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## COMMUNICATION

# One-pot synthesis of super-bright fluorescent nanogel contrast agents containing a dithiomaleimide fluorophore

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## Abstract

Fluorescent nanogels with radii ranging from 12-17 nm, high Quantum yield, concentration-independent emission, and fluorescent lifetimes of *ca.* 25 ns have been synthesised in a one-pot process. Solutions demonstrate a concentration independent emission between 0.1-10<sup>-3</sup> wt%, allowing for true quantitative imaging where dye emission is a measure of local nanoparticle concentration.

## Conceptual Insight (200 words max)

Fluorescent nanoparticles show a range of benefits over small molecule organic dyes for use in bioimaging and optical sensing; they display superior optical properties including high photostability and brightness, and can also possess greater biocompatibility than conventional organic dyes. Current limitations to the design and fabrication of nanoparticle contrast agents include synthetic complexity, and fluorophore self-quenching within the particle. Non-covalent dye encapsulation within a preformed nanoparticle provides a simple approach to fluorescent labelling, however, dye aggregation within the particles and leaching to the external environment leads to diminished optical properties. Attempts to improve performance by covalently attaching dyes is often synthetically complex, with prevention of intraparticle quenching still more demanding. In the present study we use a fluorophore that is easily incorporated into a methacrylate monomer to demonstrate a simple one-pot route to highly emissive fluorescent nanoparticles. Fluorophore

self-quenching is suppressed within these nanogel particles, leading them to outperform commonly used contrast agents.

## Introduction

Fluorescence spectroscopy and microscopy are widely used techniques for the detection and visualisation of compounds containing fluorophores. Fluorescently-labelled nanoparticles possess several features that make them more valuable for this purpose than classical fluorescent organic dyes.<sup>1</sup> Organic dyes may have low absorption coefficients, which reduces detection sensitivity, and are susceptible to photobleaching, thus limiting their use to short-term measurements. Furthermore, organic fluorophores often have short emission lifetimes, which can prohibit use in time-resolved measurements, while the toxicity of many organic fluorophores impedes their application to *in vitro* and *in vivo* analysis.<sup>2</sup> On the contrary, polymer nanoparticles with fluorescent functionality often contain multiple fluorophores leading to brighter emission, while the encapsulation of fluorophores in a macromolecular structure improves dye stability, and can impart biocompatibility to the fluorescently-labelled agent. Dyes can be incorporated either by non-covalent encapsulation, or by covalent attachment, with the latter strategy preferable as it is more efficient and stoichiometrically precise than physical absorption of the fluorophore, and also reduces leaching of the dye from the nanoparticle.<sup>1</sup> Covalent fluorophore labelling of polymer nanoparticles can be achieved in a variety of ways. Amphiphilic block copolymers can be labelled with fluorophores (using functional initiators and monomers, or by post-polymerisation functionalisation), with subsequent self-assembly providing fluorescently labelled polymer micelles.<sup>3-6</sup> Post-assembly labelling

can also be performed, using particles and dyes that are functionalised with complementary reactive groups.<sup>7-9</sup> Nanogel particles synthesised by emulsion or mini-emulsion polymerisation,<sup>10</sup> can be fluorescently-labelled by several different routes. In contrast to the encapsulation of dye molecules during<sup>11</sup> or after<sup>12, 13</sup> the (mini)emulsion polymerisation, the use of functional monomers allows for the covalent attachment of fluorophores,<sup>14, 15</sup> while post synthesis functionalisation has also been demonstrated.<sup>16, 17</sup> Fluorescent polymer nanoparticles can also be formed from fluorescent  $\pi$ -conjugated polymers. This can be achieved by performing the step-growth polymerisation in dispersion,<sup>18</sup> or by a post-polymerisation route.<sup>19, 20</sup> Current common strategies for the generation of polymeric fluorescence contrast agents for bioapplications suffer from a variety of complications during characterisation before transition to *in vitro* and *in vivo* studies.<sup>21</sup> Another difficulty in the generation of polymeric nanotheranostics is the ambiguity of labelling location, particle-to-particle variations in labelling density and the poorly understood effects of particle-dye, dye-dye and dye-solvent interactions on emission response in biologically relevant environments.<sup>21, 22</sup>

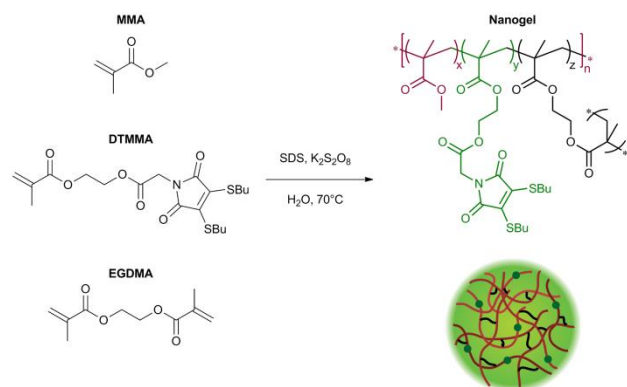
We have recently reported that the dithiomaleimide (DTM) group is a highly versatile fluorophore which can be simply incorporated into polymers and polymer nanoparticles using functional monomers and initiators for both controlled free radical and ring-opening polymerisation.<sup>23-26</sup> We demonstrated that nanoparticles containing the DTM fluorophore could be prepared by amphiphilic block copolymer synthesis using a DTM functional initiator, followed by block copolymer self-assembly in water to produce spherical micelles.<sup>25</sup> While the small molecule DTM exhibited all of the typical effects expected of a small dye molecule (quenching, short emission lifetime), when incorporated into the micellar state the DTM does not self-quench. The result was that the DTM-labelled particles showed a concentration independent molar emission and anisotropy, and a long fluorescence lifetime (20 ns). In addition, by using time-domain fluorescence-lifetime imaging (FLIM) *in vitro* it was possible to resolve assembled and dis-assembled micelles (*i.e.* unimers) due to a significantly different fluorescent lifetime for the two states.<sup>25</sup> In this work we instead chose a simpler synthetic route to DTM functional fluorescent nanoparticles, allowing synthesis in a one-step process. We utilised a recently developed DTM-functional methacrylate monomer,<sup>26</sup> in the fabrication of surfactant stabilised nanogel particles by radical cross-linking polymerisation in emulsion.

## Results and Discussion

### Nanoparticle synthesis

The fluorescently labelled DTM-functional particles (NG1-NG6) were formed by radical cross-linking emulsion polymerisation. The nanoparticle scaffold comprised of poly(methyl methacrylate) (PMMA), with crosslinking density (CLD) controlled by copolymerisation of ethyleneglycol dimethacrylate (EGDMA) as detailed in the Supporting Information. Covalent DTM dye attachment was achieved by copolymerisation of the recently reported monomer dithiomaleimide methacrylate (DTMMA),<sup>26</sup> with a

variable degree of functionalisation (*DoF*). Potassium persulfate was used as the radical initiator, and the reaction performed at 70 °C for 14 h. The reaction solvent was nanopure water (18.2 M $\Omega$ ·cm) with an MMA concentration of 10 g/L. A high concentration (2 g/L) of sodium dodecyl sulfate (SDS) was used as the surfactant to target nanogels with radii in the 10-25 nm size range, in accordance with previous work in our group.<sup>27, 28</sup> (Scheme 1).



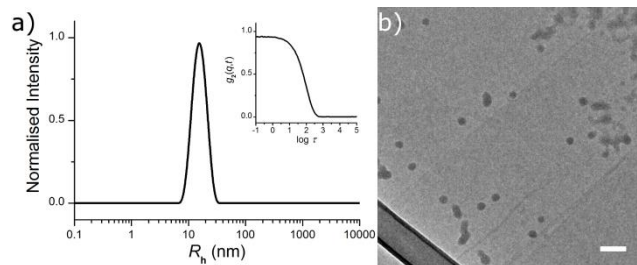
Scheme 1 Synthesis of DTM-labelled nanogels by radical cross-linking emulsion polymerisation.

*DoF* was varied over three orders of magnitude (for a constant *CLD* of 0.5 wt%) from 0.3 mol% to 0.0003 mol%, while *CLD* was also varied (for a constant *DoF* of 0.03 mol%) as 0.5 wt%, 3 wt% and 10 wt% so that the effect of both dye concentration and mobility in the nanogel core on fluorescence emission could be investigated (Table 1). After the removal of excess surfactant by exhaustive dialysis against water (18.2 M $\Omega$ ·cm), the resultant nanogel particle solutions were characterised by dynamic and static light scattering (DLS and SLS),<sup>29</sup> allowing determination of hydrodynamic radius ( $R_h$ ) and molecular weight ( $M_w$ ). For each sample, measurements were made with  $c = 5, 4, 3, 2 \& 1$  g/L, at  $\theta = 40^\circ$ - $150^\circ$  ( $10^\circ$  step), and the  $dn/dc$  was determined using a differential refractometer. In DLS measurements the autocorrelation functions showed a single mode of decay indicating the presence of a single well dispersed species in solution. From the resultant diffusion coefficient ( $D$ ), particle  $R_h$  was calculated (see SI, Fig. S2). Values of  $R_h$  were very similar for all samples, ranging from 12.0 nm to 16.5 nm (Table 1 and Fig. 1a). From these measurements, nanogel volume ( $V_{NG}$ ) could also be calculated, as the volume of a sphere with radius  $R_h$ . Particle  $M_w$  was measured by SLS, which allowed the average number of DTM units per nanogel ( $DoF_{NG}$ ) to be calculated from the molar degree of functionalisation (*DoF*) and  $M_w$  (see SI, Fig. S3). Knowledge of  $V_{NG}$  and  $DoF_{NG}$  therefore allowed the local DTM concentration within the nanogels ( $[DTM_{NG}]$ ) to be derived (see Table 1 for values and SI for further details).

Nanogels were also imaged by dry state transmission electron microscopy (TEM), using a graphene oxide (GO) substrate.<sup>30</sup> The particles showed a tendency to aggregate when dried to the GO surface (as opposed to their behaviour in solution), however, individual particles were observed to possess a spherical morphology (Fig. 1b).

Table 1. Composition and characterisation of DTM-labelled nanogels

	NG1	NG2	NG3	NG4	NG5	NG6
<i>DoF</i> (mol%)	0.3	0.03	0.03	0.03	0.003	0.0003
<i>CLD</i> (wt%)	0.5	0.5	3	10	0.5	0.5
<i>R<sub>h</sub></i> (nm)	12.0	16.5	14.0	12.5	15.6	15.3
<i>M<sub>w</sub></i> (Mg·mol <sup>-1</sup> )	2.4	11.6	4.8	4.0	9.7	7.8
<i>DoF<sub>NG</sub></i>	71	34	14	12	2.8	0.23
[DTM <sub>NG</sub> ] (mM)	16	3.0	2.0	2.3	0.30	0.025
$\tau_{\text{Av},1}$ (ns)	26.0	24.4	25.1	25.8	26.1	25.5

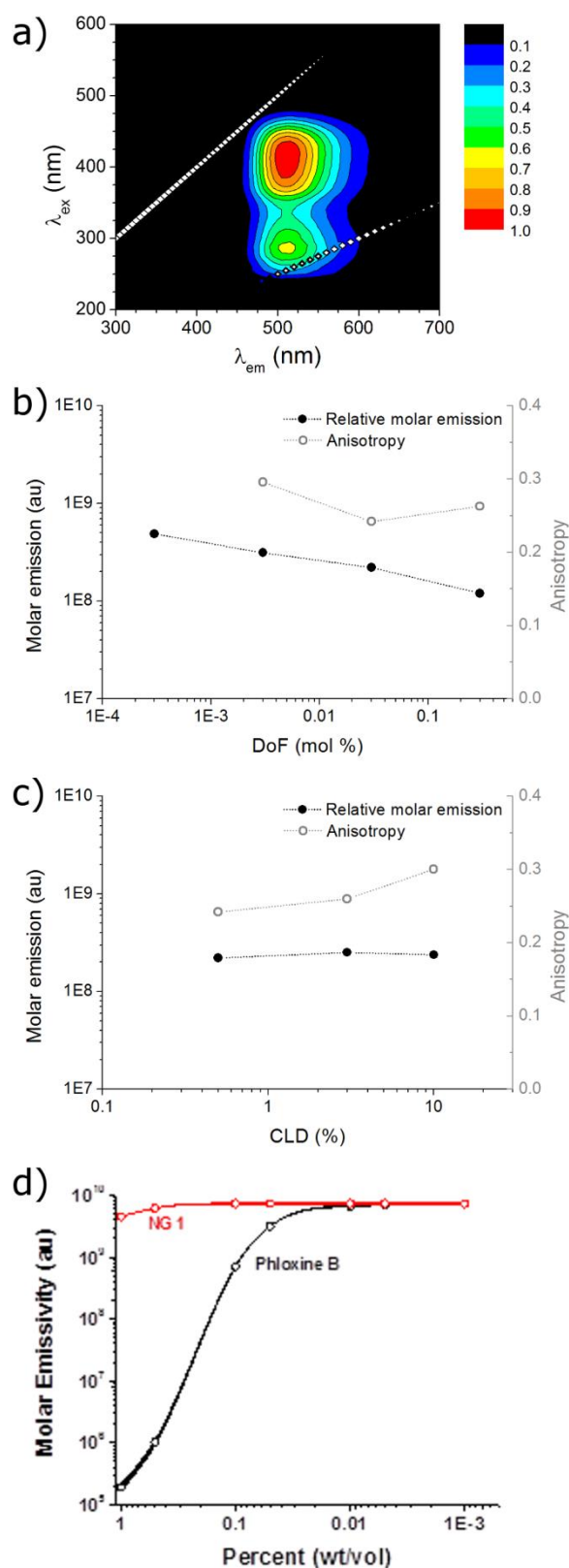


**Fig. 1** a) Size distribution obtained by DLS (detection angle of 90°) for a solution of **NG2** at 1 g/L, and the corresponding autocorrelation function (inset); b) **NG2** imaged by TEM on a GO support. The particles showed a tendency to aggregate upon drying (top right corner). Scale bar 100 nm

### Steady-state fluorescence

The excitation and emission of the nanogel solutions were characterised by recording a 2D fluorescence spectrum (for example **NG1** in Fig. 2a). Excitation maxima are observed at both *ca.* 275 nm and *ca.* 415 nm, which have the same corresponding emission maximum at *ca.* 510 nm. This is in accordance with the fluorescence profile of the small molecule DTM dyes and the DTM labelled polymers and micelles previously reported,<sup>24-26</sup> indicating that covalent attachment of the DTM unit into a nanogel particle has not significantly affected the wavelengths of excitation and emission.

The quantum yield (*Q*) for the aqueous solution of **NG1** (which has the highest DTM *DoF*) was calculated relative to the reference 5-(6)-carboxyfluorescein (5(6)-FAM) at 445 nm and found to be 54% (see SI for details). This value is 50-fold higher than the analogous small molecule dithiobutanemaleimide which has *Q* = 1.1% in methanol.<sup>25</sup> The significant difference in *Q* is attributed to the protection of the DTM fluorophore from solvent/collisional quenching afforded by the nanogel. The molar extinction coefficient ( $\epsilon$ ) for the aqueous solution of **NG1** was measured at the absorption maxima ( $\lambda_{\text{abs,max}}$ ) 405 nm. Using the particle molecular weight calculated by SLS, a value of  $\epsilon = 1.37 \times 10^5 \text{ M}^{-1} \cdot \text{cm}^{-1}$  was obtained with respect to nanogel concentration. Emission brightness can also be calculated as the product of quantum yield and extinction coefficient ( $Q \times \epsilon$ ), which is equal to  $4.51 \times 10^4 \text{ M}^{-1} \cdot \text{cm}^{-1}$  at the emission maxima ( $\lambda_{\text{em,max}}$ ) 420 nm. Values of  $\epsilon$  and brightness can also be calculated based on DTM dye concentration, using the known molarity of DTM-methacrylate used in the nanogel synthesis. In this case  $\epsilon = 1.89 \times 10^3 \text{ M}^{-1} \cdot \text{cm}^{-1}$  per dye unit at  $\lambda_{\text{em,max}} = 405 \text{ nm}$ , with the corresponding brightness of  $6.24 \times 10^2 \text{ M}^{-1} \cdot \text{cm}^{-1}$  per dye unit at 420 nm. While the other nanogel samples (**NG2-NG6**) displayed



**Fig. 2** a) 2D excitation-emission spectra with a 5 nm step for an aqueous solution of **NG1**. b) Molar emission and anisotropy for nanogels with *CLD* = 0.5 wt% with variable *DoF* (**NG1,2,5,6**), and c) *DoF* = 0.03 mol% with variable *CLD* (**NG2-4**). d) Molar emissivity vs concentration for aqueous solutions of **NG1** and Phloxine B

bright emission (*vide infra*), it was not possible to calculate quantum yield and brightness because absorption was too low at 445 nm to obtain accurate measurements as a result of the nanogels' lower *DoF*. Steady-state anisotropy (*r*) was measured for the solutions of nanogels (note that it was not possible to measure *r* for **NG6** (lowest *DoF*) due to overlap of the Raman scattering peak of water with the DTM emission at this low emission intensity). In all cases anisotropy was high ( $r > 0.2$ ) indicating that the DTM fluorophore had been successfully incorporated into a macromolecular structure (Fig. 2b&c). In comparison, we have previously shown that analogous small molecule DTM dyes have *r* *ca.* 0 in solution.<sup>25</sup>

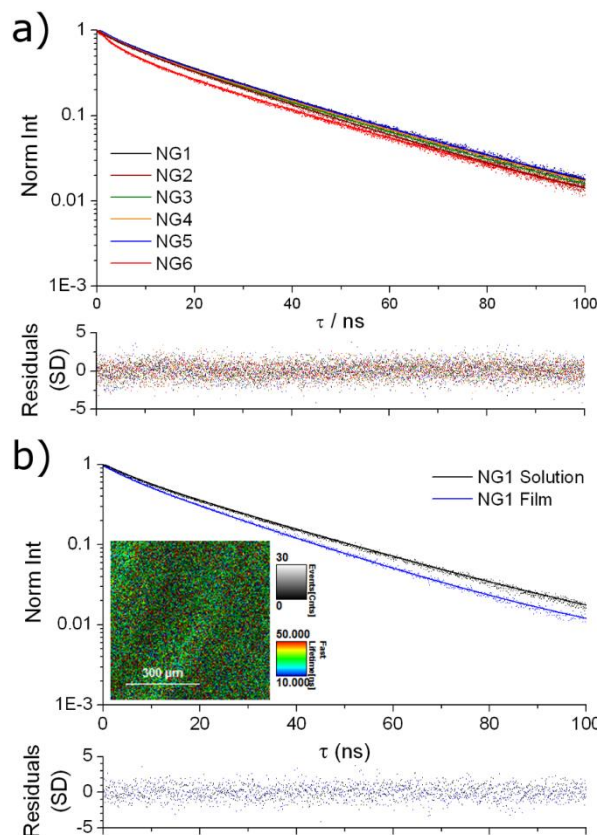
The steady-state emission spectra ( $\lambda_{\text{ex}} = 420 \text{ nm}$ ,  $\lambda_{\text{em}} = 510 \text{ nm}$ ) for nanogel samples with constant *CLD* = 0.5 wt% (**NG1,2,5,6**) showed that molar emission increased slightly by a factor of 4, as *DoF* (and also [DTM<sub>NG</sub>]) decreased by a factor of 1000 (Fig. 2b). The similarity in molar emission between **NG1** (*DoF* = 0.3 mol%) and **NG6** (*DoF* = 0.0003 mol%) shows that even at the highest *DoF* there is little intraparticle dye self-quenching, as light scattering measurements showed that **NG6** contained on average less than one fluorophore per particle (Table 1). Varying the *CLD* from 10 wt% to 0.5 wt% for nanogels with *DoF* = 0.03 mol% (**NG2-4**) was found to have almost no effect on molar emission (Fig. 2b), with values of  $2.37 \times 10^8 \pm 0.16 \times 10^8$ , suggesting that reducing the core mobility (by increasing the *CLD*) is not required to prevent dye self-quenching (Fig. 2c).

For the nanoparticles with the highest dye loading (**NG1**, *DoF* = 0.3 wt%) a study of molar emission as a function of concentration was performed. Between 0.1–10<sup>−3</sup> wt% molar emission remained constant, with only a slight decrease (*ca.* 10%) between 1–0.1 wt%. This feature would allow the DTM functional nanoparticles to be used as a quantitative imaging agent where dye emission is a true measure of local nanogel concentration. In contrast, the commonly used commercially available water soluble dye Phloxine B shows a  $\times 10^5$ -fold decrease in molar emission between 1–0.01 wt%, due to fluorophore aggregation and self-quenching (Fig. 2d). While polymeric nanoparticles which emit in the near infrared region show many benefits (reduced photon scattering and background cellular autofluorescence) over visible region emitters for *in vivo* imaging,<sup>31,32</sup> the similarity of the DTM emission to FITC<sup>33</sup> ( $\lambda_{\text{ex}}$  *ca.* 520 nm) and the high local dye concentrations and quantum yield achievable with these DTM nanogels would not preclude their potential application *in vivo*.

#### Time-resolved fluorescence spectroscopy

Solution state fluorescence lifetime was measured for the nanogel samples, using time-correlated single photon counting (TCSPC). All samples showed very similar emission decay (Fig. 3a), with an intensity averaged fluorescence lifetime ( $\tau_{\text{AV}}$ ) of *ca.* 25 ns obtained by fitting to a multi-exponential decay (details in SI). For comparison, we have previously reported that the solution state average fluorescence lifetime of small molecule DTM dyes is *ca.* 1–2 ns, whereas diblock copolymer micelles containing the DTM fluorophore at the core-corona interface had  $\tau_{\text{AV}}$  = 20 ns when assembled into spherical micelles.<sup>25</sup> These values of  $\tau_{\text{AV}}$  for the DTM functional nanogels are significantly above that of cellular autofluorescence which is caused

by endogenous fluorescent biomolecules such as keratin, collagen, flavins, metal-free porphyrins and NAD(P)H coenzymes, and is typically on the order of 2–3 ns.<sup>34</sup> This means that the DTM containing nanogels synthesised here would be ideal as *in vitro* fluorescence-lifetime imaging microscopy (FLIM) contrast agents, as we have previously demonstrated for DTM functional polymer micelles.<sup>25</sup>



**Fig. 3** Fluorescence lifetime decay spectra (points) with fitting (line) and residuals (bottom) for a) **NG1-6** in solution; b) **NG1** in solution and dried to a film, with FLIM image of the film (inset).

The fluorescence lifetime of thin films prepared by drying the nanogel solutions to a glass slide (investigated using FLIM) provided further information about the nature of the dye incorporation into the nanogel particles. In all cases near identical fluorescence decay was observed when comparing nanogel solutions and films. For example, the comparison for **NG1** is shown in Fig. 3b, with fitting of the decays indicating  $\tau_{\text{AV}}$  = 23.4 ns for the nanogel films, and  $\tau_{\text{AV}}$  = 26.0 ns for the nanogel solutions. The similarity in the fluorescence intensity decay between nanogel particles either in solution or dehydrated, indicates that encapsulation within a nanogel provides excellent protection to the DTM fluorophore's excited state from collisional quenching with the surrounding solvent. The result is a drastically increased fluorescence lifetime when compared to solutions of the small molecule DTM dyes. The data presented here demonstrates that covalent attachment of the DTM fluorophore inside a PMMA nanogel virtually eliminates both self-quenching caused by dye aggregation, and collisional quenching from the surrounding solvent. Given the quasi-



encapsulated nature of the nanogels and the macromolecular architecture, it is reasonable to conclude that the resistance to quenching in these systems is related to both long range superstructural isolation and shorter range volume exclusion due to polymeric bonding. When considering the high concentrations of fluorophore in the particle cores (up to 16 mM) the lack of self-quenching is quite remarkable, as most dyes or dye/polymer blends would be 'off-switched' in the solid state at these concentrations. This highlights the advantage of a covalent dye attachment, as opposed to non-covalent encapsulation, which is facilitated by the ease with which DTM-functional monomers can be synthesised and polymerised.

## Conclusions

A simple one-pot procedure has been used to synthesise aqueous solutions of nanogel particles labelled with a DTM fluorophore. By simply varying the ratio of reagents, the loading of DTM as well as the cross-linking density can be easily tailored, with particle radii in the range 12–17 nm. Covalent attachment of the DTM unit to the nanogel particles provides protection to the fluorophore from solvent-quenching and self-quenching, leading to very bright emission and significant fluorescence lifetimes. Furthermore, the fluorescent nanogels display a concentration independent molar emission profile up to very high concentration (0.1 wt%). When used as contrast agents these fluorescent nanogels could therefore be applied for true quantitative imaging, and for fluorescence-lifetime imaging microscopy. We are excited by the potential to expand this simple synthetic approach to prepare fluorescent particles with further functionality, for example response to an external stimulus (e.g. temperature, pH), using the vast library of functional and responsive monomers that have been previously utilised in emulsion polymerisation.

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## Notes and references

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Electronic Supplementary Information (ESI) available: Full details of experimental protocol, light scattering analysis, quantum yield and brightness calculations, and TCSPC analysis. See DOI: 10.1039/c000000x/

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## Graphical Abstract

