

Original citation:

Arasaradnam, Ramesh, Wicaksono, Alfian, O'Brien, Harrison, Kocher, Hemant M., Covington, James A. and Crnogorac-Jurcevic, Tatjana. (2017) Non-invasive diagnosis of pancreatic cancer through detection of volatile organic compounds in urine. Gastroenterology.

Permanent WRAP URL:

http://wrap.warwick.ac.uk/95165

Copyright and reuse:

The Warwick Research Archive Portal (WRAP) makes this work by researchers of the University of Warwick available open access under the following conditions. Copyright © and all moral rights to the version of the paper presented here belong to the individual author(s) and/or other copyright owners. To the extent reasonable and practicable the material made available in WRAP has been checked for eligibility before being made available.

Copies of full items can be used for personal research or study, educational, or not-for-profit purposes without prior permission or charge. Provided that the authors, title and full bibliographic details are credited, a hyperlink and/or URL is given for the original metadata page and the content is not changed in any way.

Publisher's statement:

© 2017, Elsevier. Licensed under the Creative Commons Attribution-NonCommercial-NoDerivatives 4.0 International http://creativecommons.org/licenses/by-nc-nd/4.0/

A note on versions:

The version presented here may differ from the published version or, version of record, if you wish to cite this item you are advised to consult the publisher's version. Please see the 'permanent WRAP url' above for details on accessing the published version and note that access may require a subscription.

For more information, please contact the WRAP Team at: wrap@warwick.ac.uk

1 Gastroenterology In Motion 2 3 Title: Non-invasive Diagnosis of Pancreatic Cancer Through Detection of 4 **Volatile Organic Compounds in Urine** 5 Authors: Ramesh Arasaradnam¹, Alfian Wicaksono⁴, Harrison O'Brien², Hemant M 6 Kocher³, James A Covington^{4*}, Tatjana Crnogorac-Jurcevic^{2*} 7 8 9 *Shared senior authorship 10 11 Correspondence to: 12 Professor Ramesh Arasaradnam 13 Department of Gastroenterology, University Hospital Coventry & Warwick, Applied Biological Sciences, University of Coventry 14 15 Division of Surgery | UHCW NHS Trust | Clifford Bridge Road | Coventry CV2 16 2DX, UK 17 18 ¹Department of Gastroenterology, University Hospital Coventry & Warwick, 19 Applied Biological Sciences, University of Coventry 20 ² Centre for Molecular Oncology, Barts Cancer Institute, Queen Mary University of 21 London 22 ³Centre for Tumour Biology, Barts Cancer Institute, Queen Mary University of 23 London 24 ⁴School of Engineering and Medical School, University of Warwick 25 26 Funding source: Barts Pancreas Tissue Bank is supported by PCRF. 27 28 Conflict of interest: None 29 30 Acknowledgement: Barts Pancreas Tissue Bank personnel involved in this work 31 include Vickna Balarajah, Thomas Dowe and Amina Saad. Please see 32 https://www.bartspancreastissuebank.org.uk/

33

Authors contribution: RA & TC-J designed the study, RA drafted the manuscript, AW & JC undertook analysis and RA, TC-J and JC provided interpretation, AW & JC undertook statistical analysis, JC provided technical expertise, HB & HK recruited patients and sample collection, RA, AW, HK, JC & TC-J reviewed the manuscript for intellectual content.

Word count: 998

With its incidence approaching mortality, and with over 300,000 new cases diagnosed worldwide in 2013, pancreatic ductal adenocarcinoma (PDAC) is currently the fourth leading cause of cancer-related death and predicted to become the second by 2030 (1). More than 80% of PDAC patients are diagnosed late, with locally invasive and/or metastatic disease, resulting in negligible 5-year survival. Timely detection of PDAC is hampered by several factors: lack of specific clinical symptoms in the early stages, insufficient sensitivity of current imaging modalities and a lack of accurate, clinically utilisable biomarkers (2). Thus the quest for a simple, inexpensive and non-invasive test to detect PDAC early - whilst it is still amenable to surgical resection, continues. Detection of volatile organic compounds (VOCs) has come to the fore offering a novel approach for the detection of disease. It uses the odours that emanate from urine, breath and faeces and is akin to canine 'sniffing'. These compounds are metabolic products and/or consequence of bacterial dysbiosis produced by the disease state (3, 4). We thus postulate that either altered cellular physiology or even alteration in the microbial milieu in patients with PDAC will alter the individual's metabolome profile, such that the resultant VOC patterns that are emitted provide a characteristic signature that can be detected.

Description of Technology

For this study, we used a commercial gas analysis instrument (Owlstone Lonestar), based on ion mobility spectrometry (IMS) to analyse the VOCs emanating from patients' urine samples. Patients' information is summarised in **Table 1**. Midstream urine samples were collected and frozen at -80°C within 4 hours of collection for long-term storage. Prior to analysis, each sample was defrosted and heated to 40°C for 10 minutes. The air above the sample (known as the headspace) was then passed into the Lonestar unit and the ion mobility measured. Each sample typically takes five minutes to analyse (60 seconds per sample to scan, with five repeats). Once all the samples were analysed, an existing data processing pipeline with four different classifiers was applied (Sparse Logistic Regression, Random Forest, Gaussian Process Classifier and Support Vector Machine). Age was not deemed to be a confounder based on our previous published work – supplementary material. PDAC urine samples were detected with a sensitivity (SN) of 0.91 (CI: 0.83-0.96) and specificity (SP) of 0.83 (CI: 0.73-0.90), with an area under the curve (AUC) of 0.92 (CI:0.88-0.96), using a support vector machine algorithm – **Figure 1c.** The results were

validated by random data splitting into the training and test set: the machine algorithm system was trained using the first 100 samples and the results of the remaining 62 test samples were then predicted. This achieved similar results with AUC of 0.92 (CI: 0.85-0.98), and SN and SP of 0.90 (CI: 0.74-0.98) and 0.81 (CI: 0.63-0.93), respectively. We also compared early stage disease (I/II) with healthy individuals as well as early stage I/II with late stage disease (III/IV) using the same analysis pipeline. These revealed a sensitivity (SN) of 0.91 (CI: 0.78-0.97) and specificity (SP) of 0.78 (CI: 0.69-0.86), with an area under the curve (AUC) of 0.89 (CI: 0.83-0.94), and SN of 0.82 (CI: 0.67-0.92), SP of 0.89 (CI: 0.75-0.97) with AUC of 0.92 (CI: 0.86-0.97), respectively, again using a support vector machine algorithm (**Figures 1d** and **e**).

8788

89

90

91

92

93

94

95

96

97

98

99

100

101

102

103

104

105

106

107

108

109

76

77

78

79

80

81

82

83

84

85

86

Video Description

The Lonestar is a special type of Ion Mobility Spectrometer (IMS), based on a method called FAIMS, (Field Asymmetric Ion Mobility Spectroscopy). Developed and used extensively by the security services, it has recently been employed for the detection of VOCs in medicine. It has the advantage of being more reproducible and far more sensitive than electronic noses, whilst at the same time, it does not have the limitations of mass spectrometry, which requires specialised and expensive infrastructure. FAIMS is based on measuring the way that ionised molecules move in very high electric fields and due to the differences in this movement, the instrument is able to separate complex mixtures of chemicals that are found in biological samples. It practice, this is achieved by passing ionised molecules between two plates, where an asynchronous electric field is applied. This attracts, repels or has no effect of these ions. If an ion touches one of the plates it loses its charge, and thus only ions that exit the plates without touching either side are detected. To the asynchronous electric field, an additional fixed "compensation voltage" is added that counteracts the movement. Thus, by scanning through a range of different compensation voltages we are able to measure a large range of different mobilities. To improve separation, the magnitude of the electric field is raised through a series of values to create a 3D map of a urine sample. These are then analysed using our in house data processing pipeline, previously described when analysing urine samples with different disease groups (5). In brief, first a data compression wavelet and threshold is applied to reduce the size of the dataset (which in our case is more than 50,000 data points per

sample). Once completed, important features are sought within the compressed dataset, using a Wilcoxon rank-sum test, with a 10-fold cross-validation step. Such identified features can then be used with different classification algorithms to calculate clinically relevant values e.g. AUC, SN and SP of unknown samples.

Take Home Message

We have already showed that urine is a useful bio-fluid for detection of early stage, resectable pancreatic cancer (6,7). In this proof of concept study, we demonstrate for the first time, utility of urine specimens to discriminate healthy individuals from PDAC patients also through detection of volatile organic compounds. Moreover, we have also shown the ability to separate healthy from early stage and early stage vs advanced disease. This was achieved using only 5 ml of urine sample and applying novel IMS technology, which offers a rapid, and more cost effective analysis compared with other gas analysis technologies. The obtained results demonstrate that urine analysis using IMS platform shows promise as an additional non-invasive approach for identification of patients with pancreatic cancer.

Table 1. Demographic information

	Healthy controls $(n = 81)$	Pancreatic Cancer (n=81)
Age (mean \pm SD)	51.4 ± 10.6	64.3 ± 23.7
Sex (male) %	30.9	53.1
Stage*		
I	n/a	4
IIA	n/a	5
IIB	n/a	35
III	n/a	24
IV	n/a	12

130 *One case could not be assessed

143 **References:**

- 144 1. Rahib L, et. al. Cancer Res. 2014;74(11):2913-21.
- 145 2. Kaur S, et. al. Biomark Med. 2012;6(5):597-612.
- 146 3. Arasaradnam RP, et. al. Aliment Pharmacol Ther. 2014;39(8):780-9.
- 4. Arasaradnam RP, et. al. Med Hypotheses. 2009;73(5):753-6.
- 148 5. Covington JA, et. al. Analyst. 2015;140(20):6775-81.
- 149 6. Debernardi S, et. al. Am J Cancer Res. 2015;5(11):3455-66.
- 150 7. Radon TP, et. al. Clin Cancer Res. 2015;21(15):3512-21.

151

152

Figure 1Legend

Figure 1a) Representative image of FAIMS output showing the plume of ion dispersion from a single urine sample of a healthy control (A.U. is arbitrary units). The mobility of ions determines the shape of the plume (5). **Figure 1b)** shows similar plume of ion dispersion from a single urine sample of a patient with PDAC. Note the difference in the shape of the plume as reflected by the different ion mobility shown in red. **Figure 1c)** Receiver Operating Characteristic (ROC) curve showing performance of VOCs in differentiating healthy individuals from pancreatic cancer samples with AUC of 0.92 (CI: 0.88-0.96); SN=0.91 (CI: 0.83-0.96); SP=0.83 (CI: 0.73-0.9); **Figure 1d)** Healthy individuals from early stage I/II (AUC =0.89; CI: 0.83-0.94); SN=0.91 (CI: 0.78-0.97); SP=0.78 (CI: 0.69-0.86). **Figure 1e)** shows early stage PDAC(I/II) could also be successfully separated from advanced stage (IIII/IV) pancreatic cancer with AUC 0.92 (CI: 0.86-0.97); SN=0.82 (CI: 0.67-0.92); SP=0.89 (CI: 0.75-0.97).