

## **Original citation:**

Michniewski, Slawomir, Redgwell, Tamsin, Scanlan, David J. and Millard, Andrew D.. (2017) Draft genome sequence of Bacteriophage vB\_Eco\_swan01. Genome Announcements, 5 (28). pp. 1-17. e00501.

#### **Permanent WRAP URL:**

http://wrap.warwick.ac.uk/90862

# **Copyright and reuse:**

The Warwick Research Archive Portal (WRAP) makes this work of researchers of the University of Warwick available open access under the following conditions.

This article is made available under the Creative Commons Attribution 4.0 International license (CC BY 4.0) and may be reused according to the conditions of the license. For more details see: <a href="http://creativecommons.org/licenses/by/4.0/">http://creativecommons.org/licenses/by/4.0/</a>

### A note on versions:

The version presented in WRAP is the published version, or, version of record, and may be cited as it appears here.

For more information, please contact the WRAP Team at: wrap@warwick.ac.uk







# Draft Genome Sequence of Bacteriophage vB\_Eco\_swan01

Slawomir Michniewski, a Tamsin Redgwell, a David J. Scanlan, a Andrew D. Millardb

School of Life Sciences, University of Warwick, Coventry, United Kingdom<sup>a</sup>; Microbiology and Infection Unit, Warwick Medical School, University of Warwick, Coventry, United Kingdom<sup>b</sup>

**ABSTRACT** Bacteriophage vB\_Eco\_swan01 was isolated from an ornamental pool using *Escherichia coli* MG1655 as the host. Bacteriophage vB\_Eco\_swan01 has limited similarity with other known phages at the nucleotide level and likely represents a new bacteriophage species within the *Tunavirinae*.

ere, we report the genome sequence of bacteriophage vB\_Eco\_swan01, which is capable of infecting *Escherichia coli* MG1655. This phage was isolated using an enrichment procedure involving a single-agar-layer plague assay (1). Briefly, an 80-mL filtered water sample (i.e., filtered through 0.45- $\mu$ m pores) was mixed with 5 mL of E. coli MG1655 cells, CaCl<sub>2</sub> to a final concentration of 1 mM, and incubated at room temperature for 10 min. Subsequently, 80 mL of molten 1.2% (wt/vol) LB agar (previously adjusted to 50°C) was added to the sample. The mixture was then poured into disposable petri dishes and incubated overnight at 37°C. A single plaque of ~3 mm in diameter was picked and transferred into SM buffer, followed by vortexing to release phages from the agar plug and storage at 4°C. Bacteriophage genomic DNA was extracted from a lysed culture using a phenol:chloroform method (2). Genomic DNA was prepared using the NexteraXT DNA sample preparation kit (Illumina) with modified PCR conditions consisting of 14 rounds of 97°C for 10 s, 55°C for 30 s, and 65°C for 1 min. Sequencing was performed on the MiSeq platform using V2 (2 imes 250-bp) chemistry. The resulting FASTQ files were trimmed with Sickle using default parameters (3), prior to being assembled with SPAdes version 3.7 using the "-careful" option (4). The genome was sequenced to an average depth of 344×. The resulting single contig was annotated with Prokka version 1.11 using a custom database constructed from all complete viral genomes within the European Nucleotide Archive (5) and then further annotated using hmmscan using the prokaryotic Viral Orthologous Groups (pVOG) collection of hmm profiles (6), using a cutoff value of <1E-15. Bacteriophage vB\_ Eco\_swan01 has a double-stranded DNA genome of 50.865 kb and a G+C content of 44.96%. A total of 83 open reading frames were predicted, with no tRNAs found. At the nucleotide level, vB\_Eco\_swan01 has an average nucleotide identity (ANI) of 72% with bacteriophage pSF1, an unclassified member of the family Siphoviridae, but has no significant similarity with other bacteriophages. For the majority of genes, it was not possible to predict a function for the encoded proteins. Of the 83 predicted proteins, 75 could be annotated as being part of a known pVOG. Phylogenetic analysis using genes encoding for the tape measure protein (00017) and large terminase subunit (00031) revealed that vB\_Eco\_swan01 is related to phages in the subfamily *Tunavirinae*. However, given the low ANI compared to all other phages, vB\_Eco\_swan01 likely represents a new species based on current definitions of phage classification (7). This genome adds further diversity to those bacteriophages capable of infecting E. coli.

**Accession number(s).** The draft genome sequence of bacteriophage vB\_Eco\_swan01 has been deposited in DDBJ/ENA/GenBank under the accession number LT841304.

**Received** 23 April 2017 **Accepted** 17 May 2017 **Published** 13 July 2017

Citation Michniewski S, Redgwell T, Scanlan DJ, Millard AD. 2017. Draft genome sequence of bacteriophage vB\_Eco\_swan01. Genome Announc 5:e00501-17. https://doi.org/10.1128/genomeA.00501-17.

**Copyright** © 2017 Michniewski et al. This is an open-access article distributed under the terms of the Creative Commons Attribution 4.0 International license.

Address correspondence to Andrew D. Millard, a.d.millard@warwick.ac.uk.

#### **ACKNOWLEDGMENTS**

Funding for sequencing was provided by the Medical Research Council (MRC) CLIMB grant (MR/L015080/1). T.R. and S.M. were in receipt of PhD studentships funded by the Natural Environment Research Council (NERC) CENTA DTP.

#### **REFERENCES**

- Grabow WO, Coubrough P. 1986. Practical direct plaque assay for coliphages in 100-ml samples of drinking water. Appl Environ Microbiol 52:430-433.
- 2. Rihtman B, Meaden S, Clokie MRJ, Koskella B, Millard AD. 2016. Assessing Illumina technology for the high-throughput sequencing of bacterio-phage genomes. PeerJ 4:e2055. https://doi.org/10.7717/peerj.2055.
- Joshi NA, Fass JN. 2011. Sickle: a sliding-window, adaptive, quality-based trimming tool for FastQ files (version 1.33). https://github.com/najoshi/ sickle.
- 4. Bankevich A, Nurk S, Antipov D, Gurevich AA, Dvorkin M, Kulikov AS, Lesin VM, Nikolenko SI, Pham S, Prjibelski AD, Pyshkin AV, Sirotkin AV, Vyahhi N,
- Tesler G, Alekseyev MA, Pevzner PA. 2012. SPAdes: a new genome assembly algorithm and its applications to single-cell sequencing. J Comput Biol 19:455–477. https://doi.org/10.1089/cmb.2012.0021.
- Seemann T. 2014. Prokka: rapid prokaryotic genome annotation. Bioinformatics 30:2068–2069. https://doi.org/10.1093/bioinformatics/btu153.
- Grazziotin AL, Koonin EV, Kristensen DM. 2017. Prokaryotic virus orthologous groups (pVOGs): a resource for comparative genomics and protein family annotation. Nucleic Acids Res 45:D491–D498. https://doi.org/10.1093/nar/gkw975.
- Adriaenssens EM, Brister JR. 2017. How to name and classify your phage: an informal guide. Viruses 9:70. https://doi.org/10.3390/v9040070.