> is the absolute temperature (K), <i>R</i> is the gas constant (8.3144 J·mol ⁻¹ ·K ⁻¹), α is the correction
349	coefficient (±1), and <i>E</i> is the activation energy ($J \cdot mol^{-1}$).
350	For fitting using Eq.(5), the curves need to be divided into two sections before and after 45 $^{\circ}$ C
351	according to viscosity curves shown in Fig. 5(B). Table 4 presents the values of <i>E</i> , α , and η_i
352	calculated based on Eq.(5) for the temperature ranges of 10–45 °C and 45–85 °C ($R^2 > 0.90$). It can
353	be seen that the η_i value for the pure-HPMC sample was very small $(1.31 \times 10^{-5} \text{ Pa} \cdot \text{s})$ at a low
354	temperature but high (1.99×10^{28} Pa·s) at a high temperature. In contrast, for the pure-HPS sample,
355	the η_i value at a low temperature (0.56 Pa·s) was much higher than that at a high temperature
356	(1.78×10 ⁻¹⁹ Pa·s). This difference in η_i between the two biopolymers could be ascribed to their
357	different gelation behavior, namely, HPS is a cooling-gel and HPMC is a thermal-gel. For the 5:5
358	(w/w) blend, the η_i value was similar to that for the pure-HPS paste and much higher than that for the
359	pure-HPMC at a low temperature, and between that for the pure-HPS sample and that for the
360	pure-HPMC sample at a high temperature. This implies that HPS played a dominant role in the blend
361	at a low temperature, while HPMC had a much stronger effect at a high temperature.
362	
363	Table 4 Arrhenius equation parameters (correction coefficient, α ; activation energy, <i>E</i> ; constant, η_i ;
364	and determination coefficient, R^2) for the HPS paste, the 5:5 (w/w) HPS/HPMC paste, and the
365	HPMC paste. The total biopolymer content is 15 wt%.

Sample	5–45 °C	45–85 °C

	$\eta_{\rm i} ({\rm Pa} \cdot {\rm s})$	α	$E (kJ \cdot mol^{-1})$	R^2	$\eta_{\rm i} ({\rm Pa} \cdot {\rm s})$	α	$E (kJ \cdot mol^{-1})$	R^2
НРМС	1.31×10 ⁻⁵	+1	27.7	0.9917	1.99×10 ²⁸	-1	174	0.9441
HPMC-HPS	0.47	+1	8.09	0.9219	5.67×10 ¹²	-1	72	0.9688
HPS	0.56	+1	10.7	0.9071	1.78×10^{-19}	+1	124	0.9744

366

367 The temperature affects biopolymer paste viscosity in different ways:

a) It is known that a higher temperature leads to greater chain mobility and lower viscosity.

369 Based on this effect (effect A), the viscosity is inversely proportional to temperature and α is 370 a negative value.

b) For a thermal gel like HPMC, increasing temperature is favorable for greater interaction

between hydrophobic groups and that between hydrophilic hydroxyl groups, thus enhancing

the three-dimensional network and increasing the viscosity. Based on this effect (effect B),

374 the viscosity is proportional to temperature and α is a positive value (Qing, Jinsong,

375 Changjiang, Jianpeng, & Shouqin, 2014b).

c) For a cooling gel like HPS, a higher temperature causes the breakage of intermolecular

377 hydrogen bonds and thus the three-dimensional network, leading to declined viscosity. Based

378 on this effect (effect C), the viscosity is inversely proportional to temperature and α is a

379 negative value.

In the low-temperature region (5–45 °C), only effect A existed for the pure-HPMC paste since it was in a liquid-like state and therefore, *E* was high and $\alpha = +1$. For the pure-HPS paste, which was a solid-like gel, there were both effects A and C, while effect A was greater than effect C; thus, $\alpha = +1$ and *E* was lower than that for the pure-HPMC sample. For the 5:5 (w/w) HPMC/HPS paste, both

384	effects A and C presented here, but effect C was weakened by HPMC due to its liquid-like state and
385	thus, $\alpha = +1$ and <i>E</i> was lower than that for the pure-HPMC sample. In a word, HPS played the
386	dominant role in controlling the phase structure and rheological behavior at a low temperature.
387	In the high-temperature region (45–85 °C), both effects A and B existed for HPMC since HPMC
388	underwent gelation at a high temperature, with effect A << effect B; thus, E was very high and α =
389	-1. For HPS, there was only effect A as HPS solates at a high temperature and therefore, E was very
390	high and $\alpha = +1$. For the 5:5 (w/w) mixture paste, there were both effects A and B but effect B was
391	lessened by HPS which was liquid-like; therefore, $\alpha = -1$ and <i>E</i> was lower than that for the pure-
392	HPMC sample. These results indicate that HPMC played the leading role in controlling the structure
393	and rheological behavior of the blend at a high temperature.
394	3.5.3 Effect of temperature on the phase structure of HPMC/HPS pastes
395	Fig. 5(C) shows the morphologies of the 5:5 (w/w) HPMC/HPS blends at 25 °C, 45 °C, and
395 396	Fig. 5(C) shows the morphologies of the 5:5 (w/w) HPMC/HPS blends at 25 °C, 45 °C, and 85 °C observed using an optical microscope. Two phases could be identified, the HPMC phase in
395 396 397	Fig. 5(C) shows the morphologies of the 5:5 (w/w) HPMC/HPS blends at 25 °C, 45 °C, and 85 °C observed using an optical microscope. Two phases could be identified, the HPMC phase in light and the HPS phase in dark. Under light microscopy, HPS becomes dark after being dyed with
395 396 397 398	Fig. 5(C) shows the morphologies of the 5:5 (w/w) HPMC/HPS blends at 25 °C, 45 °C, and 85 °C observed using an optical microscope. Two phases could be identified, the HPMC phase in light and the HPS phase in dark. Under light microscopy, HPS becomes dark after being dyed with iodine (Y. F. Wang, Zhang, et al., 2016). Increasing temperature resulted in an increased dark area
395 396 397 398 399	Fig. 5(C) shows the morphologies of the 5:5 (w/w) HPMC/HPS blends at 25 °C, 45 °C, and 85 °C observed using an optical microscope. Two phases could be identified, the HPMC phase in light and the HPS phase in dark. Under light microscopy, HPS becomes dark after being dyed with iodine (Y. F. Wang, Zhang, et al., 2016). Increasing temperature resulted in an increased dark area (HPS phase) but a reduced light area (HPMC phase). At 25 °C, HPMC (in light color) was shown to
 395 396 397 398 399 400 	Fig. 5(C) shows the morphologies of the 5:5 (w/w) HPMC/HPS blends at 25 °C, 45 °C, and 85 °C observed using an optical microscope. Two phases could be identified, the HPMC phase in light and the HPS phase in dark. Under light microscopy, HPS becomes dark after being dyed with iodine (Y. F. Wang, Zhang, et al., 2016). Increasing temperature resulted in an increased dark area (HPS phase) but a reduced light area (HPMC phase). At 25 °C, HPMC (in light color) was shown to be a continuous phase, with the scattering of small spherical HPS domains (dark). In contrast, at
 395 396 397 398 399 400 401 	Fig. 5(C) shows the morphologies of the 5:5 (w/w) HPMC/HPS blends at 25 °C, 45 °C, and 85 °C observed using an optical microscope. Two phases could be identified, the HPMC phase in light and the HPS phase in dark. Under light microscopy, HPS becomes dark after being dyed with iodine (Y. F. Wang, Zhang, et al., 2016). Increasing temperature resulted in an increased dark area (HPS phase) but a reduced light area (HPMC phase). At 25 °C, HPMC (in light color) was shown to be a continuous phase, with the scattering of small spherical HPS domains (dark). In contrast, at 85 °C, the HPMC phase became quite small and scattered in the HPS continuous phase.
 395 396 397 398 399 400 401 402 	Fig. 5(C) shows the morphologies of the 5:5 (w/w) HPMC/HPS blends at 25 °C, 45 °C, and 85 °C observed using an optical microscope. Two phases could be identified, the HPMC phase in light and the HPS phase in dark. Under light microscopy, HPS becomes dark after being dyed with iodine (Y. F. Wang, Zhang, et al., 2016). Increasing temperature resulted in an increased dark area (HPS phase) but a reduced light area (HPMC phase). At 25 °C, HPMC (in light color) was shown to be a continuous phase, with the scattering of small spherical HPS domains (dark). In contrast, at 85 °C, the HPMC phase became quite small and scattered in the HPS continuous phase. There should be a certain temperature at which the viscosity of HPMC and HPS became similar,
 395 396 397 398 399 400 401 402 403 	Fig. 5(C) shows the morphologies of the 5:5 (w/w) HPMC/HPS blends at 25 °C, 45 °C, and 85 °C observed using an optical microscope. Two phases could be identified, the HPMC phase in light and the HPS phase in dark. Under light microscopy, HPS becomes dark after being dyed with iodine (Y. F. Wang, Zhang, et al., 2016). Increasing temperature resulted in an increased dark area (HPS phase) but a reduced light area (HPMC phase). At 25 °C, HPMC (in light color) was shown to be a continuous phase, with the scattering of small spherical HPS domains (dark). In contrast, at 85 °C, the HPMC phase became quite small and scattered in the HPS continuous phase. There should be a certain temperature at which the viscosity of HPMC and HPS became similar, thus leading to a change in the phase distribution (continuous or discrete) of the two biopolymers.
 395 396 397 398 399 400 401 402 403 404 	Fig. 5(C) shows the morphologies of the 5:5 (w/w) HPMC/HPS blends at 25 °C, 45 °C, and 85 °C observed using an optical microscope. Two phases could be identified, the HPMC phase in light and the HPS phase in dark. Under light microscopy, HPS becomes dark after being dyed with iodine (Y. F. Wang, Zhang, et al., 2016). Increasing temperature resulted in an increased dark area (HPS phase) but a reduced light area (HPMC phase). At 25 °C, HPMC (in light color) was shown to be a continuous phase, with the scattering of small spherical HPS domains (dark). In contrast, at 85 °C, the HPMC phase became quite small and scattered in the HPS continuous phase. There should be a certain temperature at which the viscosity of HPMC and HPS became similar, thus leading to a change in the phase distribution (continuous or discrete) of the two biopolymers. We observed this transition at 45 °C (Fig. 5C) when the typical "sea-island" morphology in the

- 406 agrees with the transition indicated by the tan δ peak and the rapid change in viscosity for the 5:5 407 (w/w) mixture at 45 °C.
- 408 **3.6** Temperature-induced structural changes of HPMC/HPS solutions
- 409 Based on the comparison of HPMC/HPS blends discussed above in the context of the classic
- 410 rheological behavior of polymer solution and hybrid gels (Ford, 1999; Rachmawati, Woortman, &
- 411 Loos, 2013; Zhang, et al., 2015), we propose a schematic representation of the temperature-induced
- 412 conformational change of HPMC/HPS pastes, as shown in **Fig. 6**.
- 413 The gelation behavior of HPMC has been reported and the related mechanism has been discussed
- 414 (Haque & Morris, 1993; Haque, Richardson, Morris, Gidley, & Caswell, 1993; S. Q. Liu, Joshi, Lam,
- 415 & Tam, 2008; Viriden, Larsson, Schagerlof, & Wittgren, 2010; Y. F. Wang, et al., 2018).
- 416 Specifically, it is recognized that HPMC chains exist in solution as aggregated bundles. These
- 417 bundles are held together by the packing of the un-substituted or the sparingly soluble regions of the
- 418 cellulosic structure and by the hydrophobic clustering of methyl groups and hydroxyl groups in
- 419 regions with denser substitution. These packings and clusters are the so-called water cages and shells,
- 420 preventing the formation of inter-chain hydrogen bonding at a low temperature. The absorption of
- 421 heat can break the water cages and shells, which is kinetic of the sol-gel transition. The breakage of
- 422 the water cages and shells exposes methyl and hydroxyl groups to the surrounding water and causes
- 423 a significant increase in volume. At a higher temperature, due to both the interaction among
- 424 hydrophobic groups and that among hydroxyl groups, a cross-linked network is formed (**Fig. 6a**).
- 425 Amylose, exuded from starch granules after gelatinization, tended to coil to form hollow, left-
- 426 handed single helices coaxially twisting along the chains. These helices feature a hydrophobic cavity
- 427 inside and a hydrophilic surface outside. This compact structure of starch affords higher stability

(Lopez, de Vries, & Marrink, 2012; Rachmawati, et al., 2013; Zhou, et al., 2016). Therefore, HPS
behaves as flexible random coils with stretches of helical segments in an aqueous solution at a high
temperature. With reducing temperature, the hydrogen bonds between starch chains and water were
broken, releasing the structured water, and inter-chain hydrogen bonding is enhanced, leading a
three-dimensional gel network (Fig. 6b).





435 Fig.6 Schematic representation of the sol-gel transition of HPMC (a), HPS (b), and HPMC/HPS (c)
436 pastes.

437

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For the mixture pastes, at a low temperature, HPMC has a lower viscosity than HPS and thus
HPMC forms a continuous phase encompassing discrete domains of higher-viscosity HPS gel. At the
edge of the two phases, the hydroxyl groups of HPMC chains can lose the structured water and form
hydrogen bonding with HPS chains. Upon heating, HPS may receive enough energy to form
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442 hydrogen bonding with water, resulting in the gradual disintegration of the gel network. Meanwhile, 443 the shells and water cages of HPMC chains are disrupted and even broken to expose hydroxyl groups 444 and hydrophobic clusters. At a high temperature, HPMC forms a gel network due to the inter-chain 445 hydrogen bonding and hydrophobic interaction and becomes a high-viscosity phase scattered in the 446 continuous phase of HPS coils (**Fig. 6c**). Therefore, HPS at a low temperature, or HPMC at a high 447 temperature, plays a dominant role in viscoelastic properties, gelation behavior, and phase structure 448 for HPMC/HPS pastes.

449 **4 Conclusion**

450 This work reveals how the phase structure of HPMC/HPS pastes was influenced by the gelation 451 behavior and rheological properties of individual biopolymers. And, we have established new links 452 between rheological parameters and phase structure for such systems.

The HPMC/HPS mixture showed a typical "sea-island" phase structure. The variation in this phase conformation was significantly influenced by the viscosity of individual biopolymer, blending ratio, and temperature. With increasing HPS ratio, the continuous phase of the blend changed from HPMC to HPS at an HPS ratio of 60%. We found that this phase structure change can be well correlated to the positive or negative deviation of η_0 from the log-additivity rule.

At a low temperature, HPMC, with low viscosity, was the continuous phase, and so was HPS at a high temperature. Moreover, HPS at a low temperature, or HPMC at a high temperature, plays the dominant role in controlling the viscoelastic properties and gelation behavior. With increasing temperature, the continuous phase changed from HPS to HPMC at a critical point of 45 °C. We introduced a modified Arrhenius-type equation with correction coefficient α (±1) to explore the effect of temperature on the continuous-phase change. This phase structure change can be well

464 correlated to rheological indicators such as the presence of a tan δ peak, a sudden change in η^* , and

465 the positive-negative change in α .

- 466 The results from this work led a schematic model to describe the temperature-induced
- 467 conformational change of HPMC/HPS pastes, which may provide an insight into the understanding
- 468 of the gelling mechanism for other temperature-sensitive multi-phasic systems.

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475 **Conflicts of interest**

476 There are no conflicts of interest to declare.

477 **Reference**

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Fig. 1 Viscosity as a function of time for HPMC/HPS pastes. The shear rate was fixed at 800 s⁻¹.

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Fig. 2 A) Flow curves for 50:50 (w/w) HPMC/HPS pastes of different total biopolymer concentration at 25 °C; B) Zero-shear viscosity as a function of total biopolymer concentration, with the molecular conformation of HPMC/HPS schematically shown (the HPMC/HPS ratio was 50:50 (w/w), red lines represent HPMC chains and grey lines HPS).





Fig.3 A) Viscosity *vs*. shear rate curves for HPMC/HPS pastes with different HPS ratio at 25 °C; B) Zero-shear viscosity η_0 , with the solid line indicating measured values and the dashed line predicted values; C) Flow behavior index *n*; and D) flow consistency index *K* for HPMC/HPS pastes with different HPS ratio. The total concentration of HPMC/HPS was 15 wt%.



Fig. 4 Three-interval thixotropic curves for HPMC/HPS pastes with different HPMC/HPS ratio at 25

°C. The total biopolymer concentration was 15 wt%.



Fig.5. A) Storage modulus (G'), loss modulus (G'') and loss tangent (tan δ) vs. temperature curves for the pure-HPMC paste, the 5:5 (w/w)

HPMC/HPS paste, and the HPS paste, respectively, all having a total biopolymer concentration of 15 wt%; B) Complex viscosity vs. temperature

curves for the HPMC paste, the 5:5 (w/w) HPMC/HPS paste, and the HPS paste, respectively, all having a total biopolymer concentration of 15

wt%; C) Light-microscopic images of the dyed 5:5 (w/w) HPMC/HPS paste of 3 wt% total biopolymer concentration at 25 °C, 45 °C, and 85 °C, Ji 3 w.

respectively.



Fig. 6 Schematic representation of the sol-gel transition of HPMC (a), HPS (b), and HPMC/HPS (c)

pastes.

Highlights:

- ✓ Hydroxypropyl methylcellulose (HPMC) and hydroxypropyl starch (HPS) mixture studied
- ✓ Strong rheological and gelation effects on the phase structure of HPMC/HPS shown
- ✓ HPMC/HPS blend ratio and temperature controls continuous/discrete phase change
- \checkmark Phase structure correlated to zero-shear viscosity and Arrhenius coefficient α
- \checkmark A schematic model to describe the temperature-induced conformational change

. confo.

- Declaration of Interest -

Hydroxypropyl methylcellulose and hydroxypropyl starch: Rheological and gelation effects on the phase structure of their mixed hydrocolloid system

Yanfei Wang^{1,2,3}, Long Yu³, Qingjie Sun^{1,2}, Fengwei Xie⁴

¹College of Food Science and Engineering, Qingdao Agricultural University, Qingdao, Shandong 266109, China

²Qingdao Special Food Research Institute, Qingdao, Shandong 266109, China

³College of Food Science and Engineering, South China University of Technology, Guangzhou, Guangdong 510640, China

⁴International Institute for Nanocomposites Manufacturing (IINM), WMG, University of Warwick, Coventry CV4 7AL, United Kingdom

The authors declare that there is no conflict of interest regarding the publication of this article.